

RNA analysis of activity-dependent transcriptional profiles from BLA neurons.

a, Example bioanalyzer traces of RNA samples collected from footshock (green) (n = 3), female (red) (n = 3), on dox (black) group (n = 1). Bioanalyzer traces was used to test the quality of RNA sample for RNA microarray, the graph shows the fluorescence levels, which corresponds to RNA levels, of different RNA species of different size (nt). Bioanalyzer traces showed that footshock and female samples yielded RNA samples with RNA quality number (RQN) >6 (n = 6), while the on dox RNA sample RQN <4 (n = 1). Peaks at .02kb, 1.9kb and 4.7kb correspond to the marker, 18S rRNAs, and 28S rRNAs, respectively. b, Analysis of MAS5 normalized data of arrays from the footshock (n = 3) and female (n = 3) group.



Gene expression of candidate genetic markers in the BLA using in situ hybridization. a-g, Genes that were enriched in the array of the footshock group (green). h-p, Genes that were enriched in the array of the female group (red). q-t, Positive control for interneurons

(yellow). u,v, Positive control for excitatory neurons (yellow). Micrographs represent FISH with the exception of *Ppp1r1b* (smFISH). a-v, nuclear marker, DAPI (blue). Scale bar 100µm.



Rspo2⁺ and *Ppp1r1b*⁺ BLA neurons collectively constitute all BLA pyramidal neurons.

smFISH of *Rpso2/Camk2a* (a), *Rspo2/Gad1* (b), *Ppp1r1b/Camk2a* (c), *Ppp1r1b/Gad1* (d), coronal BLA, scale bar 200 μm. e, smFISH of *Rpso2+Ppp1r1b/Camk2a*, sagittal BLA, scale bar 250 μm. f, higher magnification expression of *Rpso2+Ppp1r1b/Camk2a*, scale bar 50μm.



c-Fos protein was visualized using IHC by an Alexa Fluor 555 secondary antibody. For improved graphical representation, images were inverted and saturation removed. a, c-Fos expression across the AP-axis of the BLA in response to shock, context, female. b, c-Fos expression across the AP-axis of the BLA in response to olfactory stimuli. c, c-Fos expression across the AP-axis of the BLA in response to gustatory stimuli. Scale bar 250μm.



response to water or no water. Results show mean ± s.e.m (a-d).



Rpso2-Cre and Cartpt-Cre mice were injected with a Cre-dependent eYFP virus into the BLA and smFISH was performed against *Rspo2* and *Ppp1r1b*, respectively. a, Quantification of the percentage of *Rpso2*⁺ BLA neurons that express eYFP (eYFP/*Rspo2*) and

the percentage of eYFP⁺ BLA neurons that express *Rspo2* (*Rpso2/eYFP*) (n = 4). b, eYFP (green) and *Rspo2* (red) expression in the BLA of virus injected Rspo2-Cre mice. c, Quantification of the percentage of *Ppp1r1b*⁺ BLA neurons that express eYFP (eYFP/*Ppp1r1b*) and the percentage of eYFP⁺ BLA neurons that express *Ppp1r1b* (*Ppp1r1b/eYFP*) (n = 4). d, eYFP (green) and *Ppp1r1b* (red) expression in the BLA of virus injected Cartpt-Cre mice. Though *Ppp1r1b* is endogenously expressed outside of the BLA, such as in the intercalated cell mass, choroid plexus, and striatum (Fig. 2c), Cre-dependent virus targeted in the Cartpt-Cre mice does not express in these off targeted cells. Fuzziness of the eYFP signal reflect the effects of the protease digestion step of the smFISH. Scale bar 250µm.





smFISH of Rspo2 and Ppp1r1b in CTB injected brains. Rspo2 (a) and Ppp1r1b (b) expression in the BLA of CeC-CTB mice. Rspo2 (c) and Ppp1r1b (d) expression in the BLA of CeL/M-CTB mice. Rspo2 (e) and Ppp1r1b (f) expression in the BLA of NAc-CTB mice. Scale

bar 250µm.



Optic fiber was unilaterally implanted above the NAc of Rspo-ChR2 mice (NAc Rpso2-ChR2). NAc Rspo2-ChR2 underwent behavioral assays. a, Optogenetic freezing test (n = 9). b, Optogenetic self-stimulation test (n = 11). c, Optogenetic place preference test (n = 9). Behavioral performance was compared against Rspo2-ChR2 (Fig. 4) using an unpaired *t-test*. No significant difference was observed across all assays. d, Optic fiber placement in the NAc of Rspo2-ChR2 mice. Significance for unpaired *t-test*, all comparison between ChR2 and eYFP groups not significant (N.S).



BLA neurons to $Rspo2^+$ BLA neurons. $Ppp1r1b^+$ BLA neurons send dense fibers to the CeL and CeM. Therefore, a population in the

CeL and/or CeM may mediate appetitive behaviors.