

Title of file for HTML: Supplementary Information

Description: Supplementary Figures and Table.

Title of file for HTML: Supplementary Data 1

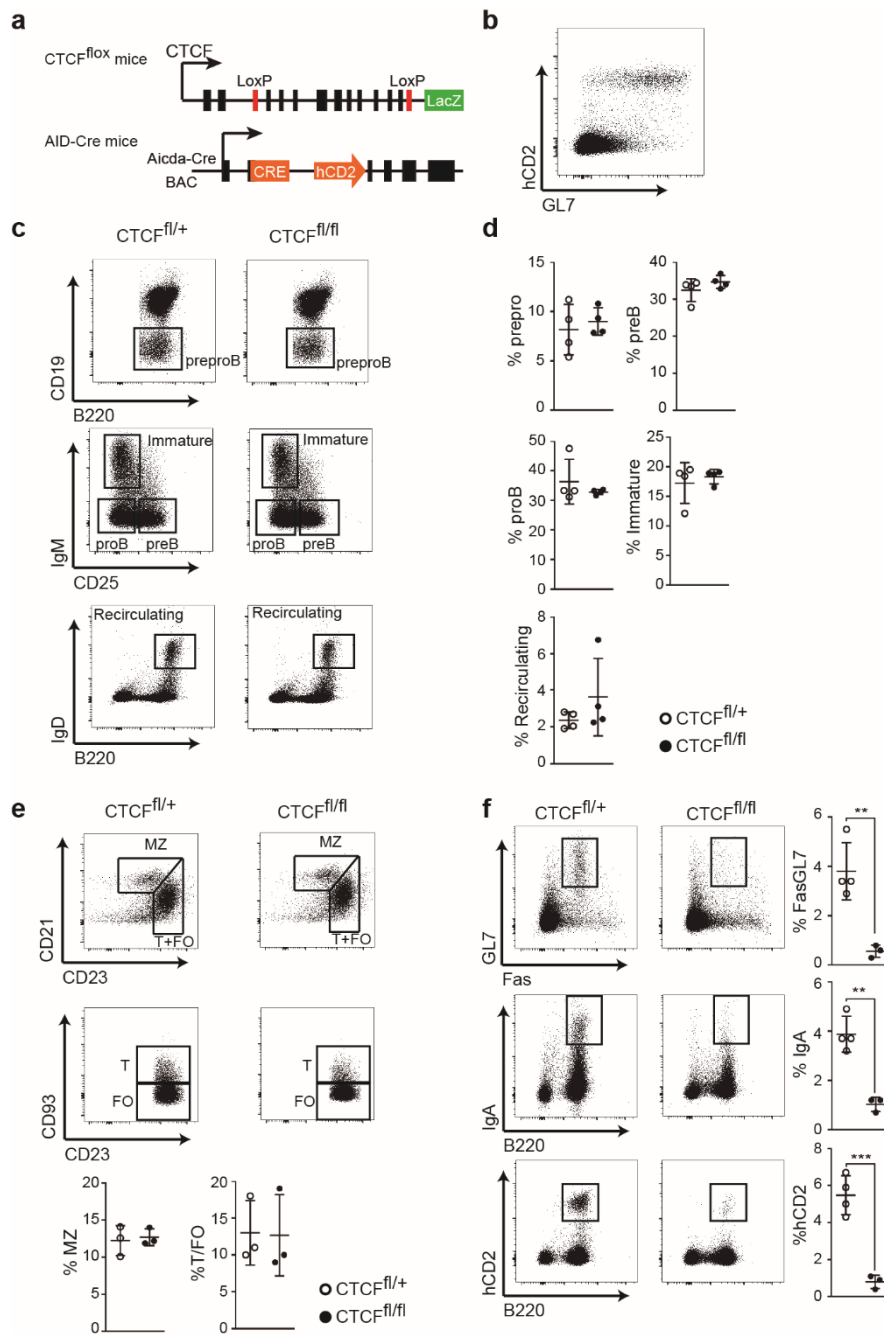
Description: List of genes differentially expressed between CTCFfl/fl and CTCFfl/+ hCD2+ sorted B cells stimulated for 48h in the presence of antiCD3/antiCD28 and T cells

