

## SUPPLEMENTAL FIGURES AND TABLES

### Biosynthesis and regulation of sulfomenaquinone, a metabolite associated with virulence in *Mycobacterium tuberculosis*\*

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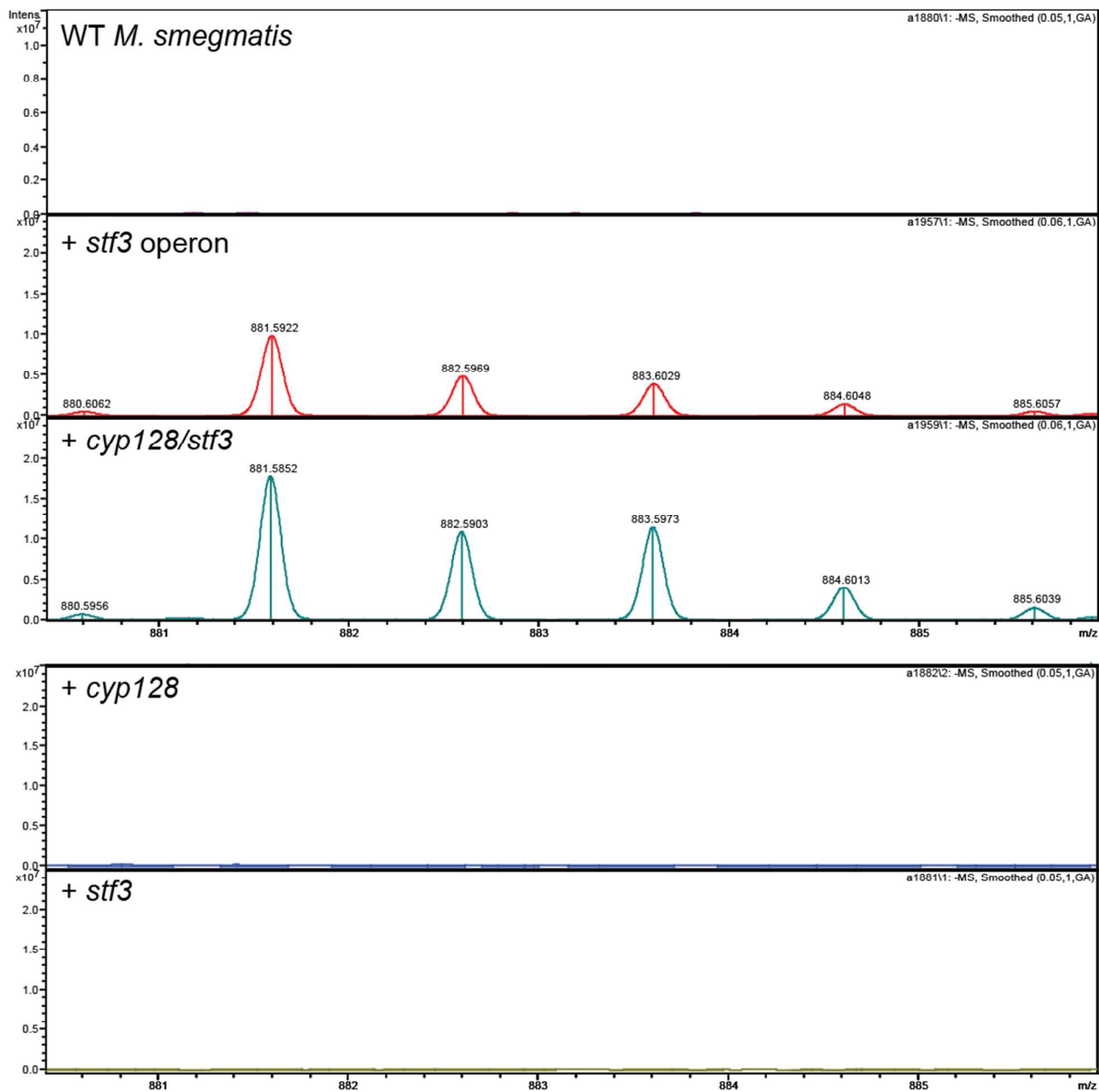
