

SmMYB36, a Novel R2R3-MYB Transcription Factor, Enhances Tanshinone Accumulation and Decreases Phenolic Acid Content in *Salvia miltiorrhiza* Hairy Roots

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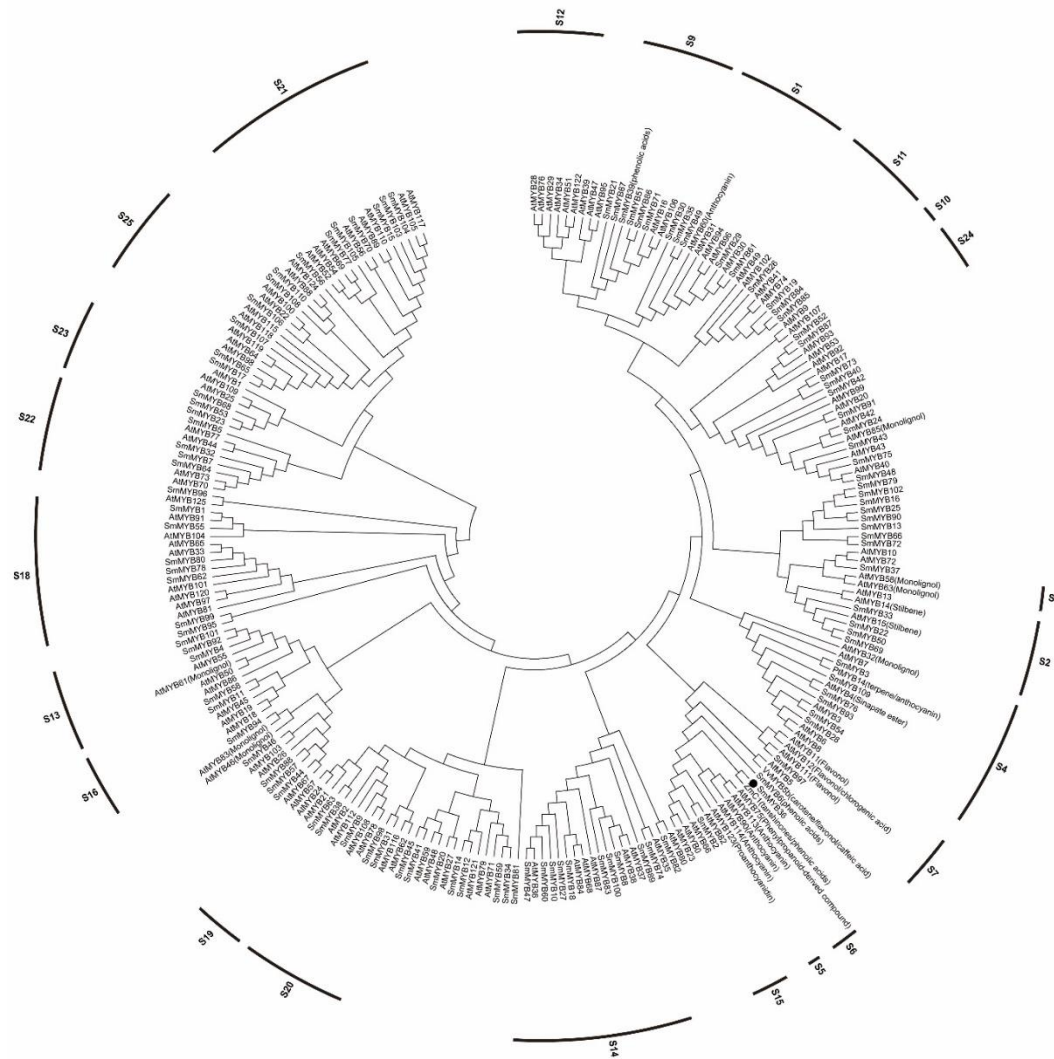
