

Supporting Information for

# DEVELOPING TARGETED HYBRID IMAGING PROBES BY CHELATOR SCAFFOLDING

Dominik Summer,<sup>†</sup> Leo Grossrubatscher,<sup>†</sup> Milos Petrik,<sup>‡</sup> Tereza Michalcikova,<sup>‡</sup> Zbynek Novy,<sup>‡</sup>  
Christine Rangger,<sup>†</sup> Maximilian Klingler,<sup>†</sup> Hubertus Haas,<sup>§</sup> Piriya Kaeopookum,<sup>†||</sup> Elisabeth von  
Guggenberg,<sup>†</sup> Roland Haubner,<sup>†</sup> and Clemens Decristoforo<sup>\*†</sup>

<sup>†</sup>*Department of Nuclear Medicine, Medical University Innsbruck, Anichstrasse 35, A-6020 Innsbruck,  
Austria*

<sup>‡</sup>*Institute of Molecular and Translational Medicine, Faculty of Medicine and Dentistry, Palacky  
University Olomouc, Hnevotinska 5, 779 00, Olomouc, Czech Republic*

<sup>§</sup>*Division of Molecular Biology / Biocenter, Medical University Innsbruck, Innrain 80-82, A-6020  
Innsbruck, Austria*

<sup>||</sup>*Ministry of Science and Technology (MOST), Thailand Institute of Nuclear Technology (TINT),  
Nakhonnayok 26120, Thailand*

**\* Corresponding author:**

*Clemens Decristoforo; Department of Nuclear Medicine, Medical University Innsbruck, Anichstrasse  
35, A-6020 Innsbruck, Austria; email: [Clemens.Decristoforo@i-med.ac.at](mailto:Clemens.Decristoforo@i-med.ac.at); Te:+4351250480951;  
Fax:+435125046780951*

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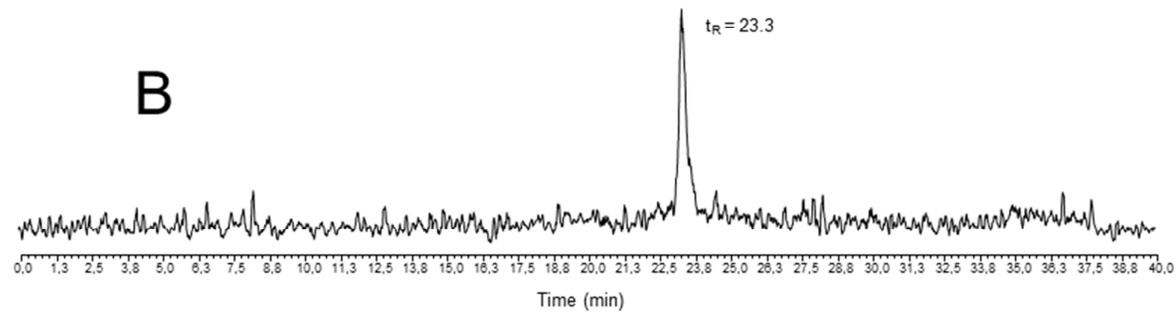
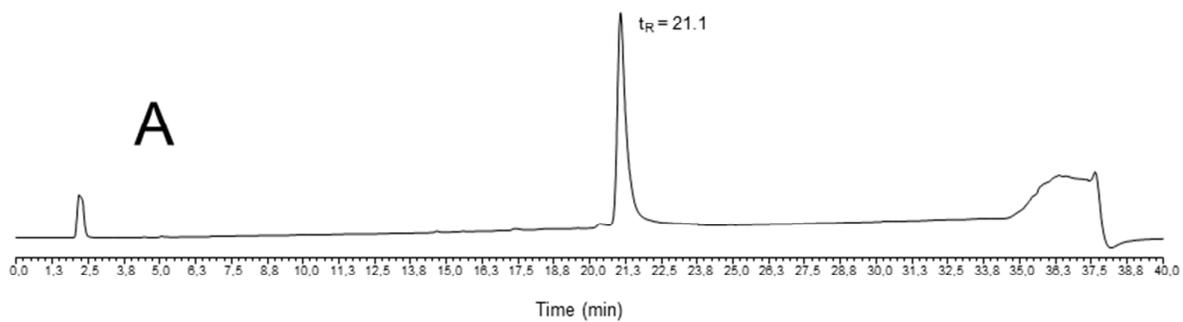
**Radioiodination general information.** The cyclic pentapeptide c(RGDyV) was synthesized according to Haubner et al.<sup>1</sup> and [Leu<sup>15</sup>]-Gastrin I was purchased from Bachem (Bubendorf, Switzerland). Iodine-125 (5 mCi) was obtained from Perkin Elmer (Waltham, MA, USA) as Na<sup>125</sup>I in 0.01 mM NaOH solution (pH 8-11). Iodine-125 labelling was performed at ambient temperature and labelled peptides were purified by analytical radio-RP-HPLC and fractions were collected manually. RP-HPLC analysis was carried out on an UltiMate 3000 RS UHPLC pump, Ultimate 3000 column compartment (25 °C oven temperature), UltiMate 3000 Variable Wavelength Detector (Thermo Fisher Scientific, Vienna, Austria, UV detection at 220 nm), GabiStar radio detector (Raytest; Straubenhardt, Germany) and a Jupiter 5 µm C<sub>18</sub> 300 Å 150 × 4.6 mm (Phenomenex Ltd. Aschaffenburg, Germany) column with following acetonitrile (ACN)/H<sub>2</sub>O/ 0.1 % trifluoroacetic acid (TFA) multistep gradients was used: Gradient A: flow rate of 1.0 mL/min; 0.0–2.0 min 20% ACN, 2.0–32.0 min 20–50 % ACN, 32.0–33.0 min 50–80 % ACN, 33.0–35.0 min 80 % ACN, 35.0–40.0 min 20 % ACN. Gradient B: flow rate of 1.5 mL/min; 0.0–2.0 min 0% ACN, 2.0–15.0 min 0–50 % ACN, 15.0–16.0 min 50–100 % ACN, 16.0–18.0 min 100 % ACN, 18.0–20.0 min 0 % ACN.