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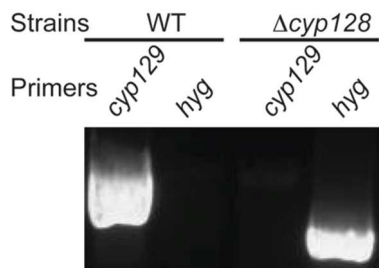
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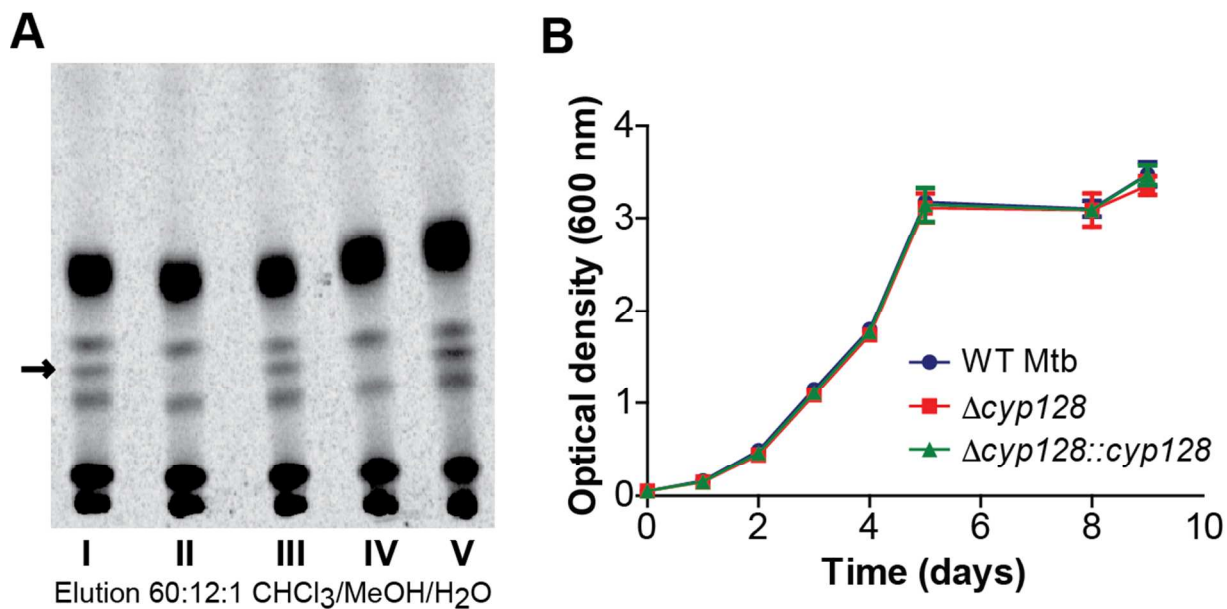
**FIGURE S1:** Mass spectra of TLE from *M. smegmatis* strains expressing SMK biosynthetic genes showing region m/z 880-886.



