# Imaging Genetics and Genomics in Psychiatry: A Critical Review of Progress and Potential

## Supplemental Information

## **The Origins of Imaging Genetics**

Guided by convergent evidence from *in vitro*, psychiatric, and behavioral candidate gene studies (e.g., Table 1), imaging genetics began in 1998 before the draft of the human genome was complete. Characterizing the replication challenge that is inherent to the field, the first two imaging genetics studies (1, 2) reached opposing conclusions on whether the missense ankyrin repeat and kinase domain containing 1 (ANKK1) C/T single nucleotide polymorphism (SNP), rs1800497 (also known as Taq1A, previously assigned to DRD2<sup>1</sup>), is associated with in vivo dopamine receptor type 2 (D2R) availability and density. Pohjalainen and colleagues (1) found that the T allele of rs1800497 is associated with reduced dopamine type 2/3 receptor availability in the striatum among 54 healthy volunteers. Contrastingly, Laruelle and colleagues (2) found no difference in binding according to rs1800497 genotype in a sample of healthy controls (n=47) and patients with schizophrenia (n=23); however, a consistent unreported trending association is observed in controls. A meta-analysis of *in vivo* and postmortem studies supports the association between the T allele and reduced D2R availability among healthy individuals (3). The mechanism underlying these functional associations remains controversial; it is plausible that they may emerge as a result of interactions between ANKK1 and DRD2 or linkage disequilibrium patterns with nearby SNPs within DRD2, or otherwise unknown interactions. Nonetheless,

\_

<sup>&</sup>lt;sup>1</sup> Notably, this SNP was initially mistakenly believed to be within the dopamine receptor type gene (*DRD2*) but actually resides downstream of *DRD2* within *ANKK1*, which codes for a protein kinase involved in signal transduction.

evidence suggests that this SNP may be associated with psychiatric phenotypes (4, 5) potentially as a result of these functional associations; (but see also (6)) and lack of GWAS significance (7).

These initial imaging genetic findings were followed in 1998 and 2000 by ligand binding studies by Heinz and colleagues in humans and rhesus monkeys that associated SPECT imaging of [I-123]β-CIT binding to polymorphisms within the serotonin transporter (*SLC6A4*) and dopamine transporter (*SLC6A3*) genes (8-11). Alongside *in vitro* studies, these initial imaging genetics studies have been highly influential, inspiring a wealth of research examining associations between these genotypes and individual differences in structural and functional neural phenotypes as well as psychiatric disorders and variability in behavior (e.g., (12)). Thus, from its ligand-based beginnings, imaging genetics has produced findings that converge with data from multiple other modalities providing potential mechanistic pathways through which genetic variation in some of the most well-studied candidate loci may impact psychiatrically relevant behavior and risk. For further historical review please see (13).

Imaging genetics did not become widespread until it employed functional magnetic resonance imaging (fMRI) to examine associations between functional polymorphisms in the apolipoprotein E (APOE), catechol-O-methyltransferase (COMT) and serotonin transporter (SLC6A4) and neural activation during memory and emotion tasks (14, 15). These studies paved the way for the broader adoption of imaging genetics in the context of functional and structural MRI due to its lower cost, wide availability and lack of ionizing radiation exposure. Further, the larger sample sizes that can be obtained using MRI have led to the development of massive datasets through data sharing and large scale studies (16-21). In addition to encouraging new ways of characterizing brain function and structure such as examining interactions within and between large scale brain networks, these large datasets allow for the application of analytic

techniques such as GWAS (22), gene x gene interaction (23), gene x environment interaction (24), and pathway analysis (25) that may link both genes and behavioral phenotypes to brain function in new and interesting ways. This extension to MRI has enabled the rapid expansion of the field and helped popularize the intermediate phenotype approach in psychiatry by helping contextualize gene – behavior relationships through the mediating effects of brain (**Figure 1A**) (26), which has subsequently been refined in the form of the research domain criteria (RDoC) for psychiatric disease (27).