**Radiosynthesis of [<sup>125</sup>I]-[Leu<sup>15</sup>]-Gastrin I via Chloramine T method.** An aliquot of 10 µL of peptide stock solution (0.48 mM in H<sub>2</sub>O) was mixed with 25 µL 0.5 M phosphate buffer, 5 µL Na<sup>125</sup>I (10 MBq) solution and after adding 20 µL of freshly water-dissolved *N*-Chloro-*p*-toluenesulfonamide sodium salt in a concentration of 7.1 mM the resulting mixture was reacted for 1 min. Thereafter 25 µL 10 % (m/v) bovine serum albumin (BSA) dissolved in 0.05 M phosphate buffer was added and after 2 minutes reaction time followed by addition of 10 µL potassium iodide solution (10 % in H<sub>2</sub>O, m/v) [<sup>125</sup>I]-[Leu<sup>15</sup>]-Gastrin I was purified using analytical RP-HPLC (gradient A).

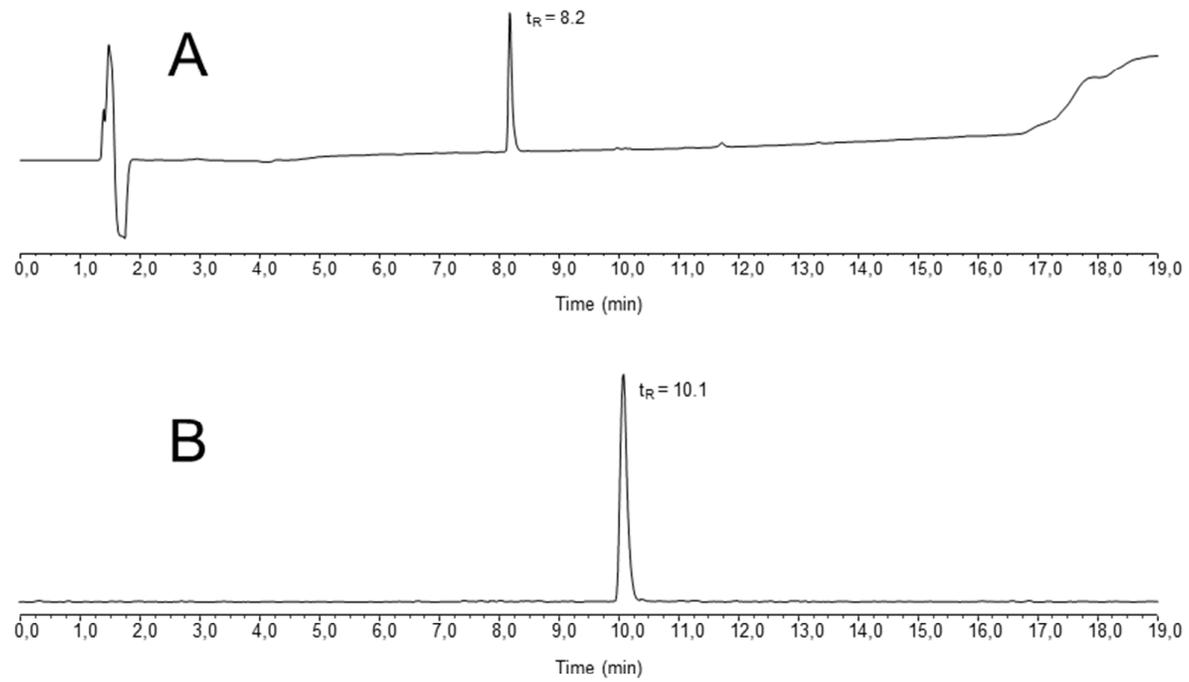
**Radiosynthesis of [<sup>125</sup>I]-c(RGDyV) via Iodogen method.** Pierce™ Iodination Reagent (ThermoFisher Scientific, Vienna, Austria) was dissolved in dichloromethane to a final concentration of 2.3 mM and after transferring 150 µL to an Eppendorf tube the organic solvent was removed under argon stream. Hereafter 150 µL 0.5 M phosphate buffer (pH 7.4), 10 µL of peptide stock solution (1.7 mM in H<sub>2</sub>O) and 5 µL Na<sup>125</sup>I (10 MBq) solution was added to the 1,3,4,6-tetrachloro-3α,6α-diphenylglycoluril coated tube and the mixture was maintained under gentle shaking. After 10 min the reaction solution was immediately transferred to a fresh Eppendorf tube and [<sup>125</sup>I]-c(RGDyV) was purified via analytical RP-HPLC (gradient B).

