Supplemental Figure S1



Supplemental Figure S1: Strategy for LpA-I and LpA-I/A-II IAC isolations: LpA-I and LpA-I/A-II particles were obtained from plasma or HDL isolated by ultracentrifugation (UC-HDL) using FPLC with columns containing resin enriched with antibodies specific for either human apoA-I or human apoA-II. The sample was applied to the column in PBS and bound material was eluted using 3M sodium thiocyanate. Elution fractions were immediately desalted using PD-10 desalting columns. Panel **(A)** shows the isolation scheme for subfractions obtained from plasma. Plasma was applied to the α -apoA-I column and all bound particles containing apoA-I were eluted and desalted. ApoA-I containing particles were applied to the α -apoA-II column. Unbound material contained only apoA-I with no apoA-II (LpA-I) and bound material containing both apoA-I and apoA-II (LpA-I/A-II) was desalted. Panel **(B)** shows the isolation scheme for subfractions obtained from UC-HDL. To maximize efficiency of the isolation UC-HDL was first applied to the α -apoA-II column. Unbound material was collected which contained UC-HDL with only apoA-I (LpA-I). Bound material was eluted, desalted, and applied to the α -apoA-I column. Bound material which contained both apoA-I and apoA-II (LpA-I/A-II) was eluted and desalted.



Supplemental Figure S2: Purification of LpA-I and LpA-I/A-II particles from plasma: Panels (A) and (B) show enlarged western blots from Figure 1 in the main text which were probed with antibodies for apoA-I and apoA-II, respectively. $3 \mu g$ of total protein per lane were loaded onto an Any-KDTM mini gel in sample buffer containing beta mercaptoethanol to reduce apoA-II to monomeric form. LpA-I particles are shown in lanes 2, 4, 7, and 9 and LpA-I/A-II particles are shown in lanes 3, 5, 8, and 10 with molecular weight markers in lanes 1 and 6.



Supplemental Figure S3: Purification of LpA-I and LpA-I/A-II particles from UC-HDL: 4-15% SDS-PAGE analysis of LpA-I and LpA-I/A-II particles segregated from UC-HDL for two representative donors. UC-HDL starting material is shown in lanes 2 and 6 and LpA-I particles are shown in lanes 3 and 7. LpA-I/A-II particles are shown in lanes 4 and 8 in non-reducing conditions and lanes 5 and 9 in reducing conditions. Each lane contains 6 μ g of total protein.



Supplemental Figure S4: Characterization of plasma LpA-I and LpA-I/A-II particles by agarose gel electrophoresis. Samples were analyzed by agarose gel electrophoresis using a quickgel lipoprotein electrophoresis system. Samples were loaded by volume (4 µI each) and migration was visualized by staining with Fat Red 7B. Plasma (lane 1) and UC-HDL (lane 2) were run as controls. LpA-I particles were loaded in lane 3 and LpA-I/A-II particles were loaded in lane 4.

Supplemental Figure S5



Supplemental Figure S5: Size and cross-linking analysis of rLpA-I and rLpA-I/A-II discoidal particles generated *in vitro.* Panel (**A**) shows a native gel containing a protein standards ladder (lane 1), rLpA-I discoidal particles (lane 2) and rLpA-I/A-II discoidal particles (lane 3). Panel (**B**) shows a 4-15% SDS Page analysis of the rLpA-I and rLpA-I/A-II particles before and after cross-linking at a molar ratio of 50:1 BS³:apoA-I. 10 µg of total protein for each sample was loaded and visualized by staining with Coommassie Blue. rLpA-I particles before and after cross-linking are shown in lanes 2 and 4, respectively. rLpA-I/A-II particles before and after cross-linking are shown in lanes 3 and 5, respectively.