#### **Gene x Environment Interaction**

Given large effects of the environment, and in particular childhood maltreatment and poverty, on the expression of psychopathology, a complete etiologic understanding requires the incorporation of environmental factors (28). The interplay between genotype and environmental factors (including adversity and advantage) may occur due to selective environmental exposure due to genotype (i.e., gene-environment correlation) or due to their interaction (i.e., gene x environment interaction, GxE). Inspired by GxE observations in traditional psychiatric genetics that have been profoundly influential (29) but have also grown increasingly contentious (30), imaging genetics has begun to interrogate GxE using single variant and polygenic approaches. For example, studies have linked a functional variant in FKBP5 that has been associated with stress-related psychopathology and disease, to threat-related amygdala responsiveness in the context of prior childhood maltreatment (31, 32). That this association occurs in the context of adversity occurring early in life is consistent with observations in clinical and molecular epigenetic research (33). In another recent example, within 3 independent samples, polygenic risk for schizophrenia was negatively associated with cortical thickness only among male

participants who used cannabis (34). While these findings are intuitively appealing and provide ample mechanistic speculation for psychopathology risk, it is important to highlight that GxE research within an imaging genetics framework is confronted by a host of unique challenges including assessment of the environment in resource intensive studies, the need to appropriately model covariates, as well as power limitations introduced by interactive terms; a more complete discussion of these unique challenges is presented in (24).

### **Supplemental References**

- 1. Pohjalainen T, Rinne JO, Nagren K, Lehikoinen P, Anttila K, Syvalahti EK, et al. (1998): The A1 allele of the human D2 dopamine receptor gene predicts low D2 receptor availability in healthy volunteers. *Molecular psychiatry*. 3:256-260.
- 2. Laruelle M, Gelernter J, Innis RB (1998): D2 receptors binding potential is not affected by Taq1 polymorphism at the D2 receptor gene. *Molecular psychiatry*. 3:261-265.
- 3. Gluskin BS, Mickey BJ (2016): Genetic variation and dopamine D2 receptor availability: a systematic review and meta-analysis of human in vivo molecular imaging studies. *Translational psychiatry*. 6:e747.
- 4. Ma Y, Wang M, Yuan W, Su K, Li MD (2015): The significant association of Taq1A genotypes in DRD2/ANKK1 with smoking cessation in a large-scale meta-analysis of Caucasian populations. *Translational psychiatry*. 5:e686.
- 5. Wang F, Simen A, Arias A, Lu QW, Zhang H (2013): A large-scale meta-analysis of the association between the ANKK1/DRD2 Taq1A polymorphism and alcohol dependence. *Human genetics*. 132:347-358.
- 6. Goldman D, Urbanek M, Guenther D, Robin R, Long JC (1998): A functionally deficient DRD2 variant [Ser311Cys] is not linked to alcoholism and substance abuse. *Alcohol*. 16:47-52.
- 7. Schizophrenia Working Group of the Psychiatric Genomics C (2014): Biological insights from 108 schizophrenia-associated genetic loci. *Nature*. 511:421-427.
- 8. Heinz A, Goldman D (2000): Genotype effects on neurodegeneration and neuroadaptation in monoaminergic neurotransmitter systems. *Neurochemistry international*. 37:425-432.
- 9. Heinz A, Goldman D, Jones DW, Palmour R, Hommer D, Gorey JG, et al. (2000): Genotype influences in vivo dopamine transporter availability in human striatum. *Neuropsychopharmacology*. 22:133-139.

10. Heinz A, Jones DW, Mazzanti C, Goldman D, Ragan P, Hommer D, et al. (2000): A relationship between serotonin transporter genotype and in vivo protein expression and alcohol neurotoxicity. *Biological psychiatry*. 47:643-649.