Title of file for HTML: Supplementary Data 2

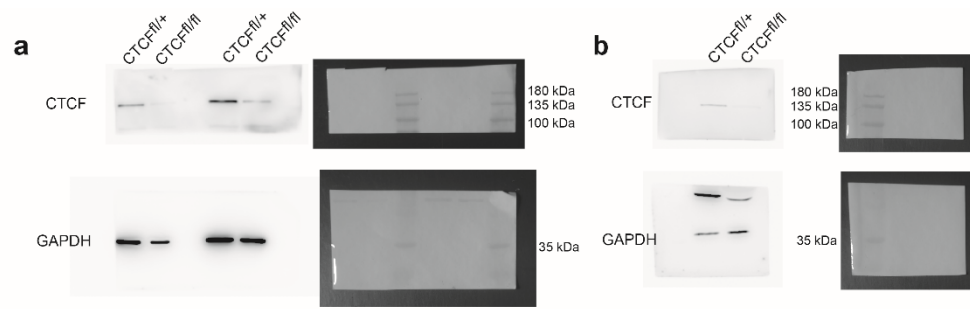
Description: List of genes differentially expressed between CTCFfl/fl and CTCFfl/+ hCD2+ sorted B cells stimulated for 48h in the presence of LPS/IL4

Title of file for HTML: Peer Review File

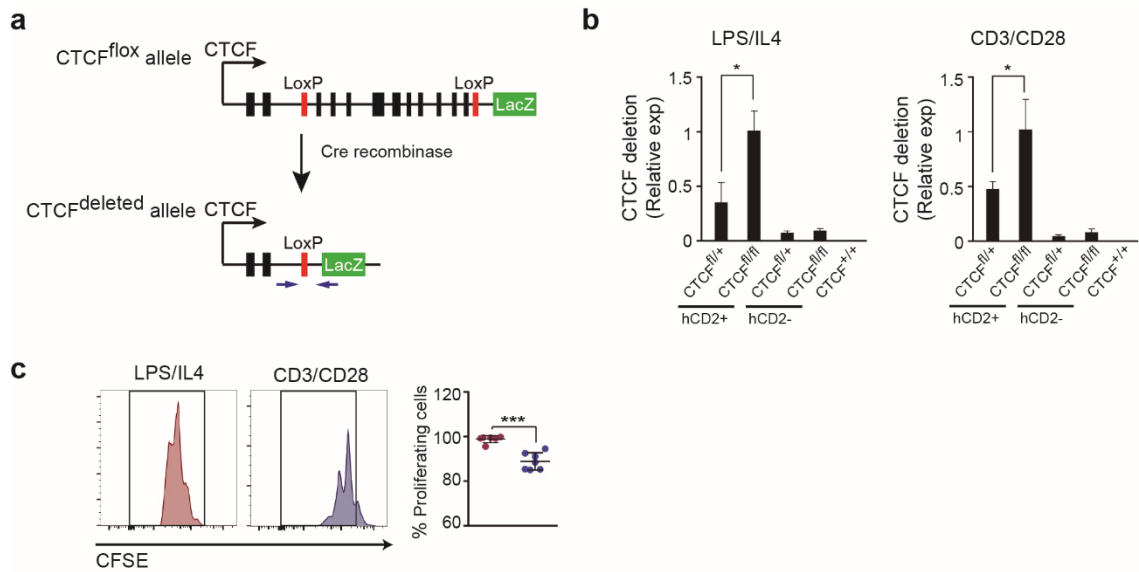
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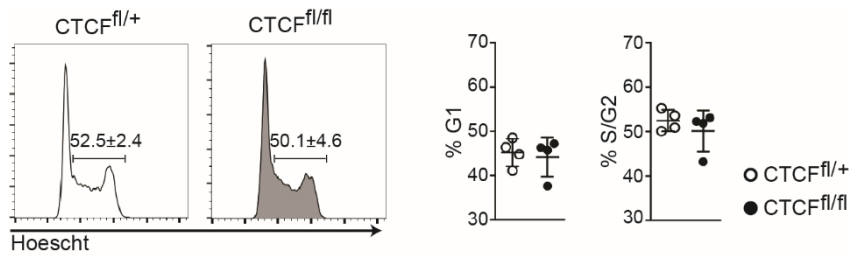
Supplementary Figure 1. B cell differentiation and constitutive GC in CTCF deficient mice. A) Representation of the constructs used for conditional depletion of CTCF in GC B cells. Mice carrying the CTCF allele flanked by LoxP sites (top) were bred to transgenic AID-CRE mice (bottom), where a cassette encoding for Cre recombinase and the hCD2 receptor is inserted in an AID-BAC. B) FACS analysis of hCD2 expression in Peyer's Patch B cells from CTCF^{fl/fl} mice. C) FACS analysis of B cell differentiation in bone marrow. Populations were defined as follows: preproB (B220⁺ CD19⁻); proB (B220⁺CD19⁺CD25⁻IgM⁻); preB (B220⁺CD19⁺CD25⁺IgM⁻); Immature (B220⁺CD19⁺CD25⁺IgM⁺); Recirculating (B220⁺CD19⁺IgD⁺). D) Quantification of the bone marrow populations shown in (C). Absolute numbers are shown in Supplementary Table 1. E) FACS analysis of B cell differentiation in spleen. B cell populations were defined as follows: Transitional (T: B220⁺CD21⁺CD23⁺⁺CD93⁻); Marginal Zone (MZ: B220⁺CD21⁺⁺ CD23⁺); Follicular (FO: B220⁺CD21⁺CD23⁺⁺CD93⁺). Quantifications are shown on the right. Absolute numbers are shown in Supplementary Table 1. All population percentages are referred to total B220⁺. CTCF^{fl/+}, white dots; CTCF^{fl/fl}, black dots. F) FACS analysis of GL7, Fas, IgA and hCD2 expression in Peyer's Patches B cells from CTCF^{fl/+} (n=4) and CTCF^{fl/fl} (n=3) mice. Plots are gated on B220⁺ cells (top) or on total live lymphocytes (middle and bottom). Quantifications are shown on the right, as percentages within B220⁺ cells. p(FasGL7)= 0.0056; p(IgA)= 0.0015; p(hCD2)= 0.0008. Statistical analysis was done by two-tailed unpaired Student's *t* test.