### **Supporting Reference**

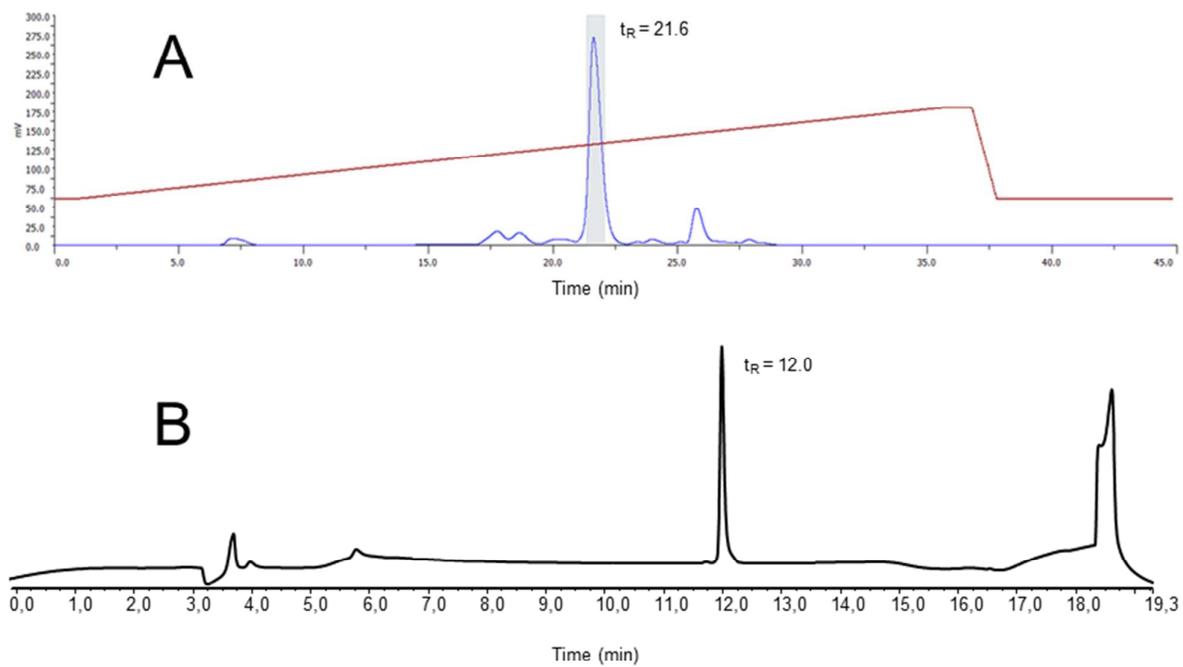
(1) Haubner, R., Wester, H. J., Reuning, U., Senekowitsch-Schmidtke, R., Diefenbach, B., Kessler, H., Stocklin, G., and Schwaiger, M. (1999) Radiolabeled α<sub>v</sub>β<sub>3</sub> integrin antagonists: A new class of tracers for tumor targeting. *J. Nucl. Med.* 40, 1061–1071.



