Effects of Near Ultraviolet and Green Radiations on Plant Growth^{1, 2}

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In an earlier paper, Klein (7) reported that the growth of nonphytosynthetic plant tissue cultures was reversibly repressed by near-ultraviolet (near-UV) and green radiations. The present paper reports our studies on the effects of these wavelengths on the growth and respiration of higher plants, on algae, and on a fungus.

Materials and Methods

Test Systems. Tagetes erecta L., Dwarf Pigmy French Marigold (Burpee 4166) was planted in soil in polyethylene boxes; radiation treatments were started immediately. Impatiens Balsamina L., Holsti Hybrid (Burpee 4107) and Lycopersicon esculentum Mill., Rutgers (Burpee 5061) were similarly grown; radiation treatments were started when the seedlings were 1 cm tall. One week after planting, the seedlings were thinned for uniformity. Day temperatures ranged from 23° to 26° and dark temperatures ranged from 19° to 23°. Experiments were terminated 1 month after planting. Experiments were repeated at least once.

Chlamydomonas reinhardti Dang. was grown in Trainor's I-N medium (19) at 20° , cells were washed 3 to 4 times in water and used to inoculate 10 ml of medium in 50 ml flasks. Five cultures for each variable were incubated on a rotary shaker at 100 cycles per minute for 96 hours at 20°. Cell density was measured in a Klett colorimeter with a green 54 filter.

Euglena gracilis bacillaris Klebs was shaken under white light at 20° as either a nongrowing culture in buffer (1) or as a growing culture in medium (9) and O_2 uptake measured manometrically at 30° in water 1, 2, and 3 days after inoculation. Other cultures were grown in darkness at 20° on a rotary shaker, the cells washed by centrifugation and then incubated in buffer or in medium for up to 72 hours in light. Chlorophylls and carotenoids were determined (18).

Sordaria fimicola Cesati and Notarius was grown in darkness at 25° in petri dishes containing Difco cornmeal agar plus 1% glucose. Disks of agar and mycelium were removed with a no. 2 corkborer and 1 disk transferred to each racer tube (17) containing the same medium. Linear growth was measured daily for 7 to 9 days. Increase of weight of mycelium was measured in 50 ml of liquid medium (5) in 125 ml flasks inoculated with agar disks of mycelium. Six tubes or flasks were used for each variable.

Radiation Procedures. White light was supplied from banks of cool-white fluorescent and incandescent lamps in a 10:1 w ratio. Marigold plants received 1000 ft-c and the balsam and tomato plants received 500 ft-c, as measured with a Weston cosine-corrected illumination meter. The photoperiod was 15 hours for the higher plants and 24 hours for the microorganisms. Supplementary near-UV radiation was supplied from integral filter black-light fluorescent lamps emitting 70 % of their radiation between 355 and 380 m μ . Supplementary green radiation (530– 585 m μ , peak at 565 m μ) was supplied from filtered green fluorescent lamps (8).

Near-UV radiations were completely removed from white light with plastic filter with a low wavelength cut-off at 385 m μ and 95 % transmission above 400 m μ (8). The green component of white light was reduced approximately 50 % with a Cinemoid no. 36 pale lavender plastic filter. Appropriate adjustment of plant to lamp distance was made to equalize total radiant energy in μ w/cm² received by the plants (fig 1).

Results

Growth of Intact Higher Plants. Removal of all of the near-UV or half of the green radiation from white light increased the fresh and dry weight, height of the plants, and the lengths of the peduncle of marigold plants (fig 2a, table I). Peduncle weight and length were reduced when either near-UV or green wavelengths were filtered but were increased above control levels when both wavelengths were filtered. Tomato plants showed a similar response (fig 2b) as did corn and bean plants. Since marigold, tomato, and corn are semi-tropical. highlight requiring species, identical experiments were performed with Impatiens, a temperate zone, semishade plant. Selective removal of near-UV and/or green wavelengths from white light caused growth responses similar to those observed in marigold (fig 2c). Thus, monocot and dicot species from widely separated families are responsive to selective removal of these wavelengths.

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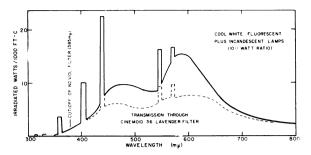


FIG. 1. Spectral energy distribution curves of light bank containing fluorescent and incandescent lamps with (dashed line) and without (solid line) plastic filters.