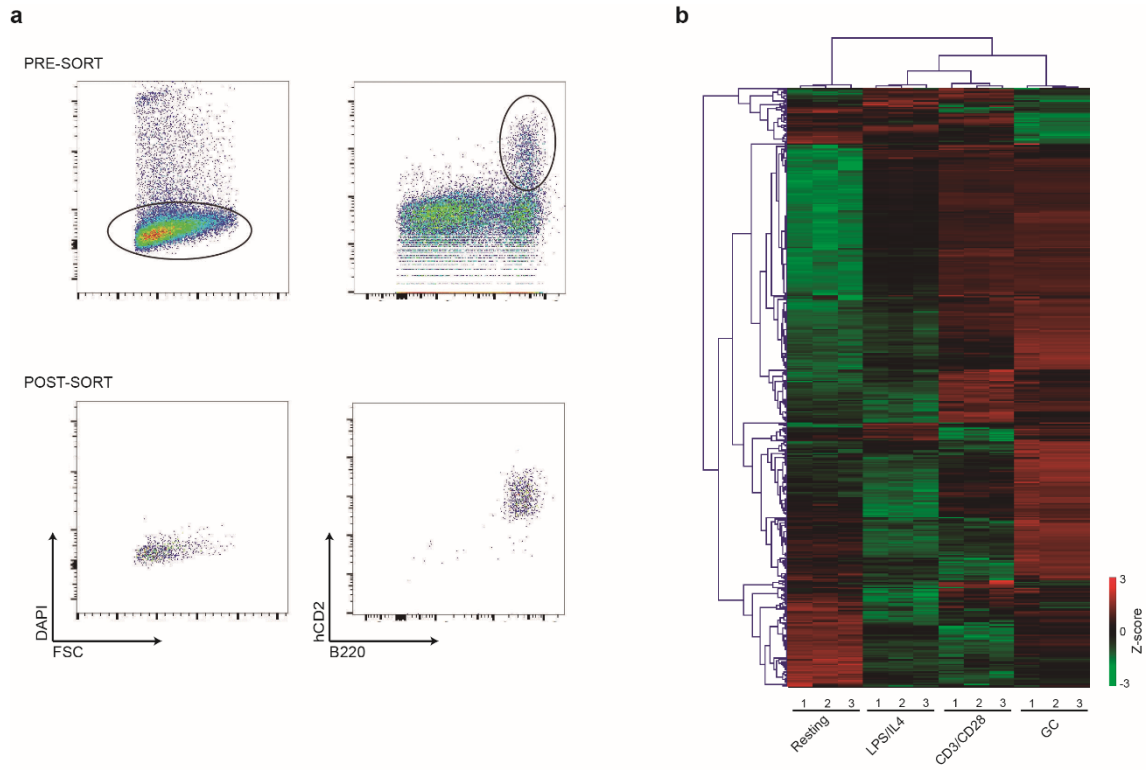
Supplementary Figure 2. Uncropped western blots. A) Western blot analysis of CTCF and GAPDH in isolated hCD2⁺ B cells from CTCF^{fl/+} and CTCF^{fl/fl} mice after 72 hours of LPS/IL4 stimulation. B) Western blot analysis of CTCF and GAPDH in isolated hCD2⁺ B cells from CTCF^{fl/+} and CTCF^{fl/fl} mice after 72 hours of stimulation in CD3/CD28 T-B co-cultures.



