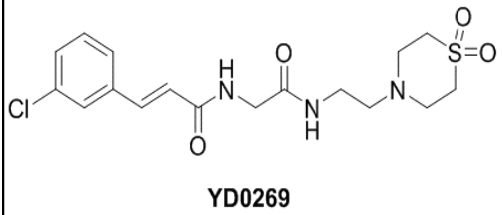
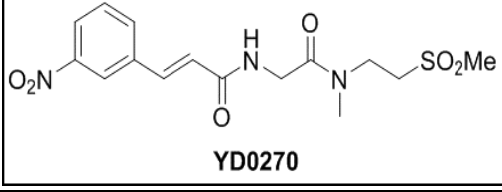
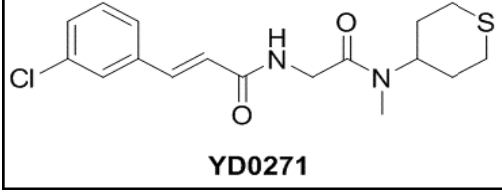
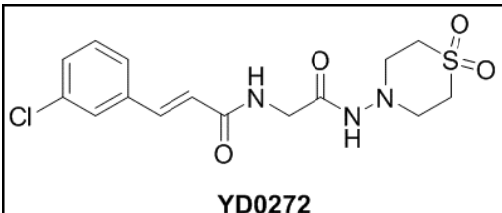
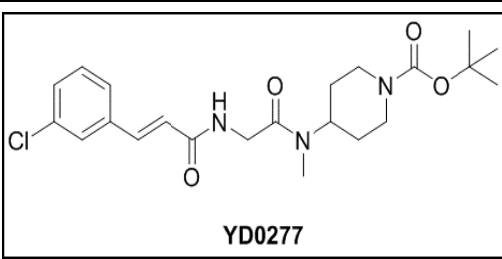
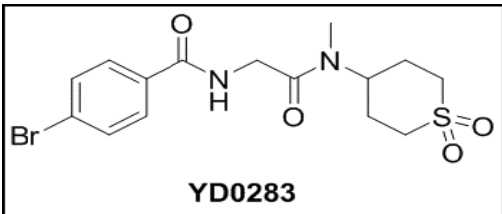
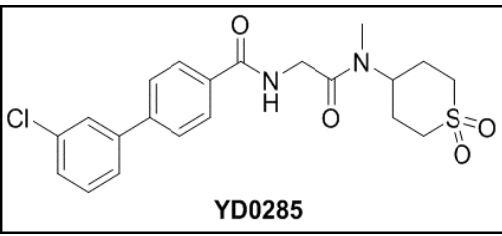
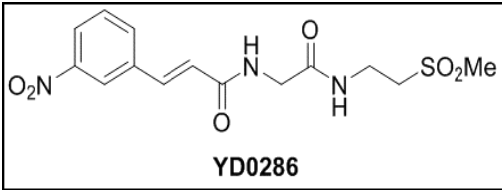


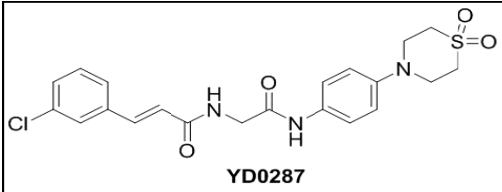
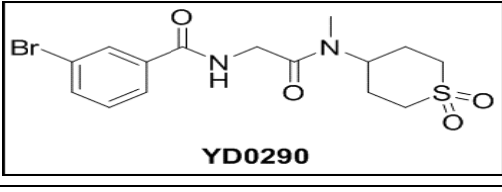
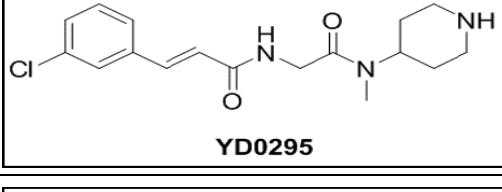
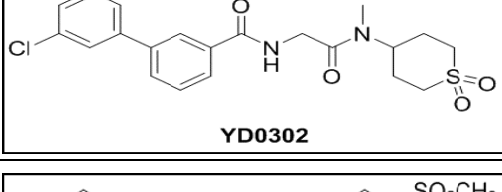
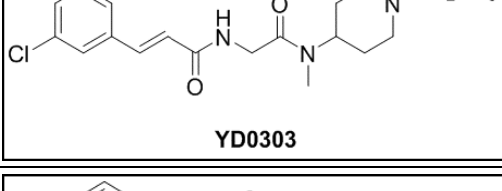
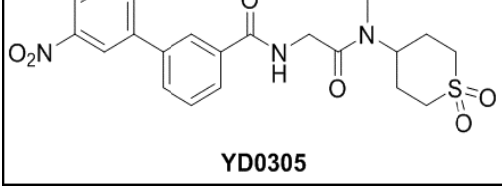
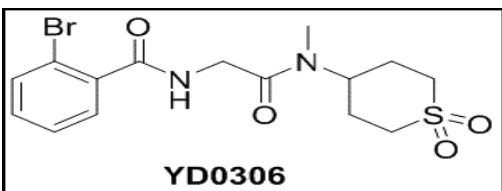
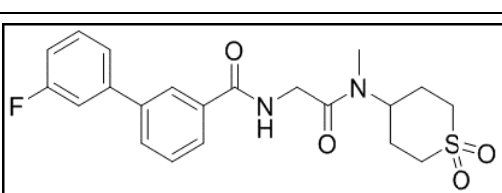
Supplementary Tables

