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# Supplementary Materials for

# Stress-induced dynamic regulation of mitochondrial STAT3 and its association with cyclophilin D reduce mitochondrial ROS production

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Fig. S1. Ser<sup>727</sup> and Tyr<sup>705</sup> of STAT3 are not required for H<sub>2</sub>O<sub>2</sub>-induced mitoSTAT3 loss. 4T1 or wild-type (WT) MEF cytosolic extracts were probed for pTyr<sup>705</sup> (**A**) or (**B**) pSer<sup>727</sup> STAT3. Blots are representative of 2 independent experiments. (**C**) 4T1 cells untreated or treated for 30' with H<sub>2</sub>O<sub>2</sub> were collected and nuclear extracts were immunoblotted for STAT3. Blots are representative of 2 independent experiments. (**D**) Mitochondrial lysates were probed for pSer<sup>727</sup> STAT3 and total STAT3 abundance and quantified by densitometry analysis. \*p=0.0229. Blots are representative of 4 independent experiments. (**E**) *STAT3*<sup>-/-</sup> MEFs were reconstituted with WT STAT3 (STAT3a) or Ser<sup>727</sup> (S727A and S727D) and Tyr<sup>705</sup> (Y705F) mutants. Mitochondrial extracts from untreated cells or cells treated with H<sub>2</sub>O<sub>2</sub> for 30' were immunoblotted for STAT3 or NDUFA9

(loading control). Cytosolic amounts of STAT3 Ser<sup>727</sup> and Tyr<sup>705</sup> mutants are shown as a control (right panel). Blots are representative of 2 independent experiments. (**F**) Mitochondrial extracts from WT MEFs untreated or treated for 30' with  $H_2O_2$  were probed with multiple STAT3 antibodies and GRIM19 (loading control). D3Z2G (Cell Signaling, rabbit monoclonal, C-terminus), 79D7 (Cell signaling, rabbit monoclonal, C-terminus), 124H6 (Cell Signaling, mouse monoclonal, C-terminus), N-term Ab (BD Biosciences, mouse monoclonal, N-terminus). Blots are representative of 2 independent experiments. (**G**) Mitochondria were isolated from HeLa cells treated for the indicated times with  $H_2O_2$  and lysates were resolved by SDS-PAGE and blotted for STAT3. Blots are representative of 2 independent experiments.



**Fig. S2. Selectivity of the mitoSTAT3 signaling pathway and relevance in vivo.** (**A**) BT474 [ER<sup>-</sup>/PR<sup>-</sup>/HER2<sup>+</sup>] human breast cancer cells were treated with OSM for the indicated times and mitochondrial extracts were probed for STAT3. Blots are representative of 2 independent experiments. (**B**) Mitochondria from WT MEFs treated with OSM for the indicated times were isolated and extracts immunoblotted for STAT3. Blots are representative of 2 independent experiments. (**C**) WT MEFs were treated with Buthionine Sulfoximine (BSO) for the indicated times and mitochondrial extracts were immunoblotted for mitoSTAT3. Blots are representative of 2 independent experiments. (**C**) WT MEFs were treated with Buthionine Sulfoximine (BSO) for the indicated times and mitochondrial extracts were immunoblotted for mitoSTAT3. Blots are representative of 2 independent experiments. (**D**) WT MEFs were treated for the indicated times with IFNβ and immunoblotted for STAT3 (mito fraction, top panel) or pTyr<sup>701</sup> STAT1 and pTyr<sup>705</sup> STAT3 (cyto fraction, bottom panel). Blots are representative of 2 independent experiments. (**E**) Mitochondrial extracts from MDA-231 cells were treated with IFNγ and mitochondrial fractions were

probed for STAT3 (top panel). Cytosolic extracts (bottom panel) were probed for pTyr<sup>701</sup> STAT1, Blots are representative of 2 independent experiments. (**F**) Mitochondrial extracts from WT MEFs (top panel) or MDA-231 cells (bottom panel) were treated with epidermal growth factor (EGF) and immunoblotted for STAT3. Blots are representative of 2 independent experiments. (**G**) 8-12 week old CD1 male mice were tail vein injected with either PBS or IL-6 ( $500\mu g/kg$ ) and liver mitochondrial (left panel) or cytosolic fractions (right panel) were immunoblotted for STAT3 and pTyr<sup>705</sup> STAT3 respectively. A non-injected control liver extract is presented in Lane 1. Blots are representative of 2 independent experiments.



Fig. S3. Inhibition of relevant kinase pathways does not affect stimulation-induced decreases in mitoSTAT3. (A) WT MEFs were treated with IL-6 either in the presence or absence of Ruxolitinib and mitochondrial and cytosolic extracts were evaluated by western blotting. Blots are representative of 2 independent experiments. (B) MDA-231 cells were pre-treated with the STAT3 inhibitor cryptotanshinone for 1H prior to OSM stimulation (5' treatment). Mitochondrial and cytosolic extracts were immunoblotted for STAT3 and pTyr<sup>705</sup> STAT3 respectively. Blots are representative of 2 independent experiments of 2 independent experiments. (C) WT MEFs were incubated with Staurosporine or H7 Dihydrochloride 30' prior to H<sub>2</sub>O<sub>2</sub> treatment and mitochondrial (top panel) and cytosolic (bottom panel) fractions were immunoblotted. Blots are representative of 2 independent experiments. (D to G) WT MEFs or MDA-231 cells were pre-incubated with inhibitors to p38 MAPK (D, SB203580), PI3K (E, LY294002), mTOR (F, Torin 1), or JNK (G, SP600125) and

treated with either OSM (5') or  $H_2O_2$  (30'). Mitochondrial and/or cytosolic extracts were probed as indicated. Blots are representative of 2 independent experiments. (**H**) MCF-7 cells were treated with IL-6 for the indicated times and mitochondrial extracts were probed for STAT3, pERK1/2, or NDUFA9 (left panel). Comparison of mitochondrial STAT3 amounts in MCF-7 compared to MDA-231 cells (right panel). Blots are representative of 3 independent experiments.



