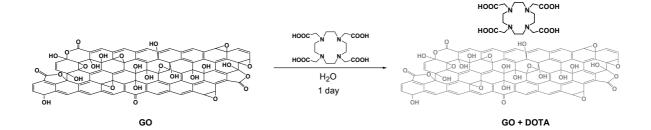
## **Supporting Information**

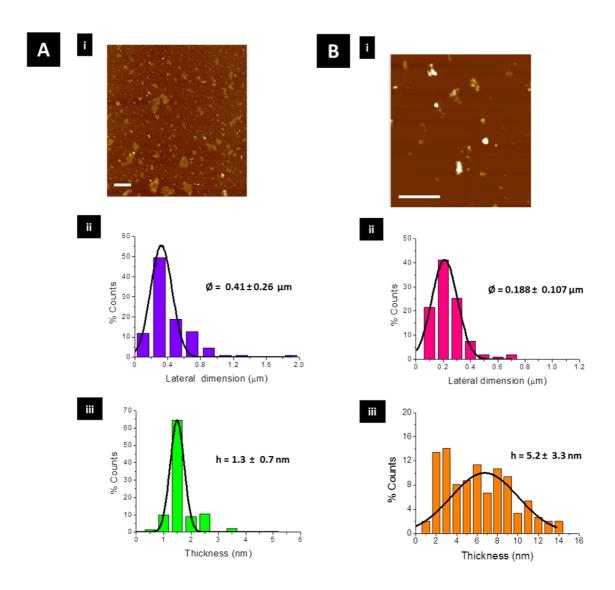
## Tissue Distribution and Urinary Excretion of Intravenously Administered Chemically Functionalized Graphene Oxide Sheets

Dhifaf A. Jasim, Cécilia Ménard-Moyon, Dominique Bégin, Alberto Bianco\*, Kostas Kostarelos\*

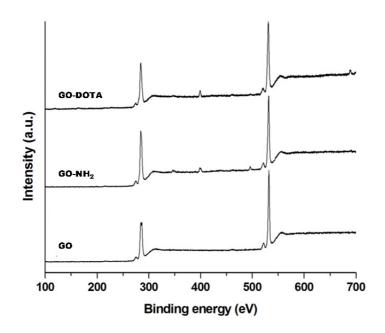
## **Supporting Figures**



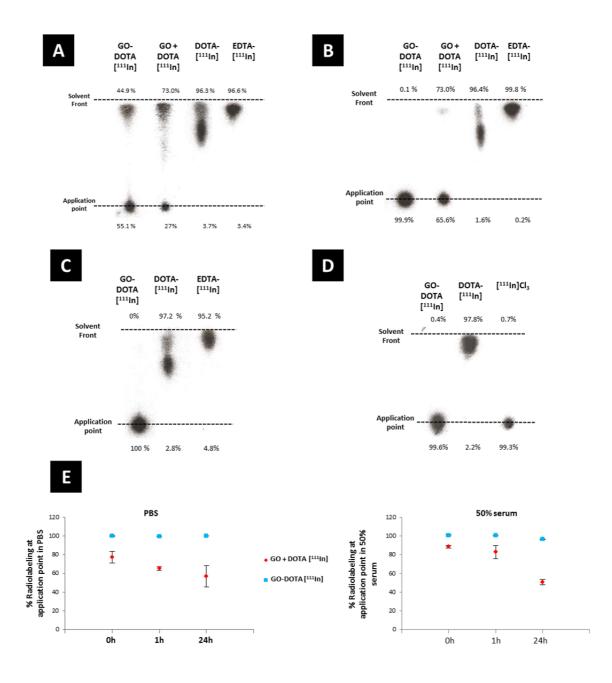
**Scheme S1:** Preparation of control sample GO + DOTA. GO was simply mixed with DOTA and treated to remove excess DOTA which was not physically adsorbed onto GO.