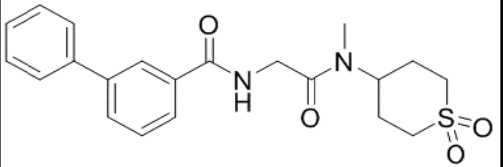
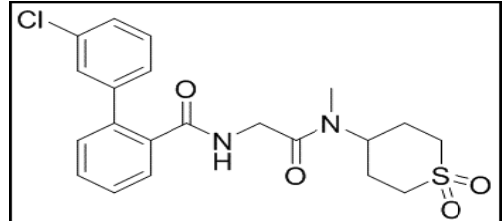
Supplementary Table 1

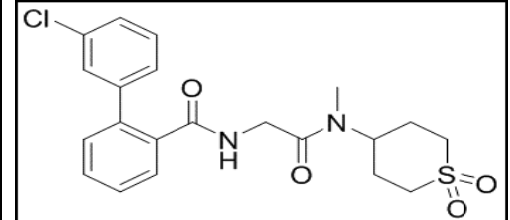
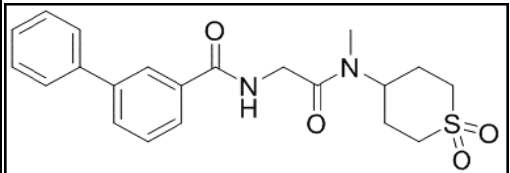
The structures of 25 compounds

No.	Compound ID	Molecular weight	Structures
1	YD-2-16	344.8138	 <chem>CC(=O)NCCNC(=O)/C=C/c1ccc(Cl)cc1</chem> YD0216
2	YD-2-40	384.8776	 <chem>CC(=O)NCCNC(=O)N1CCCN(C1)S(=O)(=O)C</chem> YD0240
3	YD-2-45	350.4326	 <chem>CC(=O)NCCNC(=O)N1CCCN(C1)S(=O)(=O)C</chem> YD0245
4	YD-2-54	384.8776	 <chem>CC(=O)NCCNC(=O)N1CCCN(C1)S(=O)(=O)C</chem> YD0254 (ML264)
5	YD-2-60	310.3687	 <chem>CC(=O)NCCNC(=O)/C=C/c1ccccc1</chem> YD0260
6	YD-2-62	358.8404	 <chem>CC(=O)NCCNC(=O)/C=C/c1ccc(Cl)cc1</chem> YD0262
7	YD-2-67	400.877	 <chem>CC(=O)NCCNC(=O)N1CCCN(C1)S(=O)(=O)C</chem> YD0267

8	YD-2-69	399.8923	 <p style="text-align: center;">YD0269</p>
9	YD-2-70	369.3929	 <p style="text-align: center;">YD0270</p>
10	YD-2-71	352.8788	 <p style="text-align: center;">YD0271</p>
11	YD-2-72	371.8391	 <p style="text-align: center;">YD0272</p>
12	YD-2-77	435.9443	 <p style="text-align: center;">YD0277</p>
13	YD-2-83	403.2914	 <p style="text-align: center;">YD0283</p>
14	YD-2-85	434.9363	 <p style="text-align: center;">YD0285</p>
15	YD-2-86	355.3663	 <p style="text-align: center;">YD0286</p>