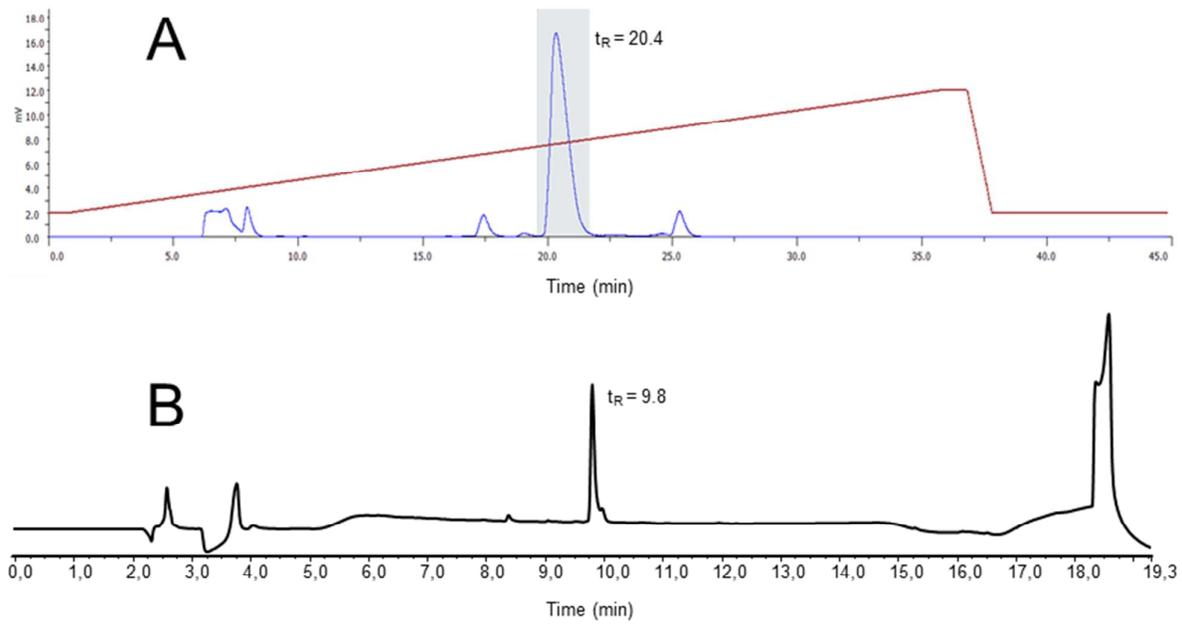
**Figure S1:** analytical RP-HPLC chromatogram of [Leu<sup>15</sup>]-Gastrin I (**A**) and [<sup>125</sup>I]-[Leu<sup>15</sup>]-Gastrin I (**B**)



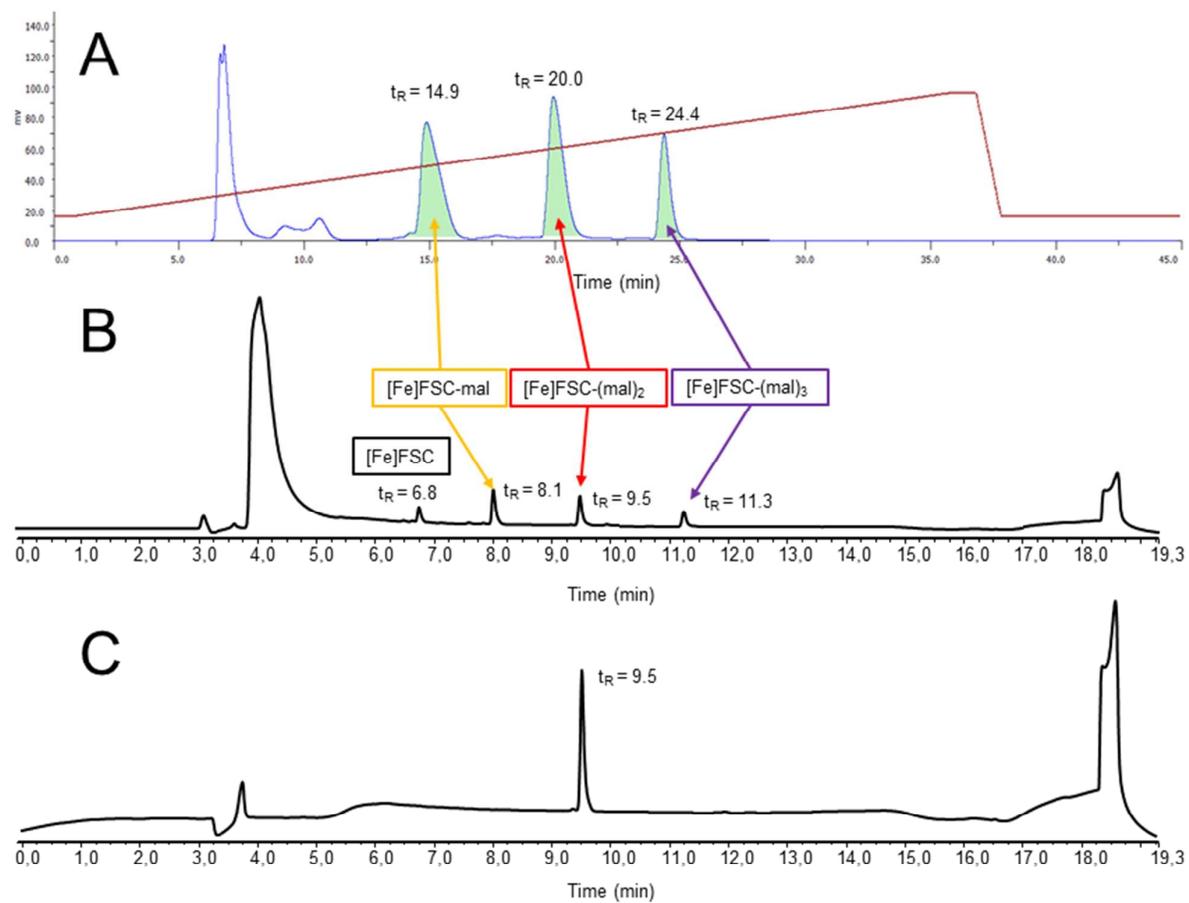
**Figure S2:** analytical RP-HPLC chromatogram of c(RGDyV) (A) and  $[^{125}\text{I}]\text{-c(RGDyV)}$  (B)