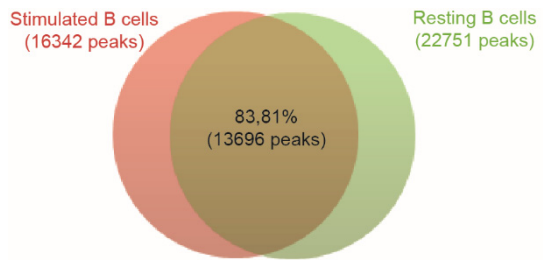
Supplementary Figure 3. Analysis of CTCF deletion and comparison of proliferation between LPS/IL4 and CD3/CD28 stimulated B cells. A) Representation of the strategy used for detection of CTCF^{fl} allele deletion by qPCR. Upper, CTCF^{fl} allele before Cre-mediated recombination. Lower, CTCF^{fl} allele after Cre-mediated recombination. Blue arrows represent primers used in the PCR. B) qPCR analysis of CTCF deletion in hCD2⁺ and hCD2⁻ cells from CTCF^{fl/+} (n=3) and CTCF^{fl/fl} (n=3) mice after 48h of stimulation with LPS/IL4 (right panel) or CD3/CD28 and T cells (left panel). p(LPS/IL4)= 0,0112; p(CD3/CD28)= 0,0283. C) CFSE staining of hCD2⁺ spleen B cells from CTCF^{fl/+} (n=5) and CTCF^{fl/fl} (n=7) mice after 72h of stimulation with LPS/IL4 or CD3/CD28 and T cells. Quantifications are shown on the right as percentages within B220⁺CD2⁺ cells. p<0.0001. Numbers indicate percentages \pm Standard Deviation. Statistical analysis was done by two-tailed unpaired Student's *t* test.



Supplementary Figure 4. RNA-seq analysis of LPS/IL4 and CD3/CD28 stimulated B cells. A) Gating strategy and post-sort analysis for selection of B cells used in RNA-seq experiments. CTCF^{fl/+} and CTCF^{fl/fl} B cells stimulated for 48h hours either in the presence of LPS/IL4 or in the presence of CD3/CD28 T stimulation were sorted as DAPI⁻, B220⁺, hCD2⁺ cells. Representative pre- and post-sort plots of B cells stimulated with CD3/CD28 T cells are shown. B) Heatmap analysis of RNA-seq data of cell cycle-related genes. Cell cycle genes (484) were chosen using GO Enrichment Analysis. Z-scores for resting, GC, LPS/IL4 and CD3/D28 stimulated B cells are represented. Clustering was performed using the average linkage method based on Pearson correlation distance. Each column shows an independent replicate.



Supplementary Figure 5. Cell cycle analysis in LPS/IL4 stimulated B cells. FACS cell cycle analysis of hCD2⁺ cells from CTCF^{fl/+} (n=4) and CTCF^{fl/fl} (n=4) mice after 48h of stimulation with LPS/IL4. Numbers indicate percentages \pm Standard Deviation. Quantification of G1 and S/G2 phase proportions is shown on the right.



Supplementary Figure 6. Comparison of CTCF binding in LPS/IL4 stimulated and naïve B cells. ChIP-seq analysis of CTCF binding peaks in stimulated B cells and resting B cells from GSE43594¹. Overlap of common peaks are depicted as Venn diagram.

Subset	Tissue	Cell number	
		CTCF ^{fl/+}	CTCF ^{fl/fl}
PreproB	Bone marrow ^a	0,95±0,20	1,24±0,13
ProB	Bone marrow ^a	3,78±0,39	4,12±0,19
PreB	Bone marrow ^a	3,60±0,68	4,36±0,28
Immature	Bone marrow ^a	1,96±0,42	2,28±0,33
Recirculating	Bone marrow ^a	0,70±0,11	1,37±0,51
Mature	Spleen ^b	3,48±1,38	3,72±1,39
Follicular	Spleen ^b	1,91±0,70	2,14±0,82
Transitional	Spleen ^b	0,23±0,06	0,27±0,10
Marginal zone	Spleen ^b	0,45±0,21	0,45±0,16

a: cell number $\times 10^5 \pm \text{SEM}$

b: cell number $\times 10^7 \pm \text{SEM}$

Supplementary Table 1. Absolute cell number of bone marrow and splenic B cell subsets in CTCF^{fl/+} and CTCF^{fl/fl} mice.

SUPPLEMENTARY REFERENCE

1. Thomas-Claudepierre AS, Schiavo E, Heyer V, Fournier M, Page A, Robert I, *et al.* The cohesin complex regulates immunoglobulin class switch recombination. *The Journal of experimental medicine* 2013, **210**(12): 2495-2502.