16	YD-2-87	447.9351	 <p style="text-align: center;">YD0287</p>
17	YD-2-90	403.2914	 <p style="text-align: center;">YD0290</p>
18	YD-2-95	335.8285	 <p style="text-align: center;">YD0295</p>
19	YD-3-2	434.9363	 <p style="text-align: center;">YD0302</p>
20	YD-3-3	413.9189	 <p style="text-align: center;">YD0303</p>
21	YD-3-5	445.4888	 <p style="text-align: center;">YD0305</p>
22	YD-3-6	403.2914	 <p style="text-align: center;">YD0306</p>
23	YD-3-7	418.4817	 <p style="text-align: center;">YD0307</p>

24	YD-3-8	400.4913	 <p>YD0308</p>
25	YD-3-9	434.9363	 <p>YD0309</p>



Supplementary Table 2

The antibodies used for Western Blot

Name	MW (KDa)	Source	Company	Dilution	Product Number
PARP	116/89	Rabbit	Cell signaling	1:1000	9532S
Caspase3	32/21/14	Rabbit	Cell signaling	1:1000	9662S
cleave-Caspase 3	19/17	Rabbit	Cell signaling	1:1000	9661S
Caspase8	57/43/18	Mouse	Cell signaling	1:1000	9746S
Caspase9	47/37/35/17	Rabbit	Cell signaling	1:1000	9502S
Caspase7	35/20	Rabbit	Cell Signaling	1:1000	9494S
CyclinD1	36	Rabbit	Cell signaling	1:1000	2978S
p21	21	Rabbit	Cell signaling	1:1000	2947S
p27	27	Mouse	BD Biosciences	1:1000	610241
Bcl2	26	Rabbit	Cell signaling	1:1000	2870S
Bclxl	30	Rabbit	Cell signaling	1:1000	2764S
DR4	55	Rabbit	Cell Signaling	1:1000	42533
DR5	55/40	Rabbit	Sigma	1:1000	D3938
BID	22/15	Rabbit	Cell signaling	1:1000	9942
Bak	25	Rabbit	Cell signaling	1:1000	9942
Bax	20	Rabbit	Cell Signaling	1:1000	9942
AKT	60	Rabbit	Cell signaling	1:1000	4685S
p-AKT(Ser473)	60	Rabbit	Cell Signaling	1:1000	9018P
GAPDH	37	Rabbit	Santa Cruz Biotechnology	1:1000	Sc-25778
Bip	78	Rabbit	Cell signaling	1:1000	3177S
IRE1 α	130	Rabbit	Cell signaling	1:1000	3294S
JNK	46,54	Rabbit	Cell signaling	1:1000	9252S
p-JNK	46,54	Rabbit	Cell signaling	1:1000	9251S
c-Jun	43,48	Rabbit	Cell signaling	1:1000	9165S
p-c-Jun	48	Rabbit	Cell signaling	1:1000	9164S
PREK	130	Rabbit	Cell signaling	1:1000	5683S
CHOP	19	Mouse	Cell signaling	1:1000	2895S
ATF6	75	Mouse	abcam	1:1000	ab37149
KLF5	55	Rabbit	JTD	1:1000	/
Anti-Rabbit IgG	/	Goat	eBio-science	1:5000	/
Anti-Mouse IgG	/	Goat	eBio-science	1:5000	/

Supplementary Table 3

The IC₅₀ values of ML264 in cell lines

	Cell lines	IC₅₀ (μM)
ER α + Breast Cancer	MCF 7	11
	T-47D	>12
TNBC	SUM 149PT	>12
	MDA-MB-231	12
	MDA-MB-453	12
	MDA-MB-468	>12
	Hs 578T	>12
	HCC 1806	>12
	HCC 1937	>12
Normal Cells	MCF 10A	7
	HC 11	NA
	NIH3T3	NA
Colorectal Cancer	HCT 116	7

Supplementary Materials and Methods

Plasmid transfection

One day before transfection, MDA-MB-231 cells were plated in growth medium without antibiotics in 30-70% confluence. Plasmid DNA were first diluted in the appropriate amount of Opti-MEM® I Medium without serum. Then Lipofectamine™ 2000 were diluted in the appropriate amount of Opti-MEM® I Medium without serum. Following that, DNA was mixed with Lipofectamine™ 2000 gently and incubated for 20 minutes at room temperature to allow complex formation. Finally, the DNA -Lipofectamine™ 2000 complexes were added to the cells and mixed gently by rocking the plate back and forth. The cells were incubated at 37°C in a CO₂ incubator for 48 hours.