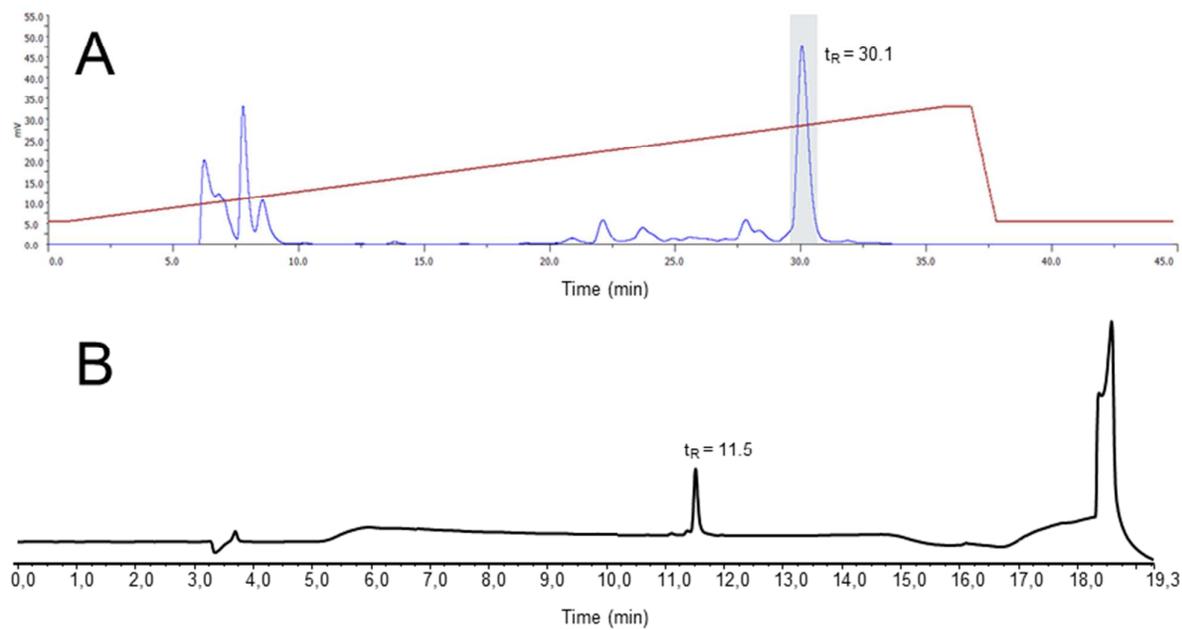
**Figure S3:** preparative RP-HPLC (A) and analytical RP-HPLC (B) of [3-MP<sup>0</sup>-D-Glu<sup>1</sup>,desGlu<sup>2-6</sup>]-minigastrin11 [MG11-SH]