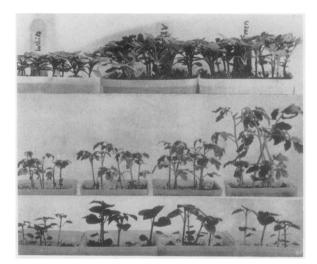


FIG. 2. Effects of selective removal from white light of near-UV and green wavelengths on growth of plants. A, marigold plants 21 days from seed showing (left to right) full white light, white light minus near-UV, and white light minus green component. B, tomato and C, *Impatiens* plants 21 days from seed showing (left to right) full white light, white light minus near-UV, white light minus green, and white light minus both near-UV and green components.

Addition of supplementary near-UV and/or green wavelengths to white light decreased the fresh weight of vegetative and reproductive structures of marigold plants but did not affect the dry weight or height of the plants (table I).

Growth of Chlamydomonas. Cell multiplication in this green alga was increased by selective removal of the near-UV component of 300 and 700 ft-c (but not 1500 ft-c) of white light, but selective reduction in the green component affected cell number only at 300 ft-c of white light (table II). The removal of both near-UV and green wavelengths was without effect.

Table II. Effect of Removal of Near-UV or Green Wavelengths from White Light on the Growth of Cultures of Chlamydomonas

Population density was measured with Klett colorimeter after 96 hours at 20°.

| Ft-c white | Control | -UV | -G | -UV, -G | |
|---------------|--------------|--------------|--------------|--------------|--|
| 1500 | 190 ± 14 | 180 ± 16 | 210 ± 20 | 190 ± 12 | |
| 700 | 313 ± 26 | 405 ± 30 | 311 ± 24 | 311 ± 27 | |
| 300 | 150 ± 10 | 208 ± 16 | 183 ± 14 | 166 ± 11 | |

Growth of Sordaria. Like some other fungi (4, 6, 23) Sordaria grew more rapidly in the presence than in the absence of white light and the light effect (found to be at nor near 420 m μ) was virtually independent of light intensity. Selective filtration of the near-UV or green wavelengths increased the linear extension of the mycelium but removal of both near-UV and green wavelengths was no more effective than removal of either near-UV or green light (table III). Dry weight of mycelium followed a similar but not identical pattern. In the absence of white light, near-UV and/or green radiations supplied at 100 μ w/cm² was without effect on dry weight of liquid cultures, but near-UV plus green light repressed the linear extension of agar cultures.

Metabolic Studies. Although most studies indicate that light has little or no direct effect on respiration (16, 22), green radiations have been reported

Table I. Effects of Near-UV (UV) and Green (G) Radiations on Growth of Marigold Plants Average data per plant of white light control are given as weights or lengths with standard errors; all other values are given as percentages of white light controls.

| | White light | White light minus : | | | White light plus: | | |
|------------------------|-------------------|---------------------|-----|--------|-------------------|-----|--------|
| | control | UV | Ğ | UV + G | UV | G | UV + G |
| Vegetative height (mm) | 41.1 ± 3.7 | 141 | 156 | 144 | 97 | 80 | 89 |
| Fr wt (g) | 0.648 ± 0.057 | 142 | 126 | 123 | 73 | 61 | 69 |
| Dry wt (g) | 0.061 | 136 | 130 | 126 | 98 | 73 | 88 |
| No. flower buds | 20.0 ± 2.1 | 190 | 135 | 135 | 112 | 100 | 120 |
| Length peduncle (mm) | 13.0 ± 1.2 | 112 | 104 | 121 | 74 | 76 | 48 |
| Fr wt peduncle (g) | 0.058 ± 0.006 | 59 | 68 | 78 | 70 | 78 | 52 |
| Dry wt peduncle (g) | 0.005 | 77 | 52 | 127 | 58 | 64 | 39 |

Table III. Effect of Removal of Near-UV (UV) or Green (G) Radiation from White Light on the Growth of Sordaria

Linear extension was measured daily for 9 days on agar medium. Mycelial dry weight was determined after 10 days in liquid medium. Dark control linear extension = 12.5 ± 0.2 mm per day; dry weight = 132.9 mg.