Supplementary Figure Legends

Figure S1. YD277 induced cell death can be partially inhibited by classical caspase inhibitor Z-VAD-fmk

- A.** MDA-MB-231 cells were seeded in a 6-well plate. Eighteen hours later, the cells were pre-treated with Z-VAD-fmk (MCE, New Jersey, USA, Cat#HY-16658, 100 μ M) for 1 hour and treated with YD277 for 36 hours. Caspase-7 and PARP cleavage were examined by Western blotting.
- B.** MDA-MB-231 cells were seeded in 48 well plates. Eighteen hours later, the cells were pre-treated with Z-VAD-fmk for 1 hour and treated with YD277 for 36 hours. The SRB assay was used to measure cell viability. Data is shown as mean \pm S.D. (n=3). **p < 0.01.

Figure S2. YD277 does not change the expression levels of DR4/5, Bak, Bax, and Bid proteins

MDA-MB-231 and MDA-MB-468 cells were treated with YD277 (1, 3, 10 μ M), ML264 (10 μ M), or DMSO for 36 h, and the expression levels of apoptosis-related proteins were detected by WB. β -actin normalized quantitative data were shown below their panel.

Figure S3. Over-expression of Bcl2/Bclxl partially rescued YD277-induced cell death

- A.** The pBabe-Bcl2 (β isoform) plasmid was used to generate retroviruses and to obtain stable Bcl2 over-expression MDA-MB-231 cell populations. Following that, the cells were transfected with pBabe-Bclxl plasmid for 36 hours and treated with YD277 (3 μ M) for another 36 hours. DMSO used as the negative control. The protein expression levels of Bclxl, Bcl2, and PARP were examined by WB. β -actin normalized quantitative data were shown below their panel.

B. The SRB assay was used to detect the cell viability. Data was shown as mean \pm S.D. (n=3) . *p < 0.05, **p < 0.01.

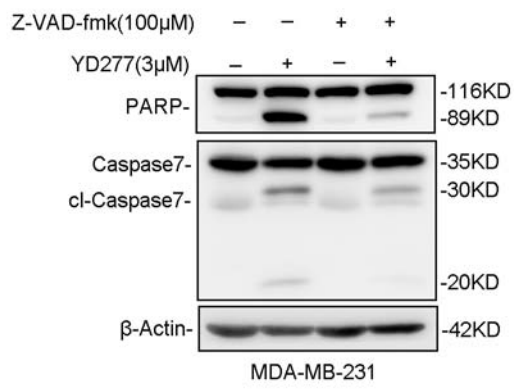
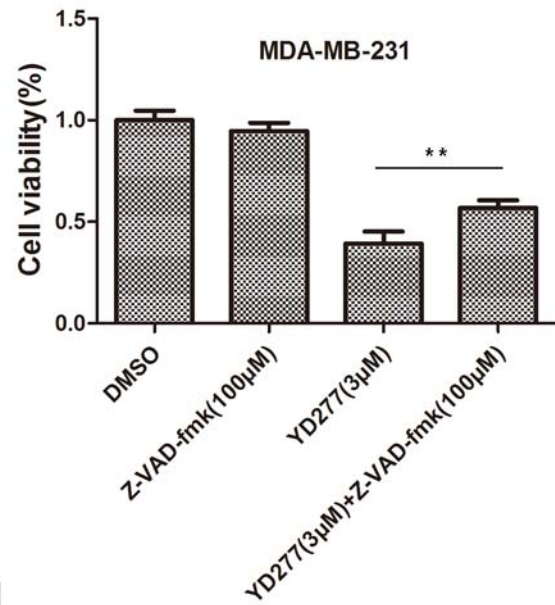
Figure S4. YD277 inhibits the activation of AKT

MDA-MB-231 cells were treated with YD277 (3 μ M) or DMSO for 36 h. MDA-MB-468 cells were treated with YD277 (3 μ M) for 48 hours. The expression levels of p-AKT and total AKT proteins were detected by WB. AKT normalized quantitative data for p-AKT was shown below the panel.