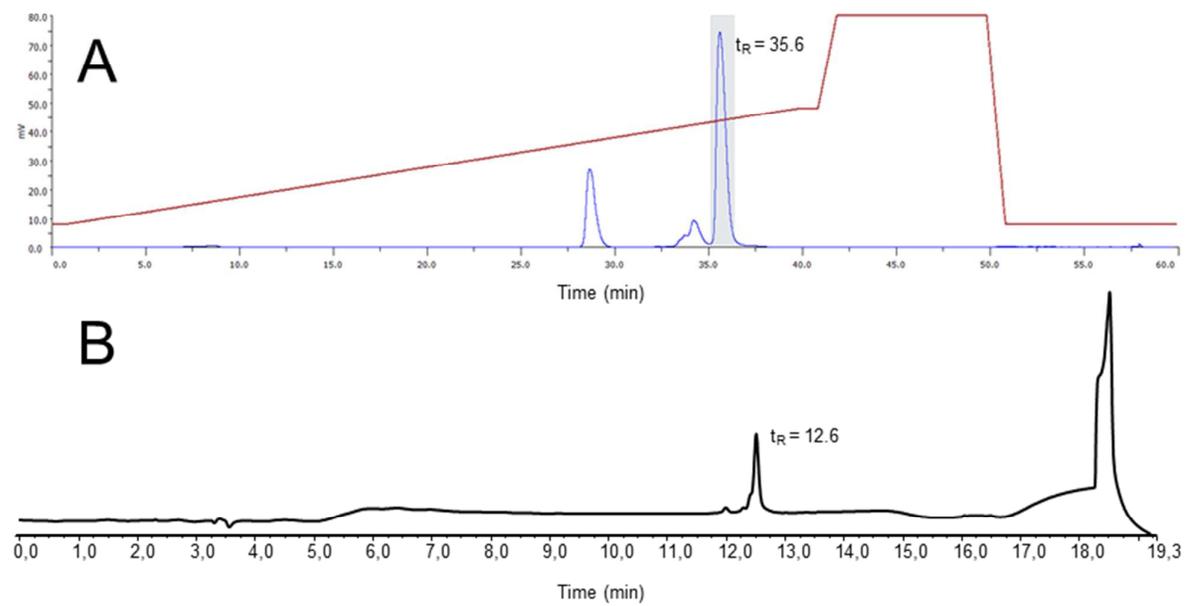
**Figure S4:** preparative RP-HPLC (**A**) and analytical RP-HPLC (**B**) of c(RGDfK)-(PEG)<sub>4</sub>-SH [RGD-SH]



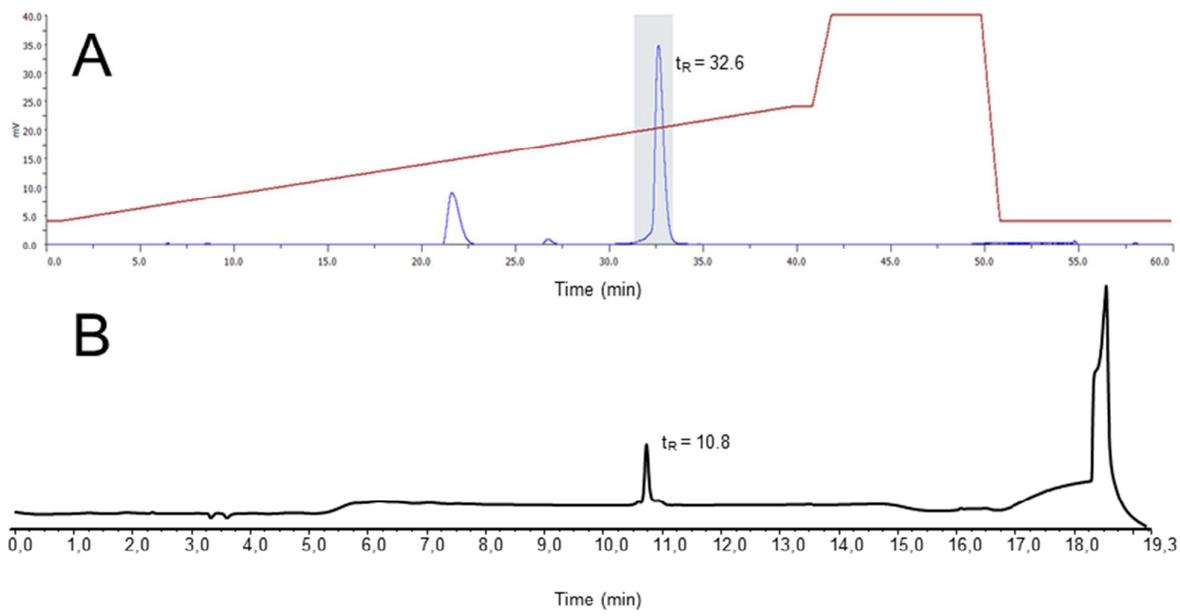
**Figure S5:** preparative RP-HPLC (**A**) and analytical RP-HPLC (**B**) of mono- and multiple (N-(3-maleinimidopropionyl) conjugated [Fe]FSC derivatives; analytical RP-HPLC (**C**) of [Fe]FSC-(N-(3-maleinimidopropionyl))<sub>2</sub> ([Fe]FSC-(mal)<sub>2</sub>)