Supplemental Figure S6. Cholesterol efflux capacity of rLpA-I particles spiked with lipid-free apoA-I and rLpA-I/A-II POPC particles. Lipid free plasma apoA-I was spiked into rLpA-I POPC particle at equivalent masses found in rLpA-I/A-II POPC particles using densitometry analysis on gels shown in Supplemental Figure 4A. Panel (A): rLpA-I POPC particles with and without addition of lipid-free plasma apoA-I were loaded by equal PC mass (20 µg/mI) and cholesterol efflux capacity was quantified in the presence and absence of cAMP. Panel (B): rLpA-I POPC particles spiked with lipid-free apoA-I and rLpA-I/A-II POPC particles were loaded by equal PC mass (20 µg/mI) and cholesterol efflux capacity was quantified in the presence and absence of cAMP. Panel (C): rLpA-I and rLpA-I/A-II POPC particles were loaded by equal PC mass (20 µg/mI) and cholesterol efflux capacity was quantified in the presence and absence of cAMP. Panel (C): rLpA-I and rLpA-I/A-II POPC particles were loaded by equal PC mass (20 µg/mI) and cholesterol efflux capacity was quantified in the presence of cAMP by measuring total cholesterol in the media using an Amplex Red Cholesterol assay kit. Cholesterol mass was normalized by total cell protein measured using the Markwell Lowry assay. Bars represent the average of three independent preparations (±SD) and asterisks denote statistically significant differences (P<0.05) found between the subfractions determined by a 2-tailed, paired student's t-test.

Supplemental Figure S7



Supplemental Figure S7: Limited proteolysis analysis of LpA-I and LpA-I/A-II particles. LpA-I and LpA-I/A-II subfractions isolated from plasma, UC-HDL, and synthetic discoidal particles were incubated sequencing grade trypsin for the indicated time points at 37°C. Reactions were quenched by addition of SDS sample buffer and boiling samples for 10 min at 100°C. Samples were frozen at -20°C until ready for analysis by SDS-PAGE. Panel (A) shows proteolytic digestion of apoA-I from LpA-I and LpA-I/A-II fractions isolated from three plasma donors. A total of 8 µg of total protein was incubated with trypsin at a ratio of 20:1 Pro:trypsin. LpA-I particles are in lanes 2, 4 and 6. LpA-I/A-II particles are in lanes 3, 5, and 7. Panel (B) shows proteolytic digestion of apoA-I from LpA-I and LpA-I/A-II fractions isolated from two donors. A total of 6 µg of total protein was incubated with trypsin at a ratio of 50:1 Pro:trypsin. LpA-I/A-II particles are in lanes 2 and 3 and LpA-I particles are in lanes 3 and 4. ApoA-II migrates as a monomer as 2-Mercaptoethanol was present in the SDS sample buffer. Panel (C) shows the proteolytic digestion of apoA-I from discoidal rLpA-I/A-II particles from three independent preps. A total of 4 µg of total protein was incubated with trypsin at a ratio of 40:1 Pro:trypsin. LpA-I particles are in lanes 2, 4 and 6 and rLpA-I/A-II particles are in lanes 3, 5 and 7.

· · ·		Lp	A-I			LpA-I/A-II						
Accession Number	Control ^a	1 ^b	2 ^b	3 ^b	4 ^b	Control ^a	1 ^b	2 ^b	3 ^b	4 ^b		
ALBU	73	383	197	270	333		29	18	12	14		
IGHM	12	165	116	142	71		43	95	88	69		
IGKC	5	96	48	70	85		56	67	100	54		
IGHG1	5	65	35	52	67	1	27	18	39	49		
K2C1	30	9	6	10	1	1	10	7	10	8		
HPTR	1	46	29	30	15		11	18	9	8		
TRFE	7	60	49	49	46							
K1C10	14	10	7	7	3	1	8	5	5	6		
IGHG4	4	54	29	26	52		27	24	25	34		
K1C9	8	3	3	8			2	4	7	6		
A2MG	1	20	9	11	27							
K22E	9	4	4	3	2		3	3	4	2		
A1AG1	5	10	6	8	10		2	2	1	2		
HPT	2	31	22	21	23					5		
K1C14	1											

Supplemental Table S1 Total peptide spectral counts for non-specifically bound proteins

^a Peptide counts for proteins in a control sample isolated from plasma applied to a dummy column containing resin and no antibodies followed by application to the α-apoA-II column.
^b Peptide counts for non-specifically bound proteins identified in individual donors.