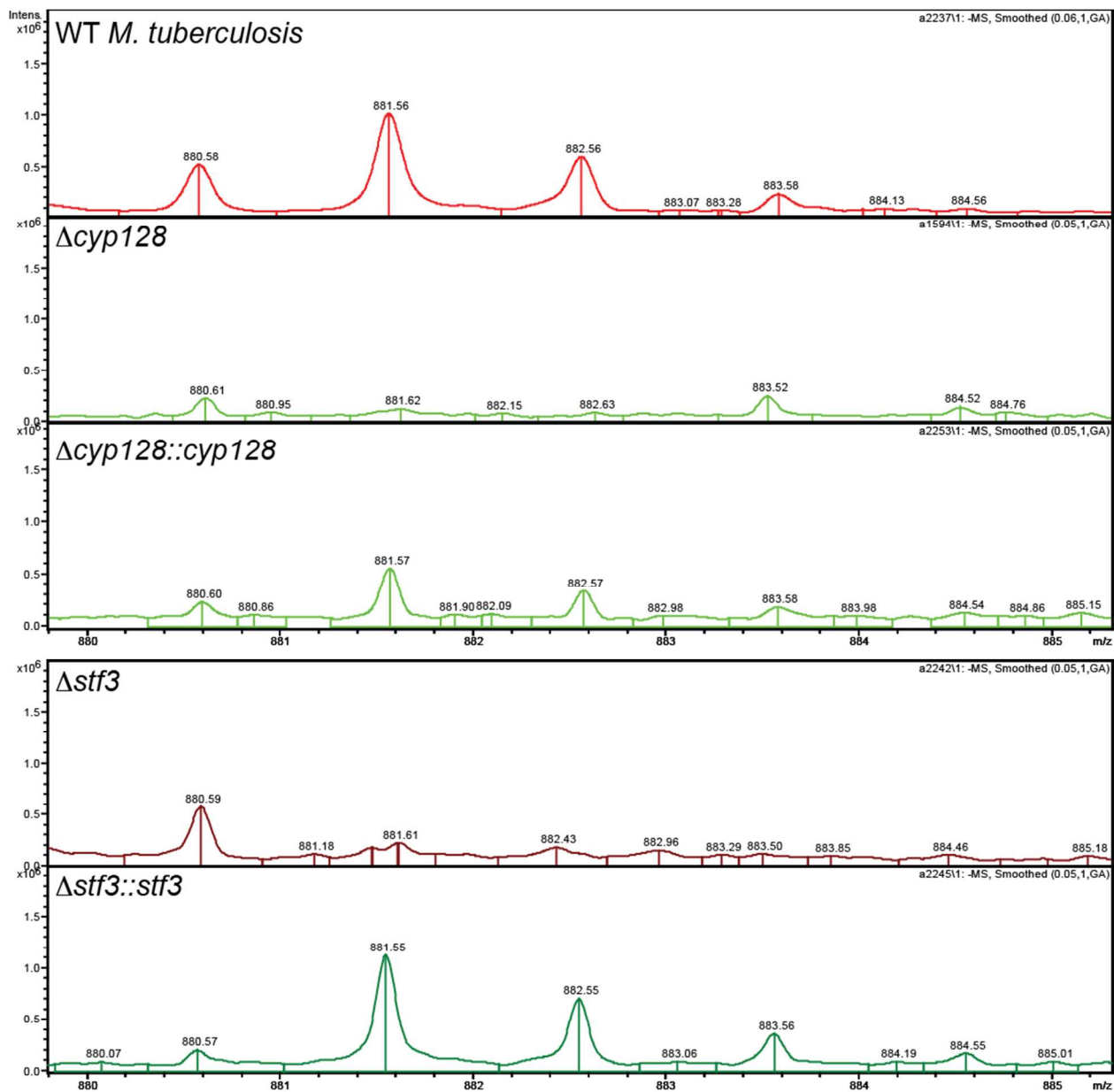
**FIGURE S2:** DNA gel from WT *M. tuberculosis* and  $\Delta cyp128$  using primers for either *cyp128* or hygromycin.