| Intensity (ft-c) | White | White -UV | White -G | White -UV -G | | |
|---------------------|---------------------------|----------------|----------------|-----------------|--|--|
| | Linear extension (mm/day) | | | | | |
| 1500 | 17.7 ± 0.2 | 21.5 ± 0.4 | 18.2 ± 0.1 | 21.5 ± 0.3 | | |
| 700 | 17.6 ± 0.4 | 17.7 ± 0.6 | 18.0 ± 0.4 | 18.0 ± 0.8 | | |
| 300 | 16.2 ± 0.4 | 15.9 ± 0.1 | 15.8 ± 0.9 | 15.8 ± 0.5 | | |
| | Dry wt (mg in 10 days) | | | | | |
| 700 | 163.2 | 204.7 | 165.2 | 216.6 | | |

to suppress the respiration of yeast cells (12, 15), the cytochrome oxidase activity and oxidation of organic acids of cauliflower mitochondria (13) and the malonate sensitive respiration of Euphorbia bracts (14). The oxidation of marigold leaf tissues from plants grown under white light supplemented with near-UV or green radiations for 21 days did not differ from that from plants grown under white light. Neither filtration or the near-UV or green wavelengths from white lights nor supplementary near-UV or green radiations to white light had any effect on the endogenous O2 uptake of stationary or growing cultures of Euglena. No changes were observed in the cytochrome oxidase activity of cauliflower mitochondria measured spectrophotometrically at 550 m_{μ} when the mitochondria isolated by the method of Laties (11) were carefully washed to remove endogenous reducing compounds. Mitochondria isolated from bean stems, Rumex tissue cultures, and cauliflower buds were used for manometric succinoxidase measurements. Light did not cause any diminution in succinoxidase activity of washed mitochondria; reductions in activity were noted only in unwashed preparations (table IV). We conclude that the observed growth changes caused by near-UV and green light are unlikely to be due to alterations in energy metabolism as determined by O₂ uptake.

There was no repression in the rates of synthesis of chlorophylls or carotenoids by Euglena cells when the near-UV or green components of the white light were filtered or when the white light was supplemented with these radiations.

Discussion

Near-UV and green wavelengths are capable of suppressing the growth of plants which also receive adequate levels of those wavelengths necessary for photosynthesis and for normal development (10, 20). Vince et al. (21) obtained reductions in leaf number, internode length, and a delay in flower induction of carnation and lettuce when white light was supplemented with green wavelengths. We have also found that the growth of intact higher plants, a green alga, a fungus, and tissue cultures is promoted by selective removal of these wavelengths from white light. Boney and Corner (2,3) stimulated the growth of sporelings of red algae by removing a fraction of the green component of white light, and similar observations have been made in our laboratory with a bacterium (H. H. Clum, unpublished). Although there is a pattern of similarity in the responses of these test systems, certain differences in detail have been found. In addition, some test systems do not respond to either removal or addition of near-UV or green wavelengths.

Our reported attempts to determine the reason for the observed growth effects are negative. Near-UV or green radiations do not appear to affect chlorophyll or carotenoid synthesis, respiration, or mitochondrial oxidations. Investigations of the mechanisms(s) are in progress.

Summary

Selective removal of near ultraviolet and green wavelengths from white light permitted enhanced growth of marigold, tomato, corn, and *Impatiens* plants, *Chlamydomonas* cells and the mycelium of *Sordaria*. Additions of near ultraviolet and green radiations caused repressions in the growth of marigold and *Sordaria*. These wavelengths do not alter the oxidative mechanisms of mitochondria, intact

 Table IV. Effect of Visible Radiation on the Oxidation of Succinate

 by Washed Cauliflower Mitochondria

| Expt | Light source | Spectrum | Total energy (mw/cm ²) | O ₂ uptake (µl/hr per vessel) | |
|------|-------------------------------|------------|---------------------------------------|---|-------|
| | | | | Dark | Light |
| 1–2 | GE Quartzline | White | 15 | 109 | 117 |
| 3 | Quartzline + 550 mµ filter | 530–585 mμ | 9 | 148 | 146 |
| 4* | Philips HPL | 550, 565 | 12 | 142 | 111 |
| 5–6 | Philips HPL | 550, 565 | 12 | 77 | 77 |

* Unwashed mitochondria.

algal cells or marigold leaf tissues. The capacity for chlorophyll and carotenoid synthesis by *Euglena* cells was unaffected by these wavelengths.