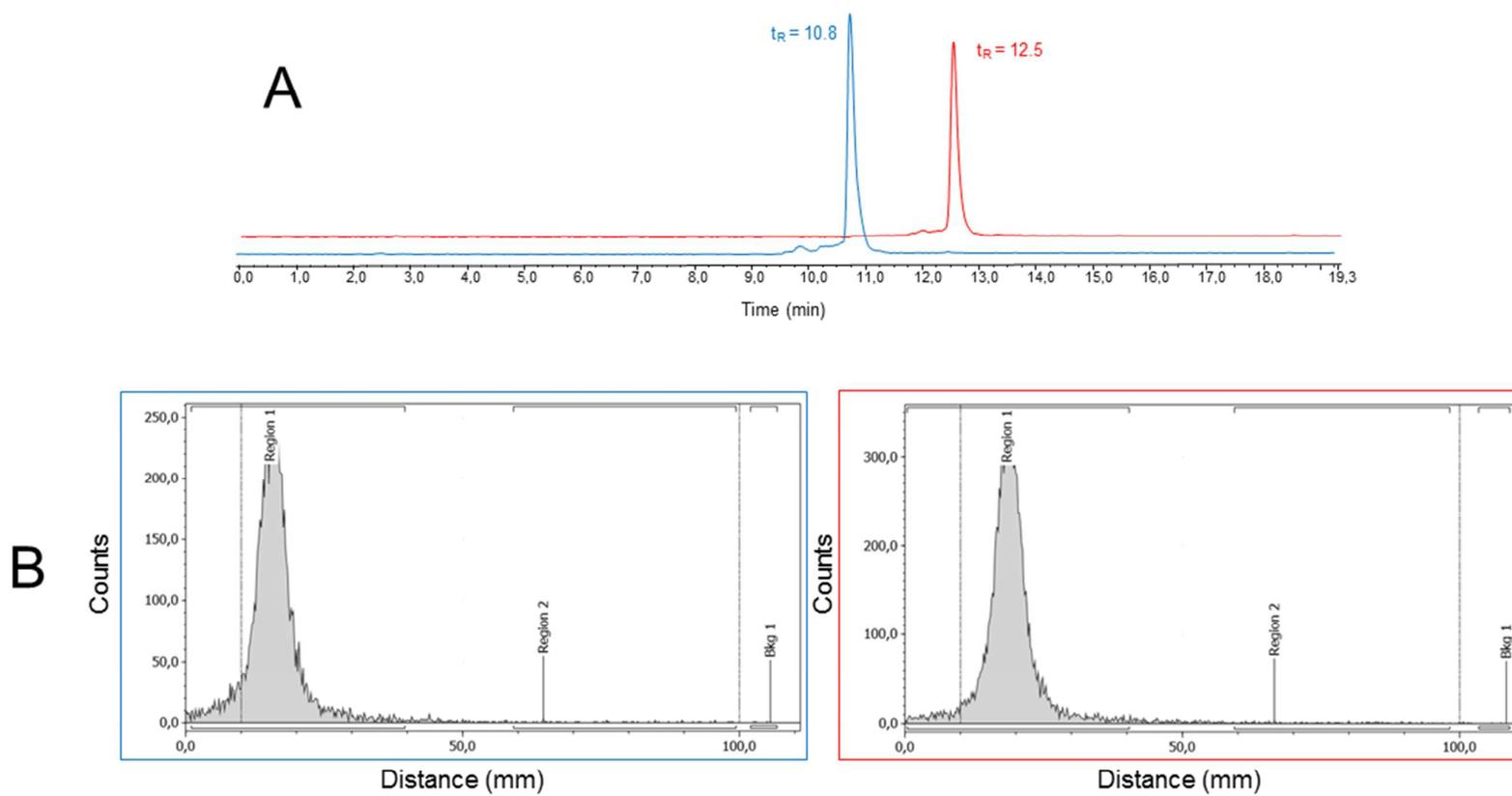
**Figure S6:** preparative RP-HPLC (A) and analytical RP-HPLC (B) of Sulfo-Cyanine7-FSC-(mal)<sub>2</sub>