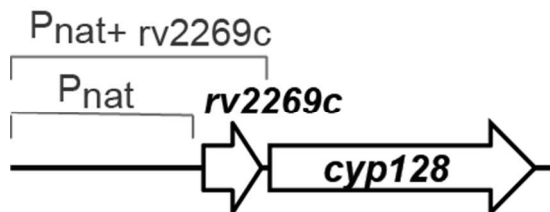
**FIGURE S3:** A) TLC analysis of TLE from *M. tuberculosis* strains grown on  $^{35}\text{S}$ -sulfate: (I) WT, (II)  $\Delta cyp128$ , (III)  $\Delta cyp128::cyp128$ , (IV)  $\Delta stf3$ , (V)  $\Delta stf3::stf3$ . Arrow indicates spot corresponding to SMK. B) Growth of  $\Delta cyp128$  and complement compared to WT in 7H9 liquid media.



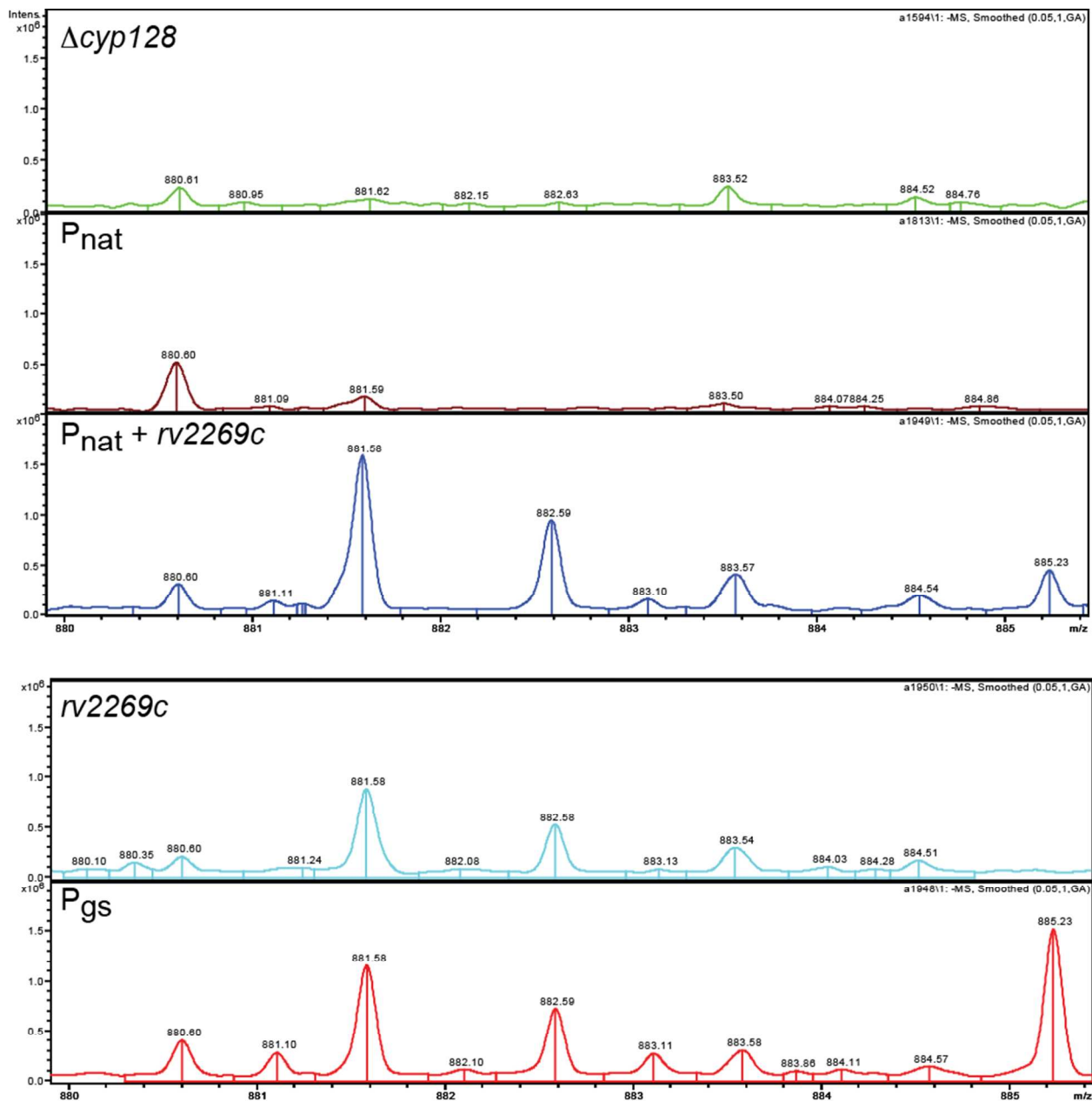
**FIGURE S4:** Mass spectra from TLE of *M. tuberculosis* WT and SMK deletion mutants with region m/z 880-886 shown.