Fig. S4. Inhibition of mitoproteases does not affect proteolysis of mitoSTAT3. (A) WT MEFs were pretreated with the calpain inhibitor MDL-28170 and subjected to  $H_2O_2$ treatment for 30'. Mitochondrial extracts were probed for STAT3. Blots are representative of 2 independent experiments. (B) Mitochondrial lysates from CDDO-Me pre-treated WT MEFs were subjected to western blotting following treatment with  $H_2O_2$ for 30'. Blots are representative of 2 independent experiments. (C)  $ClpP^{+/+}$  or  $ClpP^{-/-}$ MEFs were treated with either OSM or  $H_2O_2$  and mitochondrial and cytosolic fractions were probed for STAT3, Actin, ClpP, and CypD. Blots are representative of 2 independent experiments. (D) WT MEFs were treated with cycloheximide for 2H prior to  $H_2O_2$  stimulation and isolated mitochondrial extracts were immunoblotted. Blots are representative of 2 independent experiments. (E) Cytosolic extracts from MDA-231 cells treated with or without cycloheximide were stimulated with OSM or  $H_2O_2$  and subjected to western blotting. Blots are representative of 3 independent experiments.



Fig. S5. mitoSTAT3 inducibly binds to CypD after H<sub>2</sub>O<sub>2</sub> or cytokine stimulation. (A) Mitochondrial lysates from  $STAT3^{+/+}$  or  $STAT3^{-/-}$  MEFs untreated or treated with H<sub>2</sub>O<sub>2</sub> were incubated with GST-CypD in the presence or absence of CsA and probed for STAT3. Input is shown in bottom panel. Blots are representative of 2 independent experiments. (B) MDA-231 [ER<sup>-</sup>/PR<sup>-</sup>/HER2<sup>-</sup>] or MDA-435 [ER<sup>-</sup>/PR<sup>-</sup>/HER2<sup>+</sup>] human breast cancer cells were treated for 4H with H<sub>2</sub>O<sub>2</sub> and purified mitochondria were subjected to GST-CypD pulldown (top panel). Lower panels show the input (10% of that used for the pull downs). Blots are representative of 2 independent experiments. (C) Immunoprecipitation of STAT3 or control IgG from purified mitochondria from MDA-231 cells untreated or treated for 30' with H<sub>2</sub>O<sub>2</sub> and probed for STAT3 and CypD. Lower panels show the input (10% of that used for the immunoprecipitation) Blots are representative of 2 independent experiments. (**D** to **F**) GST-CypD pulldown of STAT3 from mitochondrial extracts following OSM treatment for the indicated times in MDA-231 cells (**D**), 4T1 cells (**E**, input presented in Fig. 2C), and in *STAT3<sup>-/-</sup>* MEFs reconstituted with WT STAT3 (**F**). Blots are representative of 2 independent experiments.



Fig. S6. Ser<sup>727</sup> is dispensable for the mitoSTAT3-CypD interaction. (A) Expression of Flag-STAT3 WT or -S727A STAT3 in 293T cells was followed by H<sub>2</sub>O<sub>2</sub> treatment and resulting whole cell extracts were incubated with GST-CypD and probed for Flag. Coomassie stain of GST-CypD input is shown. Blots are representative of 2 independent experiments. (B) 4T1 cells expressing the mutant STAT3 isoforms Y705F/S727A (YFSA) or Y705F/S727D (YFSD) were treated with H<sub>2</sub>O<sub>2</sub> and STAT3 was immunoprecipitated from mitochondrial extracts and probed for CypD. Blots are representative of 2 independent experiments. (C) *STAT3*<sup>-/-</sup> MEFs were transduced with retroviral constructs to drive expression of WT STAT3 (STAT3 $\alpha$ ) or the following STAT3 mutant constructs: S727A, Y705F, or S727D. Immunoblots of mitoSTAT3 following GST-CypD Pulldown of mitochondrial extracts isolated from cells after 30' H<sub>2</sub>O<sub>2</sub> treatment. Input as presented in fig. S1E. Blots are representative of 2 biological replicates. (D) *STAT3*<sup>-/-</sup> MEFs expressing STAT3 $\beta$ , a C-terminal truncation mutant that lacks the Ser<sup>727</sup> site, were treated with H<sub>2</sub>O<sub>2</sub> (30') and subjected to GST-CypD pulldown with blotting for STAT3. Blots are representative of 2 independent experiments. (**E**) Mitochondria from WT MEFs treated for 30' with H<sub>2</sub>O<sub>2</sub> were incubated with GST-CypD and probed for mitochondrial STAT1. Blots are representative of 2 independent experiments. (**F**)  $CypD^{+/+}$  MEFs and  $CypD^{-/-}$  MEFs were treated for the indicated times with OSM and isolated mitochondrial lysates were immunoblotted for STAT3, pERK1/2, NDUFA9 (loading control) and CypD. Blots are representative of 2 independent experiments. (**G**) WT MEFs were pre-treated with CsA (4H) then treated with H<sub>2</sub>O<sub>2</sub> for the indicated times and mitochondrial lysates were probed for STAT3. Blots are representative of 2 independent