I otal peptide spectral counts	s for prote	ins ide	entitie	d în U	C-HDI	L an	d AI-LI	S	
Protoin	Accession		UC-	HDL			_ps		
Protein	Number	1 ^a	2 ^a	3 ^a	4 ^a	1 ^a	2 ^a	3 ^a	4 ^a
Apolipoprotein A-I	APOA1	520.4	596.5	579.3	539.0		1097.4		1126.0
Apolipoprotein A-II	APOA2	155.9	149.4	177.0	175.0		225.4		204.0
Fibronectin	FINC						126.5		97.0
Apolipoprotein B-100	APOB	64.2	2.6	5.6	40.0		70.4		99.0
Apolipoprotein A-IV	APOA4	9.2	7.7	15.7	16.0		29.6		35.0
Clusterin	CLUS	3.4	2.6	1.1			63.2		70.0
Apolipoprotein D	APOD	37.8	30.7	43.7	21.0		17.3		19.0
	APOC3	29.8	47.3	24.7	32.0		23.5		26.0
Serum paraoxonase/arylesterase 1	ABOC1	3.4	21.7	12.3	2.0		24.5		21.0
Apolipoprotein C-i		20.4	30.0 2 0	20.9	20.0		0.Z 10.2		14.0
Apolipoprotein L		26.4	26.8	20.0	26.0		8.2		9.0
Apolipoprotein C-II	APOC2	29.8	35.8	21.3	30.0		4 1		3.0
Alpha-1-antitrypsin	A1AT	33.2	10.2	9.0	7.0		16.3		17.0
laGEc-binding protein	FCGBP						19.4		19.0
lg lambda-2 chain C regions	LAC2						19.4		15.0
Fibrinogen gamma chain	FIBG						10.2		12.0
lg alpha-1 chain C region	IGHA1						12.2		9.0
Phosphatidylinositol-glycan-specific phospholipase D	PHLD						13.3		14.0
Fibrinogen beta chain	FIBB						12.2		17.0
Apolipoprotein L1	APOL1	5.7	3.8	6.7	6.0		9.2		10.0
Serum amyloid A-4 protein	SAA4	2.3	5.1	9.0	13.0				
Fibrinogen alpha chain	FIBA	1.1					2.0		4.0
Ig gamma-2 chain C region	IGHG2						17.3		18.0
Complement C3	CO3	1.1					11.2		12.0
Coagulation factor V	FA5						7.1		13.0
Phospholipid transfer protein	PLTP	2.3	2.6	3.4	1.0		4.1		4.0
Complement C1q subcomponent subunit B	C1QB						16.3		12.0
Apolipoprotein F	APOF	12.6	7.7	5.6	6.0		2.0		2.0
Vitronectin	VTNC						7.1		8.0
CD5 antigen-like	CD5L						8.2		
Kininogen-1	KNG1						10.2		9.0
Ig kappa chain V-III region HAH	KV312						7.1		5.0
Immunoglobulin J chain	IGJ						3.1		
Complement C 1q subcomponent subunit C							10.2		11.0
Serum amyloid A-1 protein	SAA1 CO44	3.4		2.2	2.0		2.0		10.0
Complement C1r subcomponent	C04A	1.1		1.1			2.0		5.0 2.0
Inter alpha trypsin inhibitor haavy chain H4							2.0		2.0
Transthyretin		69			1.0		1.0		2.0
Hemopexin	HEMO	0.5					5.1		3.0
Alpha-1-acid glycoprotein 2	A1AG2						5.1		4.0
Plasminogen	PLMN						5.1		5.0
Beta-2-glycoprotein 1	APOH						7.1		7.0
Coagulation factor XI	FA11						1.0		1.0
Ig gamma-3 chain C region	IGHG3						9.2		
C4b-binding protein alpha chain	C4BPA						2.0		3.0
Vitamin D-binding protein	VTDB						1.0		2.0
Cholesteryl ester transfer protein	CETP						3.1		5.0
Ig alpha-2 chain C region	IGHA2						8.2		
Alpha-2-HS-glycoprotein	FETUA						1.0		
Prothrombin	THRB						2.0		
Alpha-1B-glycoprotein	A1BG						1.0		
Phosphatidylcholine-sterol acyltransferase	LCAT								1.0
Angiotensinogen	ANGT						4.1		
Alpha-1-antichymotrypsin	AACT						1.0		
Ceruloplasmin	CERU						2.0		
Ficolin-2	FCN2						1.0		1.0
Histidine-rich glycoprotein	HRG								1.0

^a Total peptide spectral counts for proteins identified in individual donors. Spectral counts were normalized to the total spectral count summed across all proteins for the subfraction in the donor with the maximal total spectral count within each method; i.e. spectral counts for UC-HDL were normalized to max total spectral counts with the UC-HDL donors and spectral counts for AI-LPs were normalized to the max total spectral counts with the AI-LPs donors.