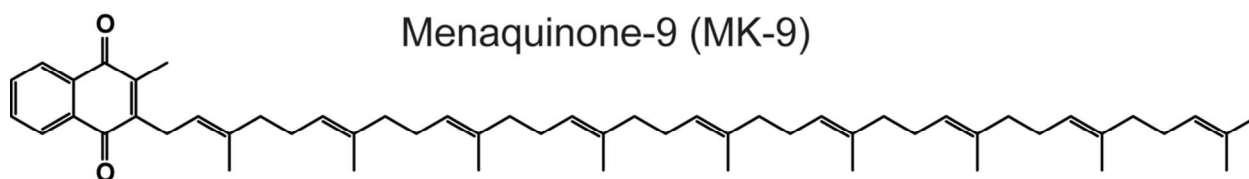
**FIGURE S5:** Scheme depicting promoters for *cyp128*.



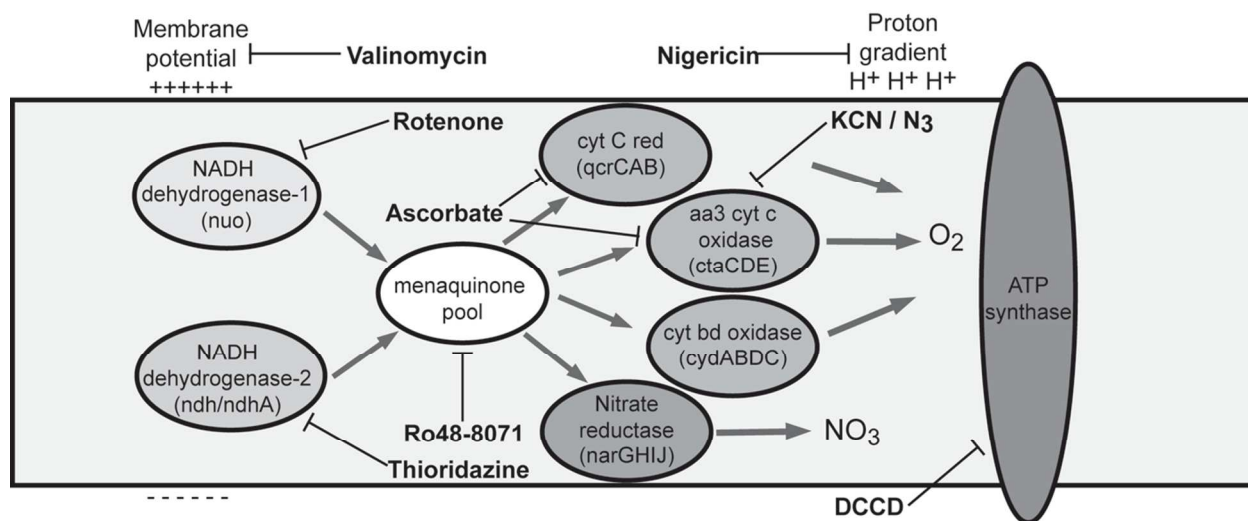
**FIGURE S6:** Mass spectra of TLE from  $\Delta cyp128$  complementation strains with *cyp128* under control of three different promoters with region *m/z* 880-886 shown.