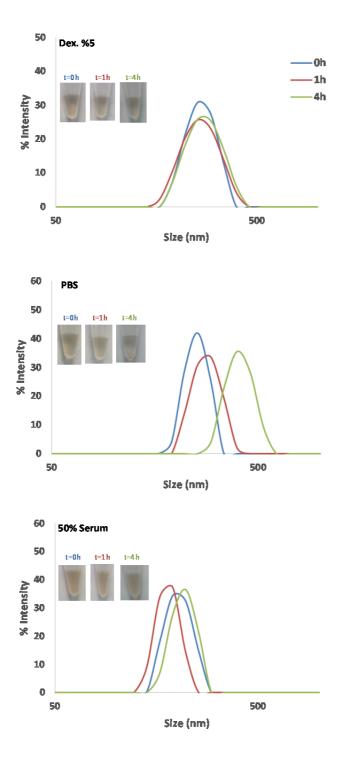
**Supporting Figure 1:** AFM of (A) GO and (B) GO-DOTA. (i) AFM images; (ii) lateral size distribution measured by counting the lateral dimension; (iii) thickness distribution obtained from height sections. Percent counts were obtained by counting more than 100 sheets obtained from several images per sample. Scale bars are  $1\mu m$ .



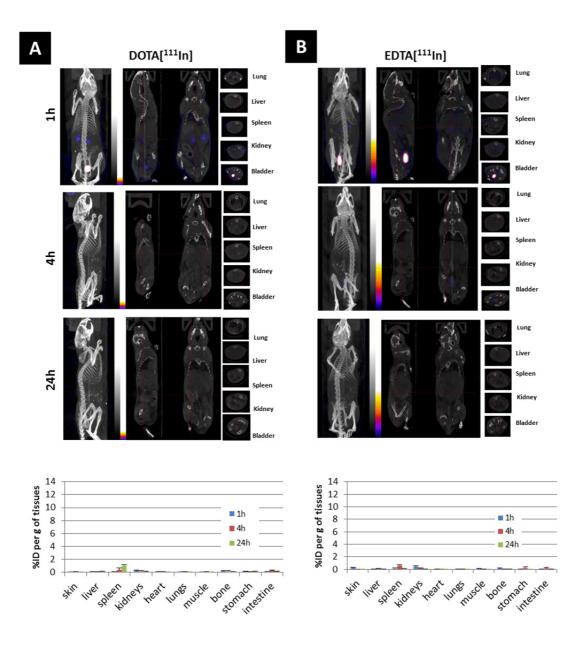
**Supporting Figure 2:** XPS of GO, GO-NH<sub>2</sub> and GO-DOTA showing the appearance of the N (1s) peak at ~400 eV, compared to XPS of GO.



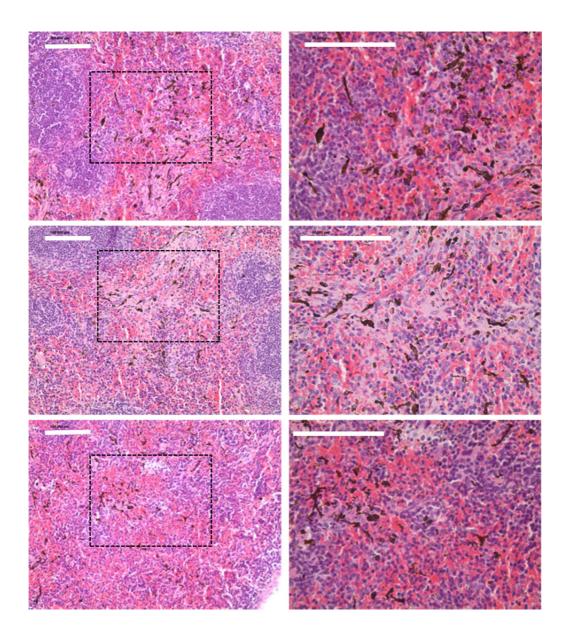
**Supporting Figure 3:** Thin layer chromatography of radiolabeled constructs showing: (A) labeling efficiency after radiolabeling with <sup>111</sup>In of the chemically conjugated (GO-DOTA) compared to the physically adsorbed control (GO + DOTA); (B) removal of free <sup>111</sup>In from GO-DOTA[<sup>111</sup>In] sample by centrifugation for 30 min compared to the physically adsorbed control (GO + DOTA]; (C) no free (unbound) <sup>111</sup>In was detected before injecting samples in mice; (D) DOTA conjugation efficiency detected by using a modified running buffer at pH 9; (E) radiolabeling stability up to 24h of the chemically conjugated sample (GO-DOTA[<sup>111</sup>In]) compared to the physically adsorbed control (GO + DOTA[<sup>111</sup>In]) at 37°C, in both 50% serum and PBS.