Total peptide spectral counts for proteins identified in the LpA-I and LpA-I/A-II subfractions isolated from plasma

			In/	A_I			Ln۸-	1/4-11	
Protein	Accession	, a	 a	 	, a	, a	a	v A-11 م a	, a
	Nulliber	1	2	3	4	1	2	3	4
Apolipoprotein A-I	APOA1	740.8	1061.2	938.0	749.8	1063.0	11//.9	1098.6	1022.3
Apolipoprotein A-li	APOA2	2.2	2.4	1.0	2.2	188.0	1/1.5	151.4	181.3
Fibronectin Anolinoprotoin P 100		191.1	197.0	244.0	270.4	70.0	F1 7	72.0	05.0
Apolipoprotein B-100		02.0	00.Z 05.8	97.0 86.0	75.0	11/1 7	108.5	13/1	90.0 142.6
Clusterin		137.1	73.1	87.0	79.5	75.7	61.8	70.8	72 9
Apolipoprotein D	APOD	27.0	37.1	32.0	24.6	62.7	73.1	57.1	49.7
Apolipoprotein C-III	APOC3	8.6	16.8	10.0	9.0	39.0	56.7	34.8	34.3
Serum paraoxonase/arvlesterase 1	PON1	79.9	59.9	55.0	60.4	44.9	40.4	37.2	38.7
Apolipoprotein C-I	APOC1	8.6	20.4	10.0	9.0	23.6	32.8	33.5	27.6
Apolipoprotein E	APOE	16.2	27.5	22.0	24.6	31.9	37.8	53.4	63.0
Apolipoprotein M	APOM	11.9	10.8	13.0	9.0	15.4	17.7	16.1	18.8
Apolipoprotein C-II	APOC2	13.0	31.1	16.0	20.1	24.8	36.6	22.3	24.3
Alpha-1-antitrypsin	A1AT	37.8	15.6	25.0	26.9	11.8	3.8	11.2	19.9
IgGFc-binding protein	FCGBP	42.1	12.0	37.0	31.3	43.7	20.2	14.9	17.7
Ig lambda-2 chain C regions	LAC2	36.7	32.3	24.0	25.7	39.0	21.4	18.6	29.8
Fibrinogen gamma chain	FIBG	36.7	38.3	48.0	69.4	7.1	6.3	8.7	8.8
Ig alpha-1 chain C region	IGHA1	43.2	27.5	32.0	38.1	17.7	16.4	42.2	18.8
Phosphatidylinositol-glycan-specific phospholipase D	PHLD	41.0	10.8	35.0	49.2	16.6	3.8	11.2	19.9
Fibrinogen beta chain	FIBB	35.6	35.9	34.0	49.2	7.1	6.3	3.7	9.9
Apolipoprotein L1	APOL1	8.6	7.2	9.0	14.5	11.8	3.8	5.0	17.7
Serum amyloid A-4 protein	SAA4	2.2	4.8	5.0	5.6	3.5	5.0	7.4	8.8
Fibrinogen alpha chain	FIBA	29.2	27.5	34.0	41.4	1.2	2.5	1.2	4.4
ry gainina-2 chain C region		34.6	0.6	10 0	44.0 22.6	40.1	40.4	00.0	42.0
Congulation factor V	EA5	36.7	20.4	13.0	16.8	24		12	1.0
Phospholinid transfer protein	PLTP	13.0	6.0	11.0	11.2	16.6	13.9	8.7	13.3
Complement C1g subcomponent subunit B	C10B	7.6	15.6	11.0	15.7	5.9	5.0	5.0	7.7
Apolipoprotein F	APOF	6.5	7.2	4.0	2.2	8.3	8.8	7.4	11.1
Vitronectin	VTNC	18.4	12.0	9.0	13.4	9.5	6.3	5.0	8.8
CD5 antigen-like	CD5L	14.0	20.4	13.0	4.5	3.5	16.4	7.4	3.3
Kininogen-1	KNG1	18.4	15.6	15.0	15.7				
Ig kappa chain V-III region HAH	KV312	11.9	4.8	10.0	12.3	5.9	8.8	14.9	6.6
Immunoglobulin J chain	IGJ	11.9	16.8	11.0	7.8	4.7	12.6	11.2	3.3
Complement C1q subcomponent subunit C	C1QC	7.6	14.4	7.0	11.2	3.5	6.3	5.0	7.7
Serum amyloid A-1 protein	SAA1	2.2	2.4	2.0	3.4	8.3	2.5	6.2	6.6
Complement C4-A	CO4A	9.7	8.4	2.0	7.8	9.5	6.3	2.5	2.2
Complement C1r subcomponent		14.0	7.2	9.0	10.1	1.2	1.3	1.2	2.2
Fidelet ideloi 4		0.5	2.4	11.0	20.7				1.1
Transthyratin		9.7 7.6	2.4 4.8	9.0	7.8	24	13		1 1
Hemonexin	HEMO	10.8	4.8	10.0	7.0	2.7			
Alpha-1-acid glycoprotein 2	A1AG2	8.6	6.0	8.0	9.0	5.9	5.0	2.5	4.4
Plasminogen	PLMN	3.2	2.4	5.0	3.4		1.3		
Beta-2-glycoprotein 1	APOH		4.8	7.0	10.1				
Coagulation factor XI	FA11	5.4	4.8	5.0	11.2				
Ig gamma-3 chain C region	IGHG3	50.8	25.2	35.0	50.4	26.0		34.8	37.6
C4b-binding protein alpha chain	C4BPA	7.6	3.6	3.0	3.4				1.1
Vitamin D-binding protein	VTDB	6.5	1.2	7.0	6.7				
Protein AMBP	AMBP	7.6	4.8	3.0	3.4		1.3		
Cholesteryl ester transfer protein	CETP	1.1		1.0		7.1		3.7	5.5
lg alpha-2 chain C region	IGHA2	25.9	13.2	19.0		14.2	11.3	26.1	
Alpha-2-HS-glycoprotein	FEIUA	5.4	2.4	4.0	4.5				
Prothrombin	THRB	2.2	3.6	1.0	3.4				
Alpha-Lo-glycoprotein	AIBG	5.4 7.6		∠.U 1 0	4.5 1 5				
Complement factor n Phoenbatidyloboling-storol acyltransforace		1.0		1.0	4.5			 1 0	 5 5
Andiotensingan		3.2	36			3.0 1.2		i.z	5.5
Alpha-1-antichymotrynsin	AACT	43	3.6	20	22				
Ceruloplasmin	CFRU	4.3	1.2	2.0	4.5				
Ficolin-2	FCN2	3.2		1.0	2.2				
Complement C1s subcomponent	C1S	6.5	1.2		2.2				
Inter-alpha-trypsin inhibitor heavy chain H2	ITIH2	4.3		1.0	1.1				
Platelet basic protein	CXCL7			1.0	3.4				