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Literature Cited

- 1. APP, A. A. AND A. T. JAGENDORF. 1963. Repression of chloroplast development in *Euglena gracilis* by substrates. J. Protozool. 10: 340-43.
- BONEY, A. D. AND E. D. S. CORNER. 1962. The effect of light on the growth of sporelings of the intertidal red algae, *Plumaria clegans* (Bonnem.) Schm. J. Marine Biol. Assoc. U.K. 42: 65-92.
- 3. BONEY, A. D. AND E. D. S. CORNER. 1963. The effect of light on the growth of sporelings of the red algae *Antithamnion plumula* and *Brongniartella* bysoides. J. Marine Biol. Assoc. U.K. 43: 319-25.
- CANTINO, E. C. AND E. A. HORENSTEIN. 1956. The stimulatory effect of light upon growth and CO₂ fixation in *Blastocladiella*. Mycologia 48: 777–83.
- 5. EL ANI, A. S. 1964. Self-sterile auxotrophs and their relation to heterothallism in *Sordaria fimicola*. Science 145: 1067–68.
- 6. GOLDSTEIN, S. 1963. Studies of a new species of *Thraustochytrium* that displays light stimulated growth. Mycologia 58: 799-811.
- KLEIN, R. M. 1964. Repression of tissue culture growth by visible and near visible radiation. Plant Physiol. 39: 546-39.
- 8. KLEIN, R. M. 1965. An inexpensive filter system for photomorphogenic research. Photochem. Photobiol. In press.
- KLEIN, R. M., M.-F. MORSELLI, AND J. WANSOR. 1963. Effect of ultraviolet radiation on growth of autotrophic and obligate heterotrophic cultures of *Euglena*. J. Protozool. 10: 223-25.

- KWACK, B. H. AND S. DUNN. 1961. Effect of light quality on plant maturity. I. Duration of growth nutrient supply and photoperiod. Lloydia 24: 75-80.
- 11. LATIES, G. G. 1953. The physical environment and oxidative and phosphorylative capacities of higher plant mitochondria. Plant Physiol. 28: 557-75.
- MATILE, P. 1962. Wirkung des sichtbaren Lichtes auf das Atmung von Hefe. Ber. Schweiz. Botan. Ges. 72: 236-61.
- MATILE, P. 1962. Lichthemmung der oxydation von Krebscyclus-Säuren durch Blumenkohl-mitochondrien. Experientia 18: 133-36.
- MATILE, P. 1962. Über die Licht-atmung der brakteen von Euphorbia pulcherrima. Planta 58: 193-98.
- MATILE, P. AND A. FREY-WYSSLING. 1962. Atmung and Wuchstum von Hefe im Licht. Planta 58: 154-63.
- ROSENSTOCK, G. AND A. RIED. 1960. Der Einfluss sichtbarer Strahlung auf die Pflanzenatmung. Handb. Pflanzenphysiol. 12 (2): 259-33.
- RYAN, F. J. 1950. Selected methods in Neurospora genetics. Methods Medical Research 3: 51-70.
- SMITH, J. H. C. AND A. BENITEZ. 1955. Chlorophylls: Analysis in plant materials. In: Modern Methods of Plant Analysis, Vol. IV. K. Paech and M. V. Tracey, eds. Springer-Verlag, Berlin. p 142–96.
- TRAINOR, F. R. 1958. Control of sexuality in Chlamydomonas chlamydogama. Am. J. Botany 45: 621-26.
- 20. VEEN, R. AND G. MEIJER. 1959. Light and Plant Growth. Philips Tech. Lab. Eindhoven.
- VINCE, D., J. BLAKE, AND R. SPENCER. 1964. Some effects of wavelength of the supplementary light on the photoperiodic behaviour of the long-day plants, carnation and lettuce. Physiol. Plantarum 17: 119-25.
- 22. WEINTRAUB, R. L. 1944. Radiation and plant respiration. Botan. Rev. 10: 384-459.
- 23. ZURZYCKA, A. 1963. Studies on photomorphosis Aspergillus giganteus mut. alba. III. Acta Biol. Cracovienaia. Ser. Botanica 6: 235-51.