# Figure S1

Control of anti Nm-fHbp, anti-NHBA, anti-LPS and anti-Mbp antibody specificity on *E.coli* BL21*ΔompA* recipient strain. Bacterial cells from BL21*ΔompA*(pET) strain were fixed with 2% formaldehyde solution and incubated with specific antibodies in the presence (lower panels) or absence (upper panels) of membrane permeabilization by 0,1% Brij96. Anti Nm-fHbp or anti-NHBA antibody binding on the recipient strain was followed by incubating cells with alexa fluor 594-labelled anti-rabbit antibodies (red), while anti-LPS and anti-Mbp antibody binding was followed using alexa fluor 488-labelled anti-mouse antibody (green). In A, B, C, each right panel represents a magnification of the corresponding left panel. **A**) Cells were incubated with rabbit anti-Nm-fHbp and mouse anti-LPS; **B**) Cells were incubated antibodies rabbit anti-NHBA and mouse anti-LPS antibodies; **C**) Cells were incubated and mouse anti-Mbp antibody.

### Figure S2

Assessment of Nm-fHbp and NHBA localization by FACS analysis and confocal microscopy in BL21 WT strain. **A)** After IPTG induction bacterial cells from BL21(pET) (red histograms), BL21(pET\_Nm-fHbp) and BL21(pET\_NHBA) (blue histograms) strains were incubated first with anti-Nm-fHbp and anti-NHBA specific antibodies, respectively, and subsequently with FITC-labelled anti-rabbit secondary antibodies. Fluorescence was measured by Fluorescence-activated Cell Sorting (FACS). **B)** Bacterial cells from BL21 (pET\_Nm-fHbp) (left) and BL21(pET\_NHBA) (right) strains were fixed with 2% formaldehyde solution and incubated with anti-Nm-fHbp and anti-NHBA antibodies and with a mouse

monoclonal antibody specific for the core region of LPS. To analyze an intracellular staining samples were first treated with 0,1% Brj96 5 min at room temperature RT (+brij96). The binding of anti-Nm-fHbp and anti-NHBA antibodies was visualized with alexa fluor 594-labelled anti-rabbit antibodies (red), while anti-LPS antibody binding was followed using alexa fluor 488-labelled anti-mouse antibody (green).

## Figure S3

Assessment of localization of Nm-fHbp expressed in *E. coli* BL21-C43 strain by FACS analysis and confocal microscopy. A) SDS-PAGE analysis of total cell extracts from BL21-C43 and BL21-C43 (pET Nm-fHbp) strains. B) BL21-C43 (pET Nm-fHbp) (blue histogram) and BL21-C43 (pET) (red histogram) strains were incubated first with rabbit anti-Nm-fHbp specific antibody, and subsequently with FITC-labelled anti-rabbit secondary antibodies. Fluorescence was measured by Fluorescence-activated Cell Sorting (FACS). C) Bacterial cells from BL21-C43 (pET Nm-fHbp) strain were fixed with 2% formaldehyde solution and incubated with rabbit anti-Nm-fHbp antibody in the presence of a mouse monoclonal antibody specific for the core region of LPS in the presence (lower panels) or absence (upper panels) of permeabilization treatment with 0,1% Brij96. The binding of anti-Nm-fHbp antibody was visualized with alexa fluor 594-labelled anti-rabbit antibodies (red), while anti-LPS antibody binding was followed using alexa fluor 488-labelled anti-mouse antibody (green). Each right panel represents a magnification of the corresponding left panel.

### Figure S4

Effect of Slam1 on surface localization of Nm-fHbp. A) BL21 A ompA, BL21 WT and BL21-C43 strains transformed with pET Nm-fHbp, pET Nm-fHbp+pACYC and pET Nm-fHbp+pACYC slam1 plasmids were grown in LB at 37°C. At OD<sub>600</sub> = 0.5, 1 mM IPTG was added and after 2 hours bacterial cells were collected by centrifugation and incubated with anti-Nm-fHbp specific antibodies and subsequently with FITC-labelled anti-rabbit secondary antibodies. Fluorescence was measured bv Fluorescence-activated Cell Sorting (FACS). B) BL21*∆ompA*(pET\_Nm-fHbp)(pACYC) BL21 *AompA*(pET Nmand (pACYC slam1) strains were grown as described above and the binding of anti-Nm-fHbp antibodies was followed by FACS in the presence or absence of different concentration of either fHbp-derived peptide<sub>35-50</sub> or NHBA-derived peptide<sub>408-421</sub>

### Figure S5

Effect of Slam1 on surface localization of NHBA in BL21*∆ompA*. Bacterial cells co-transformed with either pET\_NHBA + pACYC or pET\_NHBA + pACYC\_slam1 plasmids were grown in LB at 37°C. At OD<sub>600</sub> = 0.5, 1 mM IPTG was added and after 2 hours bacterial cells were collected by centrifugation and incubated with anti-NHBA specific rabbit antibodies and subsequently with FITC-labelled anti-rabbit secondary antibodies. Fluorescence was measured by Fluorescence-activated Cell Sorting (FACS).