Figure S5. YD277 induces IRE1 α in MDA-MB-231 cells at the transcriptional level

- A.** YD277 increased *IRE1 α* mRNA levels in a dose dependent manner. SYBR Select Master Mix (Life technologies) was used for quantitative RT-PCR. MDA-MB-231 cells were treated with YD277 (1, 3, 10 μ M) for 24 hours. Total RNA was extracted for RT-qPCR. The expression levels of *IRE1 α* was normalized to *GAPDH*. Human *IRE1 α* forward primer sequence is 5'-TGTCAGAGATCTCCTCCGAG -3' and its reverse primer sequence is 5'-TCTCTCGTGGCTGCACAGCTCCA -3'. Human *GAPDH* forward primer sequence is 5'- GAAAGCCTGCCGGTGACTAA -3' and its reverse primer sequence is 5'- GCCCAATACGACCAAATCAGAGA -3'. *, p<0.05, student's t-test.
- B.** YD277 increased *IRE1 α* mRNA levels in a time dependent manner. YD277 (1 μ M) was used to treat MDA-MB-231 cells for 12, 24, 36, and 48 hours. Total RNA was extracted for RT-qPCR. The expression levels of *IRE1 α* was normalized to *GAPDH*. *P < 0.05, **P < 0.01, Student's t-test.
- C.** YD277 increased *IRE1 α* mRNA levels in an actinomycin D dependent manner. YD277(1 μ M) was used to treat MDA-MB-231 cells for 12 or 24 hours. Actinomycin D(1 μ g/ml)was added with YD277 at the same time as

indicated. Total RNA was extracted for RT-qPCR. The expression levels of *IRE1α* was normalized to *GAPDH*. **P < 0.01, Student's t-test.