^a Total peptide spectral counts for proteins identified in individual donors. Spectral counts were normalized to the total spectral count summed across all proteins for the subfraction in the donor with the maximal total spectral count.

Total peptide spectral counts for proteins identified in the LpA-I and LpA-I/A-II subfractions isolated from UC-HDL

······································												
Dustain	Accession		Lp	A-I		LpA-I/A-II						
Protein	Number	1 ^a	2 ^a	3 ^a	4 ^a	1 ^a	2 ^a	J/A-II 3 a 591.0 174.0 174.0 26.0 1.0 48.0 40.0 13.0 28.0 37.0 36.0 20.0 6.0 7.0 14.0 2.0 11.0 3.0 3.0	4 ^a			
Apolipoprotein A-I	APOA1	776.6	762.2	837.7	787.4	632.3	646.6	591.0	605.8			
Apolipoprotein A-II	APOA2	29.9	52.8	42.3	56.3	171.3	161.3	174.0	177.5			
Apolipoprotein B-100	APOB	12.6	3.7		35.3	1.3			3.6			
Apolipoprotein A-IV	APOA4	4.7	13.5	2.6	2.2	7.6	17.8	26.0	18.0			
Clusterin	CLUS	6.3		1.3				1.0				
Apolipoprotein D	APOD	9.4	23.3	17.9	5.5	40.3	34.0	48.0	20.4			
Apolipoprotein C-III	APOC3	31.4	38.0	28.2	28.7	46.6	51.8	40.0	55.2			
Serum paraoxonase/arylesterase 1	PON1	4.7	20.9	14.1	5.5	3.8	10.4	13.0	1.2			
Apolipoprotein C-I	APOC1	22.0	23.3	16.7	25.4	29.0	44.4	28.0	42.0			
Apolipoprotein E	APOE	3.1	7.4	1.3	15.5	22.7	11.8	37.0	49.2			
Apolipoprotein M	APOM	33.0	36.8	33.3	23.2	31.5	20.7	36.0	19.2			
Apolipoprotein C-II	APOC2	40.9	49.1	28.2	37.5	34.0	41.4	20.0	36.0			
Alpha-1-antitrypsin	A1AT	50.3	8.6	11.5	6.6	13.9	3.0	6.0	6.0			
Apolipoprotein L1	APOL1	12.6	1.2	2.6	8.8	7.6		7.0	6.0			
Serum amyloid A-4 protein	SAA4	6.3	8.6	9.0	12.1	2.5	5.9	14.0	7.2			
Phospholipid transfer protein	PLTP	6.3	2.5	5.1	2.2	1.3	3.0	2.0	1.2			
Apolipoprotein F	APOF	6.3	4.9	5.1	2.2	8.8	4.4	11.0	6.0			
Serum amyloid A-1 protein	SAA1	1.6	1.2	1.3	3.3	2.5		3.0	3.6			