**FIGURE S7:** Structure of menaquinone-9 (MK-9).



**FIGURE S8:** Schematic of *M. tuberculosis* electron transport chain and the inhibitors screened against WT *M. tuberculosis* and SMK mutants.



**TABLE S1:** Minimum inhibitory concentrations (MIC<sub>90</sub>) values for WT, SMK mutants and complements.

Chemical stress	MIC	Cell wall inhibitors	MIC
H <sub>2</sub> O <sub>2</sub>	110 mM	INH	0.06 µg/ml
NaNO <sub>3</sub> , pH 5.5	5 mM	ETA	5 µM
SDS	0.025%	ETH	6 µM

H<sub>2</sub>O<sub>2</sub> hydrogen peroxide; NaNO<sub>3</sub> sodium nitrate, SDS sodium dodecyl sulfate, INH isoniazid, ETA ethionamide, ETH ethambutol.

**TABLE S2:** Strains used in this study

Strains	Genotype	Source
<b><i>M. smegmatis</i></b>		
mc <sup>2</sup> 155	Wild type	
mc <sup>2</sup> 155 <i>rv2269c</i>	pKMS101; Kn <sup>r</sup> , contains <i>rv2269c</i>	This study
mc <sup>2</sup> 155 <i>cyp128</i>	pKMS102; Kn <sup>r</sup> , contains <i>cyp128</i>	This study
mc <sup>2</sup> 155 <i>stf3</i>	pKMS103; Kn <sup>r</sup> , contains <i>stf3</i>	This study
mc <sup>2</sup> 155 <i>stf3</i> operon	pKMS104; Kn <sup>r</sup> , contains <i>rv2269c</i> , <i>cyp128</i> , <i>stf3</i>	This study
mc <sup>2</sup> 155 <i>cyp128</i> , <i>stf3</i>	pKMS105; Kn <sup>r</sup> , contains <i>cyp128</i> and <i>stf3</i>	This study
<b><i>M. tuberculosis</i></b>		
H37Rv	Wild type	
H37Rv $\Delta$ <i>cyp128</i>	Hyg <sup>r</sup> , hyg cassette disrupting <i>cyp128</i>	This study
H37Rv $\Delta$ <i>cyp128::cyp128</i>	Hyg <sup>r</sup> , Kan <sup>r</sup> , complemented strain of $\Delta$ <i>cyp128</i>	This study
H37Rv $\Delta$ <i>stf3</i>	Hyg <sup>r</sup> , <i>stf3</i> interrupted by hyg resistance cassette	Ref <sup>1</sup>
H37Rv $\Delta$ <i>stf3::stf3</i>	Hyg <sup>r</sup> , Kan <sup>r</sup> , complement with <i>stf3</i> under the glutamine synthase promoter, modified pMV306 <sup>2</sup>	Ref <sup>1</sup>
H37Rv $\Delta$ <i>rv2269c</i>	Hyg <sup>r</sup> , <i>hsp60</i> promoter disrupting <i>rv2269c</i>	This study

**TABLE S3:** Plasmids used in this study

<b>Reference name</b>	<b>Description</b>	<b>Source</b>
pMV261	Kn <sup>r</sup> , pAL5000 origin, ColE1 origin, multiple cloning site, Phsp60 promoter	Ref <sup>2</sup>
pMV306	Kn <sup>r</sup> , A derivative of pMV261 lacking the Phsp60 promoter	Ref <sup>2</sup>
pKMS101	pMV261 derivative; contains <i>rv2269c</i>	This study
pKMS102	pMV261 derivative; contains <i>cyp128</i>	This study
pKMS103	pMV261 derivative; contains <i>stf3</i>	This study
pKMS104	pMV261 derivative; contains <i>rv2269c</i> , <i>cyp128</i> , and <i>stf3</i>	This study
pKMS105	pMV261 derivative; contains <i>cyp128</i> and <i>stf3</i>	This study
pKMS110	Plasmid used for <i>cyp128</i> disruption with hyg cassette	This study
pKMS109	Plasmid used for <i>rv2269c</i> disruption with hyg cassette	This study
pKMS133	Kn <sup>r</sup> , a derivative of pMV306 encoding <i>cyp128</i> with <i>rv2269c</i> promoter.	This study
pKMS130	Kn <sup>r</sup> , a derivative of pMV306 encoding <i>cyp128</i> with P <sub>nat</sub> (upstream 1 kb of the first gene in the putative operon) + <i>rv2269c</i> as the promoter.	This study
pKMS118	Kn <sup>r</sup> , a derivative of pMV306 encoding <i>cyp128</i> with P <sub>nat</sub> .	This study