- 11. Heinz A, Higley JD, Gorey JG, Saunders RC, Jones DW, Hommer D, et al. (1998): In vivo association between alcohol intoxication, aggression, and serotonin transporter availability in nonhuman primates. *The American journal of psychiatry*. 155:1023-1028.
- 12. Caspi A, Hariri AR, Holmes A, Uher R, Moffitt TE (2010): Genetic sensitivity to the environment: the case of the serotonin transporter gene and its implications for studying complex diseases and traits. *The American journal of psychiatry*. 167:509-527.
- 13. Bigos KL, Weinberger DR (2010): Imaging genetics--days of future past. *NeuroImage*. 53:804-809.
- 14. Egan MF, Goldberg TE, Kolachana BS, Callicott JH, Mazzanti CM, Straub RE, et al. (2001): Effect of COMT Val108/158 Met genotype on frontal lobe function and risk for schizophrenia. *Proceedings of the National Academy of Sciences of the United States of America*. 98:6917-6922.
- 15. Hariri AR, Mattay VS, Tessitore A, Kolachana B, Fera F, Goldman D, et al. (2002): Serotonin transporter genetic variation and the response of the human amygdala. *Science*. 297:400-403.
- 16. Thompson PM, Stein JL, Medland SE, Hibar DP, Vasquez AA, Renteria ME, et al. (2014): The ENIGMA Consortium: large-scale collaborative analyses of neuroimaging and genetic data. *Brain imaging and behavior*. 8:153-182.
- 17. Swartz JR, Waller R, Bogdan R, Knodt AR, Sabhlok A, Hyde LW, et al. (2015): A Common Polymorphism in a Williams Syndrome Gene Predicts Amygdala Reactivity and Extraversion in Healthy Adults. *Biological psychiatry*.
- 18. Holmes AJ, Hollinshead MO, O'Keefe TM, Petrov VI, Fariello GR, Wald LL, et al. (2015): Brain Genomics Superstruct Project initial data release with structural, functional, and behavioral measures. *Scientific data*. 2:150031.
- 19. Schumann G, Loth E, Banaschewski T, Barbot A, Barker G, Buchel C, et al. (2010): The IMAGEN study: reinforcement-related behaviour in normal brain function and psychopathology. *Molecular psychiatry*. 15:1128-1139.
- 20. Satterthwaite TD, Elliott MA, Ruparel K, Loughead J, Prabhakaran K, Calkins ME, et al. (2014): Neuroimaging of the Philadelphia neurodevelopmental cohort. *NeuroImage*. 86:544-553.
- 21. Barch DM, Burgess GC, Harms MP, Petersen SE, Schlaggar BL, Corbetta M, et al. (2013): Function in the human connectome: task-fMRI and individual differences in behavior. *NeuroImage*. 80:169-189.
- 22. Hibar DP, Stein JL, Renteria ME, Arias-Vasquez A, Desrivieres S, Jahanshad N, et al. (2015): Common genetic variants influence human subcortical brain structures. *Nature*. 520:224-229.
- 23. Demers CH, Drabant Conley E, Bogdan R, Hariri AR (2016): Interactions Between Anandamide and Corticotropin-Releasing Hormone Signaling Modulate Human Amygdala

- Function and Risk for Anxiety Disorders: An Imaging Genetics Strategy for Modeling Molecular Interactions. *Biological psychiatry*.
- 24. Bogdan R, Pagliaccio D, Baranger DA, Hariri AR (2016): Genetic Moderation of Stress Effects on Corticolimbic Circuitry. *Neuropsychopharmacology*. 41:275-296.
- 25. Inkster B, Nichols TE, Saemann PG, Auer DP, Holsboer F, Muglia P, et al. (2010): Pathway-based approaches to imaging genetics association studies: Wnt signaling, GSK3beta substrates and major depression. *NeuroImage*. 53:908-917.
- 26. Meyer-Lindenberg A (2010): Intermediate or brainless phenotypes for psychiatric research? *Psychological medicine*. 40:1057-1062.
- 27. Cuthbert BN (2014): The RDoC framework: facilitating transition from ICD/DSM to dimensional approaches that integrate neuroscience and psychopathology. *World psychiatry : official journal of the World Psychiatric Association*. 13:28-35.
- 28. Teicher MH, Samson JA (2013): Childhood maltreatment and psychopathology: A case for ecophenotypic variants as clinically and neurobiologically distinct subtypes. *The American journal of psychiatry*. 170:1114-1133.
- 29. Caspi A, Sugden K, Moffitt TE, Taylor A, Craig IW, Harrington H, et al. (2003): Influence of life stress on depression: moderation by a polymorphism in the 5-HTT gene. *Science*. 301:386-389.
- 30. Duncan LE, Keller MC (2011): A critical review of the first 10 years of candidate gene-by-environment interaction research in psychiatry. *The American journal of psychiatry*. 168:1041-1049.
- 31. White MG, Bogdan R, Fisher PM, Munoz KE, Williamson DE, Hariri AR (2012): FKBP5 and emotional neglect interact to predict individual differences in amygdala reactivity. *Genes, brain, and behavior.* 11:869-878.
- 32. Holz NE, Buchmann AF, Boecker R, Blomeyer D, Baumeister S, Wolf I, et al. (2015): Role of FKBP5 in emotion processing: results on amygdala activity, connectivity and volume. *Brain structure & function*. 220:1355-1368.
- 33. Klengel T, Mehta D, Anacker C, Rex-Haffner M, Pruessner JC, Pariante CM, et al. (2013): Allele-specific FKBP5 DNA demethylation mediates gene-childhood trauma interactions. *Nature neuroscience*. 16:33-41.
- 34. French L, Gray C, Leonard G, Perron M, Pike GB, Richer L, et al. (2015): Early Cannabis Use, Polygenic Risk Score for Schizophrenia and Brain Maturation in Adolescence. *JAMA psychiatry*. 72:1002-1011.