^a Total peptide spectral counts for proteins identified in individual donors. Spectral counts were normalized to the total spectral count summed across all proteins for the subfraction in the donor with the maximal total spectral count.

Spectral count distribution for proteins across LpA-I and LpA-I/A-II subfractions isolated from plasma

Accession		Lp	A-I	1		, LpA	-I/A-II	1	LpA-I	LpA-I/A-II	LpA-I	, LpA-I/A-II	P-
Number	1 ^a	2 ^a	3 ^a	4 ^a	1 ^a	2 ^a	3 ^a	4 ^a	Avg	Avg	SD	SD	value b
APOA1	41.1%	47.4%	46.1%	42.3%	58.9%	<u>-</u> 52.6%	53.9%	57.7%	44.2%	55.8%	3.0%	3.0%	0.031
APOA2	1.1%	1.4%	0.7%	1.2%	98.9%	98.6%	99.3%	98.8%	1.1%	98.9%	0.3%	0.3%	0.000
FINC	63.7%	63.8%	69.6%	71.0%	36.3%	36.2%	30.4%	29.0%	67.0%	33.0%	3.8%	3.8%	0.003
APOB	62.6%	60.8%	57.4%	57.1%	37.4%	39.2%	42.6%	42.9%	59.5%	40.5%	2.7%	2.7%	0.006
APOA4	44.7%	46.9%	39.1%	34.5%	55.3%	53.1%	60.9%	65.5%	41.3%	58.7%	5.6%	5.6%	0.054
	04.4% 20.1%	54.2% 22.7%	25.1%	52.1% 33.1%	35.6% 60.0%	45.8%	44.9% 64.1%	47.9% 66.0%	33.2%	43.5%	5.5% 2.4%	5.5% 2.4%	0.098
APOC3	18.1%	22.8%	22.3%	20.7%	81.9%	77.2%	77.7%	79.3%	21.0%	79.0%	2.1%	2.1%	0.000
PON1	64.0%	59.7%	59.6%	61.0%	36.0%	40.3%	40.4%	39.0%	61.1%	38.9%	2.0%	2.0%	0.002
APOC1	26.8%	38.3%	23.0%	24.5%	73.2%	61.7%	77.0%	75.5%	28.1%	71.9%	7.0%	7.0%	0.008
APOE	33.7%	42.1%	29.2%	28.1%	66.3%	57.9%	70.8%	71.9%	33.3%	66.7%	6.4%	6.4%	0.014
APOM	43.6%	37.9%	44.6%	32.3%	56.4%	62.1%	55.4%	67.7%	39.6%	60.4%	5.7%	5.7%	0.036
	34.3% 76.2%	46.0% 80.5%	41.7% 60.1%	45.3% 57.4%	65.7% 23.8%	54.0% 10.5%	58.3% 30.0%	54.7% 42.6%	41.8% 70.8%	58.2% 20.2%	5.4% 10.1%	5.4% 10.1%	0.056
FCGBP	49.0%	37.2%	71.3%	63.9%	23.0 <i>%</i>	62.8%	28.7%	36.1%	55.4%	44.6%	15.2%	15.2%	0.531
LAC2	48.5%	60.1%	56.3%	46.3%	51.5%	39.9%	43.7%	53.7%	52.8%	47.2%	6.5%	6.5%	0.451
FIBG	83.8%	85.9%	84.7%	88.7%	16.2%	14.1%	15.3%	11.3%	85.8%	14.2%	2.1%	2.1%	0.000
IGHA1	70.9%	62.7%	43.1%	66.9%	29.1%	37.3%	56.9%	33.1%	60.9%	39.1%	12.3%	12.3%	0.175
PHLD	71.3%	74.0%	75.8%	71.2%	28.7%	26.0%	24.2%	28.8%	73.1%	26.9%	2.2%	2.2%	0.000
FIBB	83.4%	85.1%	90.1%	83.2%	16.6%	14.9%	9.9%	16.8%	85.4%	14.6%	3.2%	3.2%	0.000
SAA4	42.2% 37.8%	00.0% 48.7%	04.4% 40.2%	40.1%	57.6% 62.2%	34.5% 51 3%	33.0% 59.8%	54.9% 61.2%	54.5% 41.4%	45.7%	12.4% 5.0%	5.0%	0.534
FIBA	96.1%	91.6%	96.5%	90.4%	3.9%	8.4%	3.5%	9.6%	93.6%	6.4%	3.1%	3.1%	0.000
IGHG2	52.4%	46.3%	35.2%	51.6%	47.6%	53.7%	64.8%	48.4%	46.4%	53.6%	7.9%	7.9%	0.426
CO3	80.7%	100.0%	100.0%	85.9%	19.3%			14.1%	91.6%	8.4%	9.9%	9.9%	0.004