**TABLE S4:** Primers used in this study. Restriction enzymes sequences are in bold and enzyme in parentheses.

<b>Primer name</b>	<b>Sequence</b>	<b>Description</b>
okms102	cacttcgcaat <b>ggcca</b> acgatgcgcgacccttagcg	5' pKMS101, pKMS104 (MscI )
okms109	actgttctacgcctctctga <b>atcgat</b> agggtcatga	3' pKMS101 (ClaI)
okms100	ccagcgtcagaaacaatgtg	5' pKMS102, pKMS105
okms101	cgtgacaacgggctgcttag	3' pKMS102
okms103	cacttcgcaat <b>ggcca</b> acgatgcgcgacccttagcg	5' pKMS103 (MscI)
okms112	actgttctacgcctctctga <b>atcgat</b> gtcg	3' pKMS103, pKMS104, pKMS105 (ClaI)
okms126	ccgtacgt <b>ctcgagg</b> tgagcaactgaccg	pKMS110 KO 5' cyp128 (XhoI)
okms127	caccatga <b>agctt</b> ggctcagaccaacgctcgggc	pKMS110 KO 5' cyp128 (HindIII)
okms128	ccg <b>ggta</b> ccgaatagagggtggtcgagc	pKMS110 KO 3' cyp128 (KpnI)
okms129	cggtact <b>taag</b> cgaacgctcgggtgtg	pKMS110 KO 3' cyp128 (AflII)
okms213	cg <b>ggta</b> ccgtggccaacgatgcgcg	5' pKMS133 (KpnI)
okms196	gtcgacat <b>cgat</b> gcacggcgaagcggttac	3' pKMS133 (ClaI)
okms179	<b>ttc</b> gaaatgaccgcgacacagtccc	5' pKMS118 (BstBI)
okms180	gacat <b>cgatt</b> gcgcggtcagaccaac	3' pKMS118 cyp128 (ClaI)
okms181	gc <b>ggta</b> ccgtggcttgccatgtcgttatgag	5' pKMS130 (KpnI)
okms196	gtcgacat <b>cgat</b> gcacggcgaagcggttac	3' pKMS130 (ClaI)
okms122	gtacgt <b>ctcgag</b> tgttaggccctcggccagcg	pKMS109 KO 5' rv2269 (XhoI)
okms123	gatccagat <b>at</b> caactgggcccgactgtgtagg	pKMS109 KO 5' rv2269 (EcoRV)
okms124	gacaggact <b>ctaga</b> cgcgaattattgcatgcccg	pKMS109 KO 3' rv2269 (XbaI)
okms125	gactagag <b>ggta</b> ccagcagtgctctcatag	pKMS109 KO 3' rv2269 (KpnI)

**References:**

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- (2) Stover, C. K.; la Cruz, de, V. F.; Fuerst, T. R.; Burlein, J. E.; Benson, L. A.; Bennett, L. T.; Bansal, G. P.; Young, J. F.; Lee, M. H.; Hatfull, G. F. New use of BCG for recombinant vaccines. *Nature* **1991**, *351* (6326), 456–460 DOI: 10.1038/351456a0.