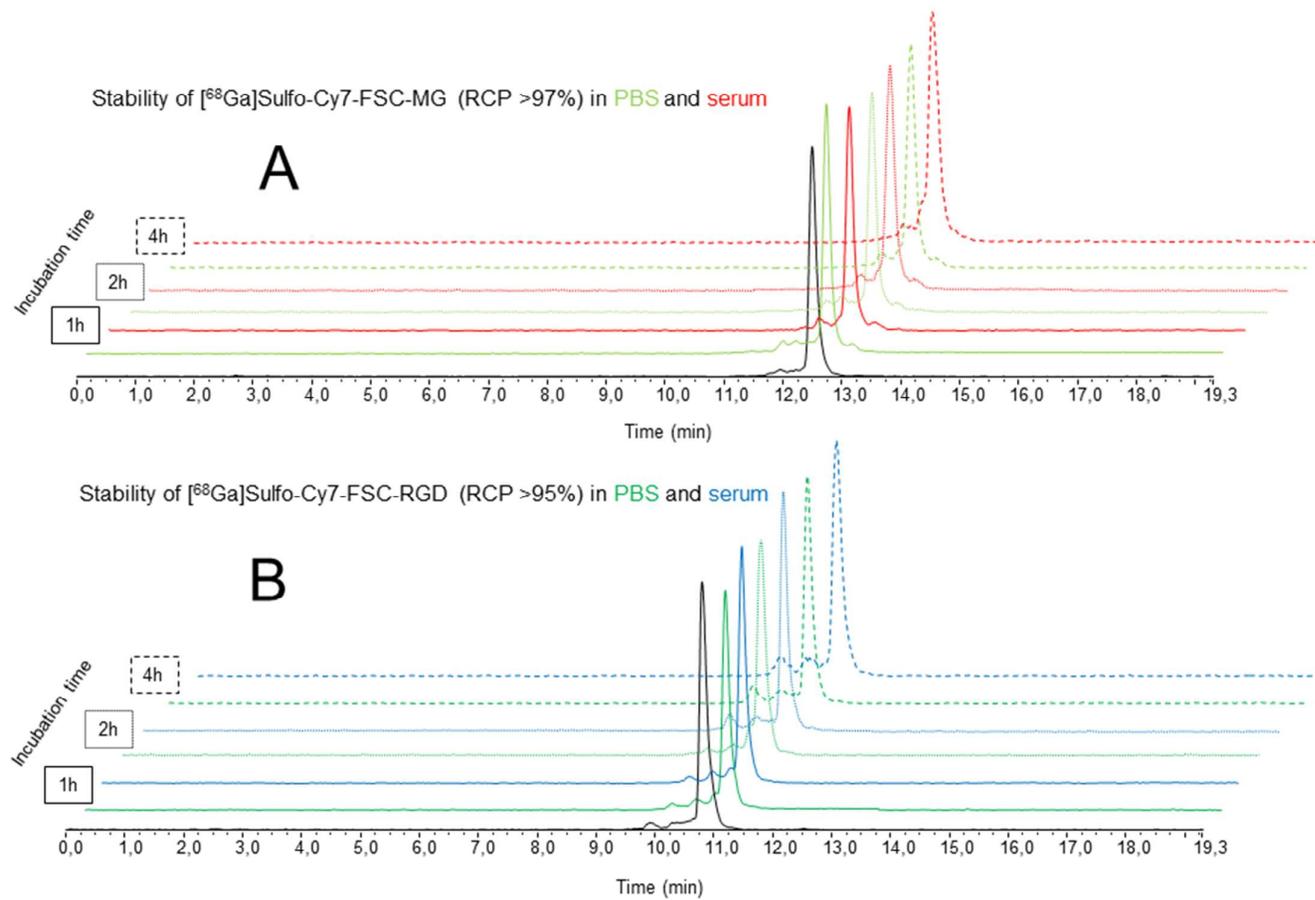
**Figure S7:** preparative RP-HPLC (A) and analytical RP-HPLC (B) of **Sulfo-Cy7-FSC-MG**



**Figure S8:** preparative RP-HPLC (A) and analytical RP-HPLC (B) of Sulfo-Cy7-FSC-RGD



**Figure S9:** analytical radio-RP-HPLC (A) and radio-ITLC (B) of [ $^{68}\text{Ga}$ ]Sulfo-Cy7-FSC-MG and [ $^{68}\text{Ga}$ ]Sulfo-Cy7-FSC-RGD



**Figure S10:** analytical radio-RP-HPLC to assess stability of [<sup>68</sup>Ga]Sulfo-Cy7-FSC-MG (**A**) and [<sup>68</sup>Ga]Sulfo-Cy7-FSC-RGD (**B**)