FA5	93.9%	100.0%	91.3%	93.8%	6.1%		8.7%	6.2%	94.8%	5.2%	3.7%	3.7%	0.000
PLTP	43.9%	30.2%	55.9%	45.8%	56.1%	69.8%	44.1%	54.2%	43.9%	56.1%	10.6%	10.6%	0.334
	56.1% 43.0%	/5.5% // 0%	68.9% 34.0%	66.9% 16.8%	43.9% 56.1%	24.5% 55.1%	31.1% 65.1%	33.1%	66.9% 35.1%	33.1%	8.1% 13.0%	8.1% 13.0%	0.025
VTNC	43.3 <i>%</i>	65.5%	64 4%	60.3%	34.0%	34.5%	35.6%	39.7%	64.1%	35.9%	2.6%	2.6%	0.002
CD5L	79.8%	55.4%	63.6%	57.4%	20.2%	44.6%	36.4%	42.6%	64.1%	35.9%	11.1%	11.1%	0.085
KNG1	100.0%	100.0%	100.0%	100.0%					100.0%	0.0%	0.0%	0.0%	
KV312	66.8%	35.2%	40.2%	65.0%	33.2%	64.8%	59.8%	35.0%	51.8%	48.2%	16.4%	16.4%	0.843
IGJ	71.5%	57.1%	49.6%	70.3%	28.5%	42.9%	50.4%	29.7%	62.1%	37.9%	10.6%	10.6%	0.106
	68.1%	69.5%	58.5%	59.1%	31.9%	30.5%	41.5% 75.6%	40.9%	03.8% 24.00/	30.2%	5.8%	5.8%	0.062
CO4A	20.7% 50.7%	40.7% 57.1%	24.4% 44.6%	33.0% 78.0%	19.3%	12 Q%	75.0% 55.4%	22.0%	57.6%	00.2% 42.4%	12.5%	12.5%	0.002
C1R	92.2%	85.1%	87.9%	82.0%	7.8%	14.9%	12.1%	18.0%	86.8%	13.2%	4.3%	4.3%	0.000
PLF4	100.0%	100.0%	100.0%	95.9%				4.1%	99.0%	1.0%	2.1%	2.1%	0.000
ITIH4	100.0%	100.0%	100.0%	100.0%					100.0%	0.0%	0.0%	0.0%	
TTHY	76.2%	79.2%	100.0%	87.6%	23.8%	20.8%		12.4%	85.7%	14.3%	10.7%	10.7%	0.007
HEMO	100.0%	100.0%	100.0%	100.0%					100.0%	0.0%	0.0%	0.0%	0.050
	59.4% 100.0%	54.3% 65.5%	100.0%	100.9%	40.0%	40.7% 34.5%	23.1%	33.1%	04.2% 91.4%	35.6%	9.0%	9.0%	0.059
APOH		100.0%	100.0%	100.0%					100.0%	0.0%	0.0%	0.0%	0.017
FA11	100.0%	100.0%	100.0%	100.0%					100.0%	0.0%	0.0%	0.0%	
IGHG3	66.1%	100.0%	50.2%	57.3%	33.9%		49.8%	42.7%	68.4%	31.6%	22.1%	22.1%	0.194
C4BPA	100.0%	100.0%	100.0%	75.2%				24.8%	93.8%	6.2%	12.4%	12.4%	0.006
VTDB	100.0%	100.0%	100.0%	100.0%					100.0%	0.0%	0.0%	0.0%	0.000
	13 2%	19.2%	21.2%	100.0%	86.8%	20.6%	78.8%	100.0%	94.0% 11.5%	3.2% 88.5%	10.4%	10.4%	0.003
IGHA2	64.6%	53.7%	42.2%		35.4%	46.3%	57.8%		53.5%	46.5%	11.2%	11.2%	0.643
FETUA	100.0%	100.0%	100.0%	100.0%					100.0%	0.0%	0.0%	0.0%	0.010
THRB	100.0%	100.0%	100.0%	100.0%					100.0%	0.0%	0.0%	0.0%	
A1BG	100.0%		100.0%	100.0%					100.0%	0.0%	0.0%	0.0%	
CFAH	100.0%		100.0%	100.0%					100.0%	0.0%	0.0%	0.0%	0.000
	23.3%				16.1% 26.7%		100.0%	100.0%	/.8% 86.6%	92.2%	13.5%	13.5%	0.032
ANGT	100.0%	100.0%	100.0%	100.0%	20.770				100.0%	0.0%	0.0%	0.0%	0.223
CERU	100.0%	100.0%	100.0%	100.0%					100.0%	0.0%	0.0%	0.0%	
FCN2	100.0%		100.0%	100.0%					100.0%	0.0%	0.0%	0.0%	
C1S	100.0%	100.0%		100.0%					100.0%	0.0%	0.0%	0.0%	
ITIH2	100.0%		100.0%	100.0%					100.0%	0.0%	0.0%	0.0%	
CXCL7			100.0%	100.0%					100. 0 %	0.0%	0.0%	0.0%	