A**B****Figure S1**

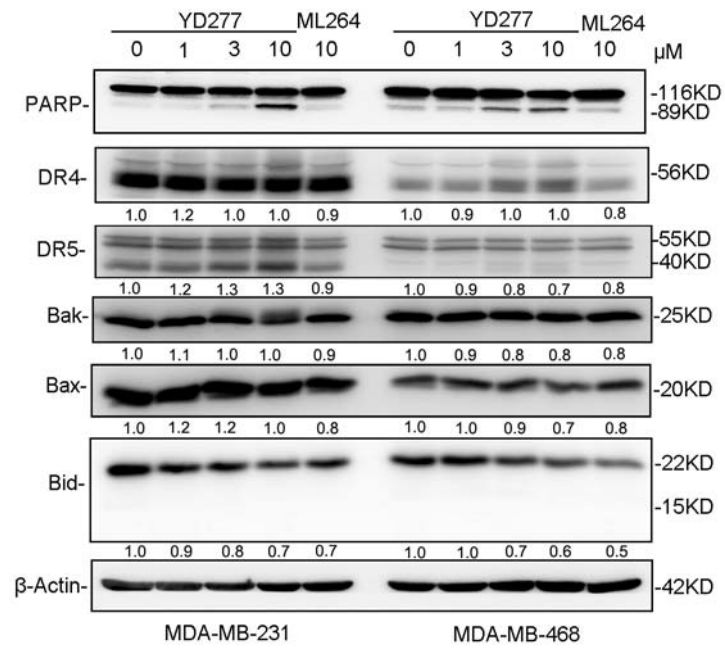


Figure S2

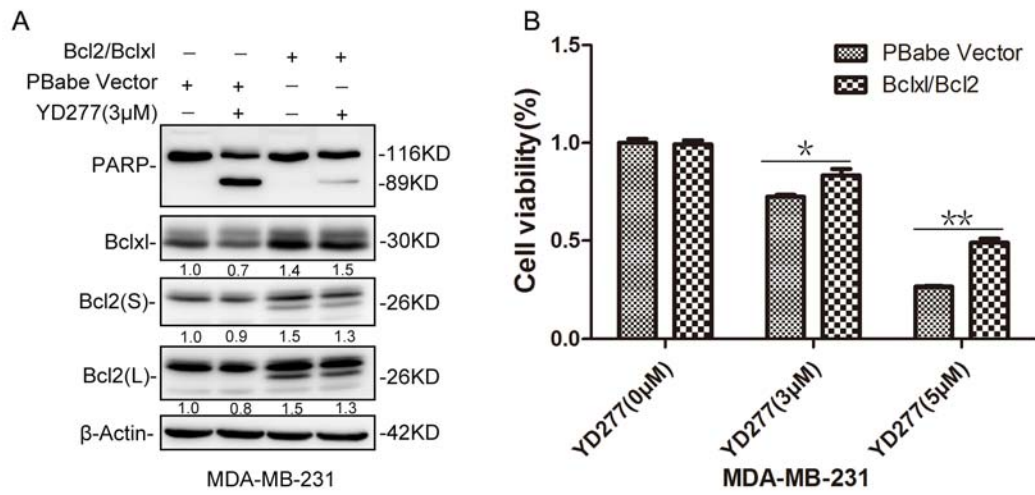


Figure S3

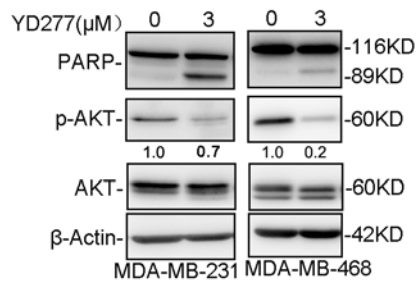


Figure S4

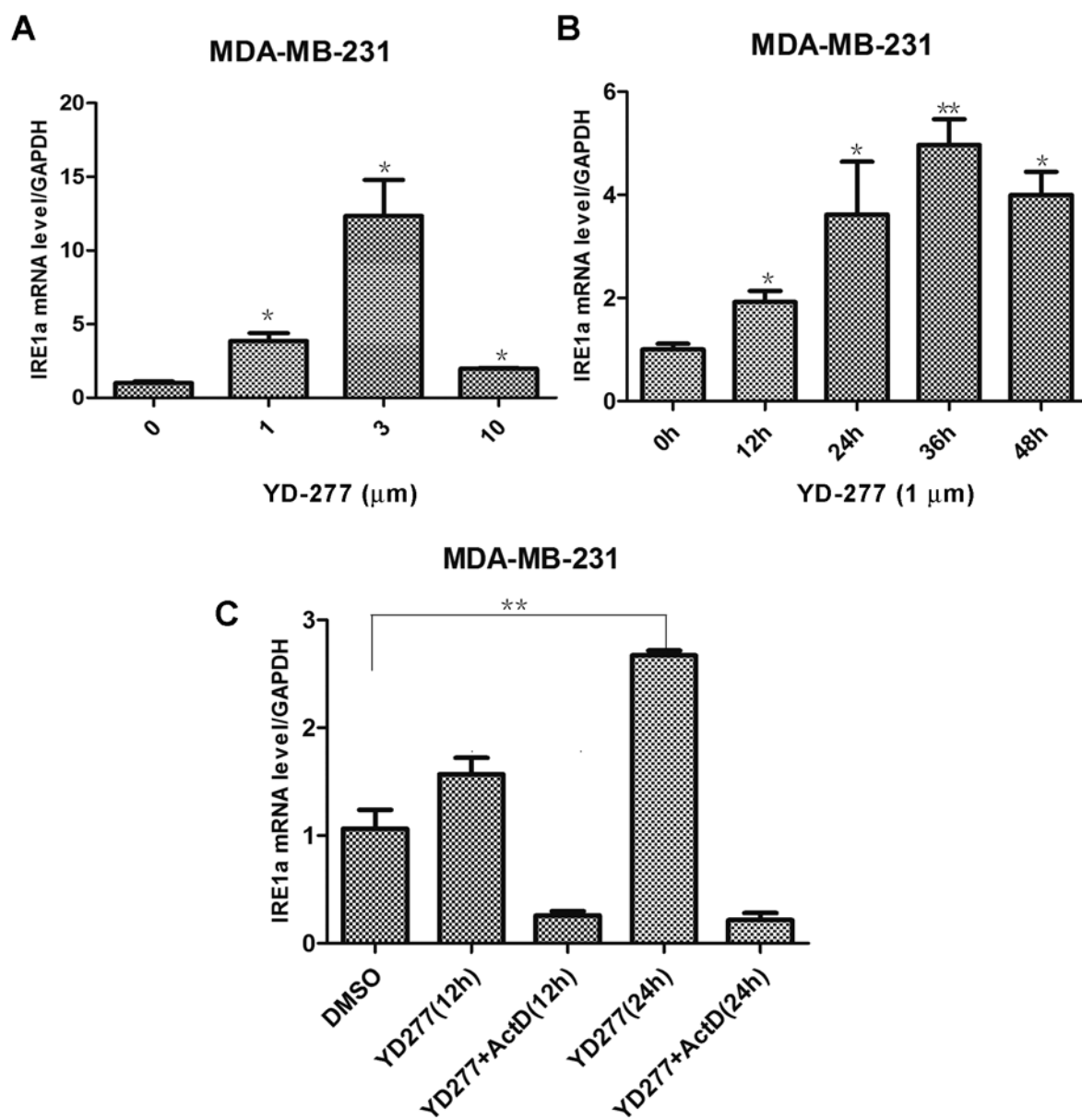


Figure S5