**Supporting Figure 4:** Colloidal stability of the GO-DOTA sheets dispersed in Dextrose, PBS and serum (50%) (from top to bottom) for t=0, 1h and 4h. The mean size of the dispersed material was measured using dynamic light scattering using Malvern Zetasizer Nano ZS (UK).



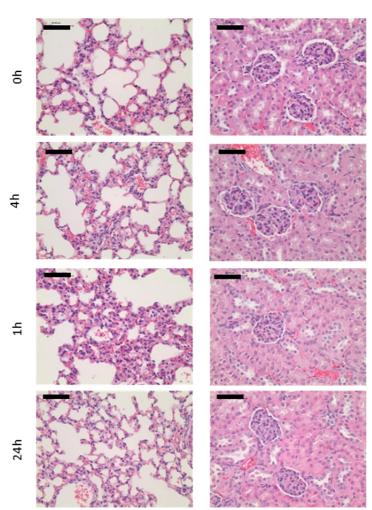
**Supporting Figure 5**: Biodistribution of **(A)** DOTA[<sup>111</sup>In] and **(B)** EDTA[<sup>111</sup>In]. Whole body SPECT/CT imaging of a C57BL/6 mouse injected with DOTA[<sup>111</sup>In] and EDTA[<sup>111</sup>In], respectively, imaged at different time points (1, 4, 24 h) showing from left to right whole body, sagittal, coronal and transverse views. Gamma scintigraphy bar charts showing organ biodistribution (n=4/group).



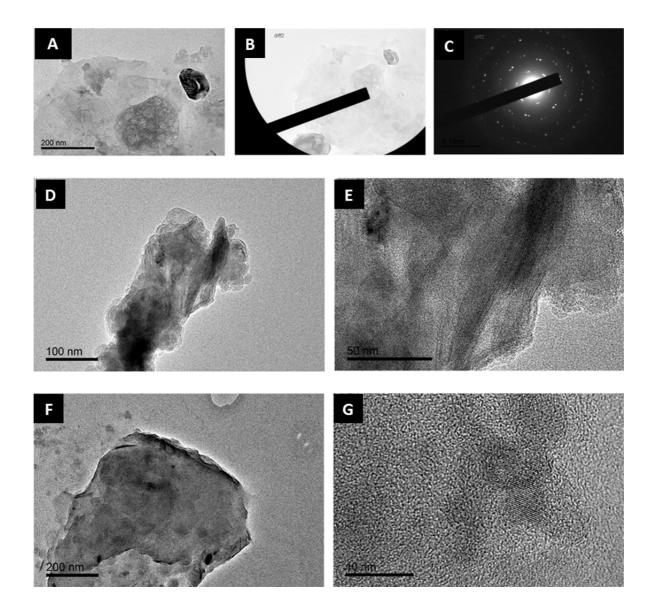
**Supporting Figure 6:** Histology using H & E staining of spleen sections showing GO-DOTA accumulation at 24h post-injection. Right hand-side images are higher magnification images of the boxes on the left hand-side sections. All scale bars are 100  $\mu$ m.



Kidney



**Supporting Figure 7:** Histology using H & E staining of lung (left) and kidney (right) tissue sections from mice injected with 50  $\mu$ g of GO-DOTA chelated with non-radioactive InCl<sub>3</sub> after 1, 4 and 24h compared to uninjected mice at time 0h. All scale bars are 50  $\mu$ m.



**Supporting Figure 8:** HR-TEM of GO-DOTA found in the urine after 24h **(A)** and their corresponding SAED diffraction patterns **(B, C)**. Other HR-TEM images of GO-DOTA found in the urine **(D, E)** and corresponding magnification images showing ordered areas **(F, G)**.