^a Distribution of peptide spectral counts between LpA-I and LpA-I/A-II subfractions isolated from plasma. Total spectral counts for a given protein were summed across LpA-I and LpA-I/A-II subfractions for individual donors. Spectral counts for each individual are expressed as a percentage of that total.

^b Statistically significant differences (P<0.05) are highlighted in red as calculated by a 2-tailed student's t-test between LpA-I and LpA-I/A-II subfractions.

Supplemental Table S6
Protein grouping for gene ontology analysis.

Function	Acute Inflammatory Response	Acute- Phase Response	Complement Activation	Hemostasis	Immune Response	Lipid Metabolic Process	Lipid Transport	Metal Ion Binding	Protease Inhibitor
	FINC	FINC	CLUS	FINC	APOA4	APOB	APOB	FINC	A1AT
	CLUS	A1AT	LAC2	APOB	CLUS	APOA4	APOA4	APOA4	CO3
	A1AT	SAA4	IGHA1	CLUS	LAC2	CLUS	CLUS	PON1	KNG1
	LAC2	FIBA	IGHG2	A1AT	IGHA1	APOD	APOD	APOE	CO4A
	IGHA1	SAA1	CO3	FIBG	FIBB	APOC3	APOC3	FIBG	ITIH4
	SAA4	ITIH4	C1QB	FIBB	APOL1	PON1	APOC1	FA5	AMBP
	FIBA	A1AG2	KV312	FIBA	FIBA	APOC1	APOE	KNG1	FETUA
	IGHG2	FETUA	C1QC	CO3	IGHG2	APOE	APOM	C1R	ANGT
	CO3	THRB	CO4A	FA5	CO3	APOM	APOC2	HEMO	AACT
	C1QB	AACT	C1R	KNG1	C1QB	APOC2	APOL1	THRB	ITIH2
5	KV312		IGHG3	SAA1	VTNC	PHLD	PLTP	CERU	HRG
mbel	C1QC		C4BPA	PLF4	KV312	APOL1	APOF	C1S	
n Nu	SAA1		IGHA2	PLMN	IGJ	CO3	APOH	HRG	
ssio	CO4A		CFAH	APOH	C1QC	PLTP	CETP		
Acce	C1R		FCN2	FA11	SAA1	APOF	LCAT		
	ITIH4		C1S	THRB	CO4A	TTHY			
	A1AG2			CXCL7	C1R	APOH			
	IGHG3			HRG	PLF4	VTDB			
	C4BPA				IGHG3	CETP			
	IGHA2				C4BPA	LCAT			
	FETUA				IGHA2	ANGT			
	THRB				CFAH				
	CFAH				FCN2				
	AACT				C1S				
	FCN2				CXCL7				
	C1S								