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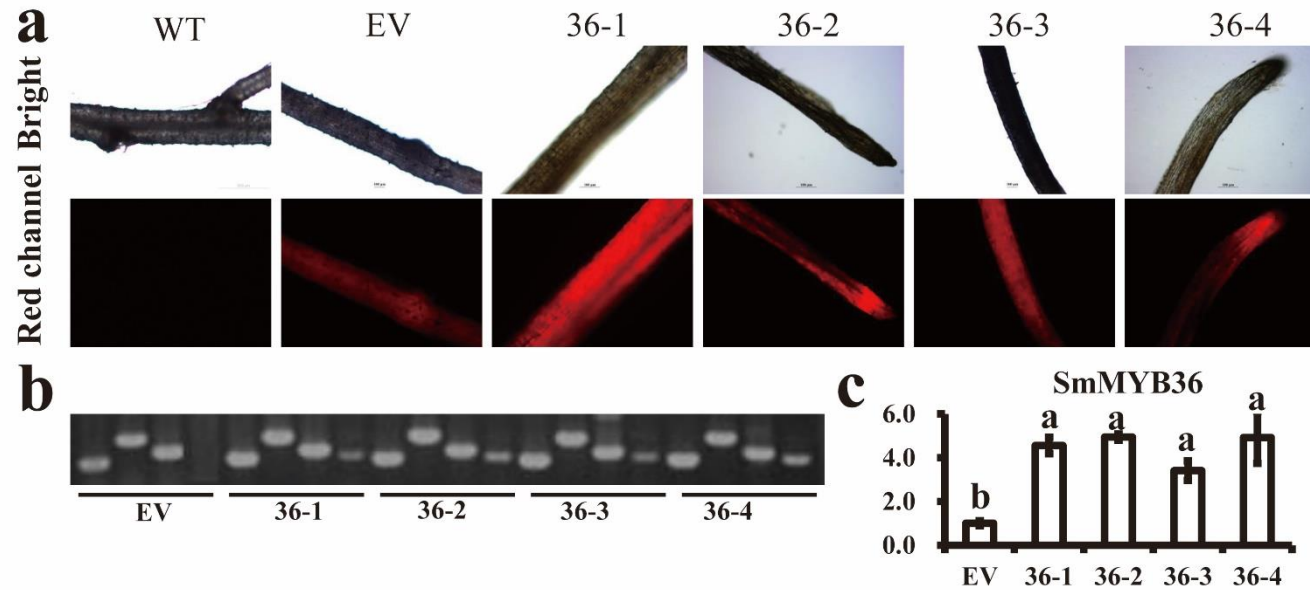
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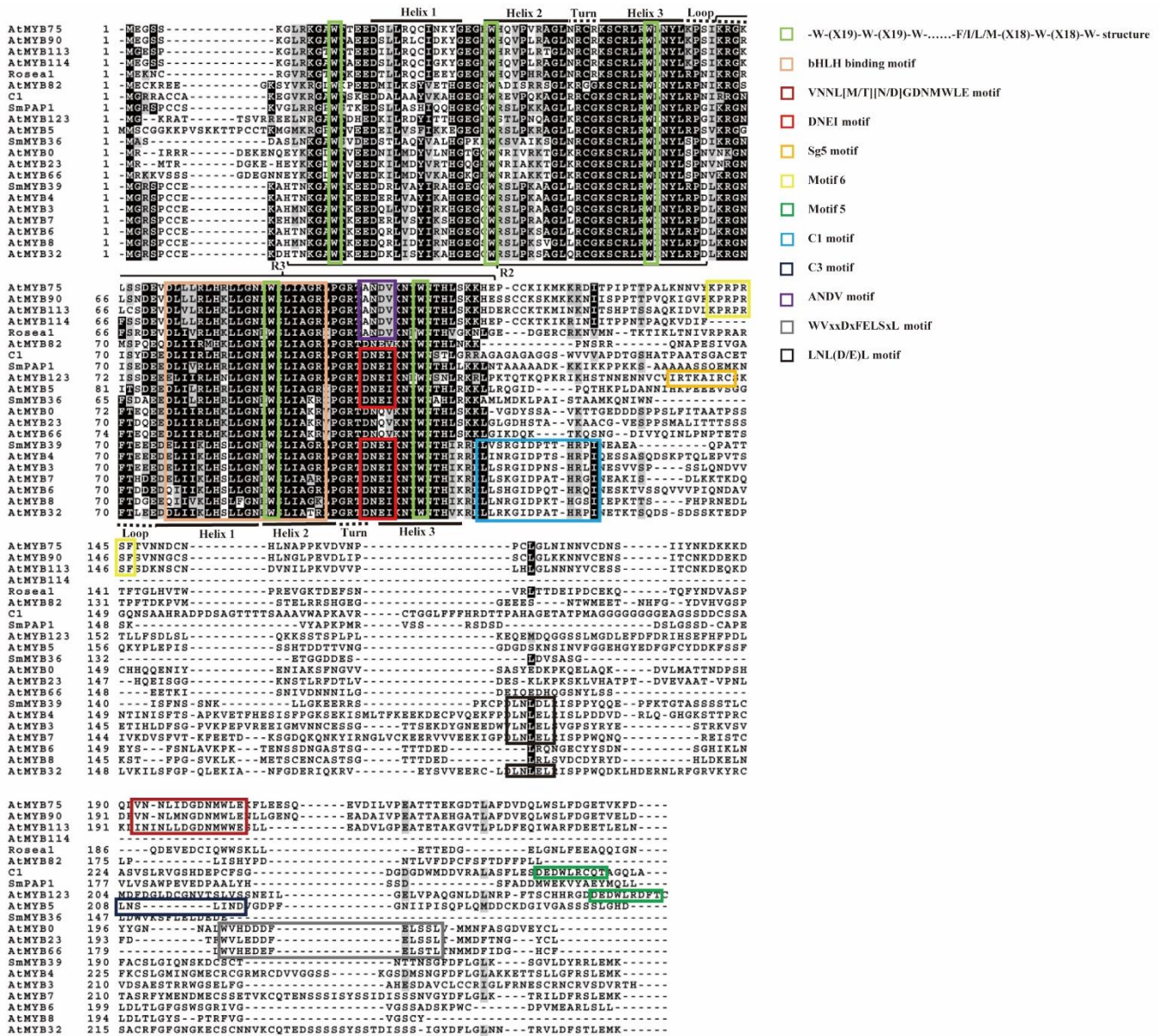
Zongsuo Liang, liangzs@ms.iswc.ac.cn



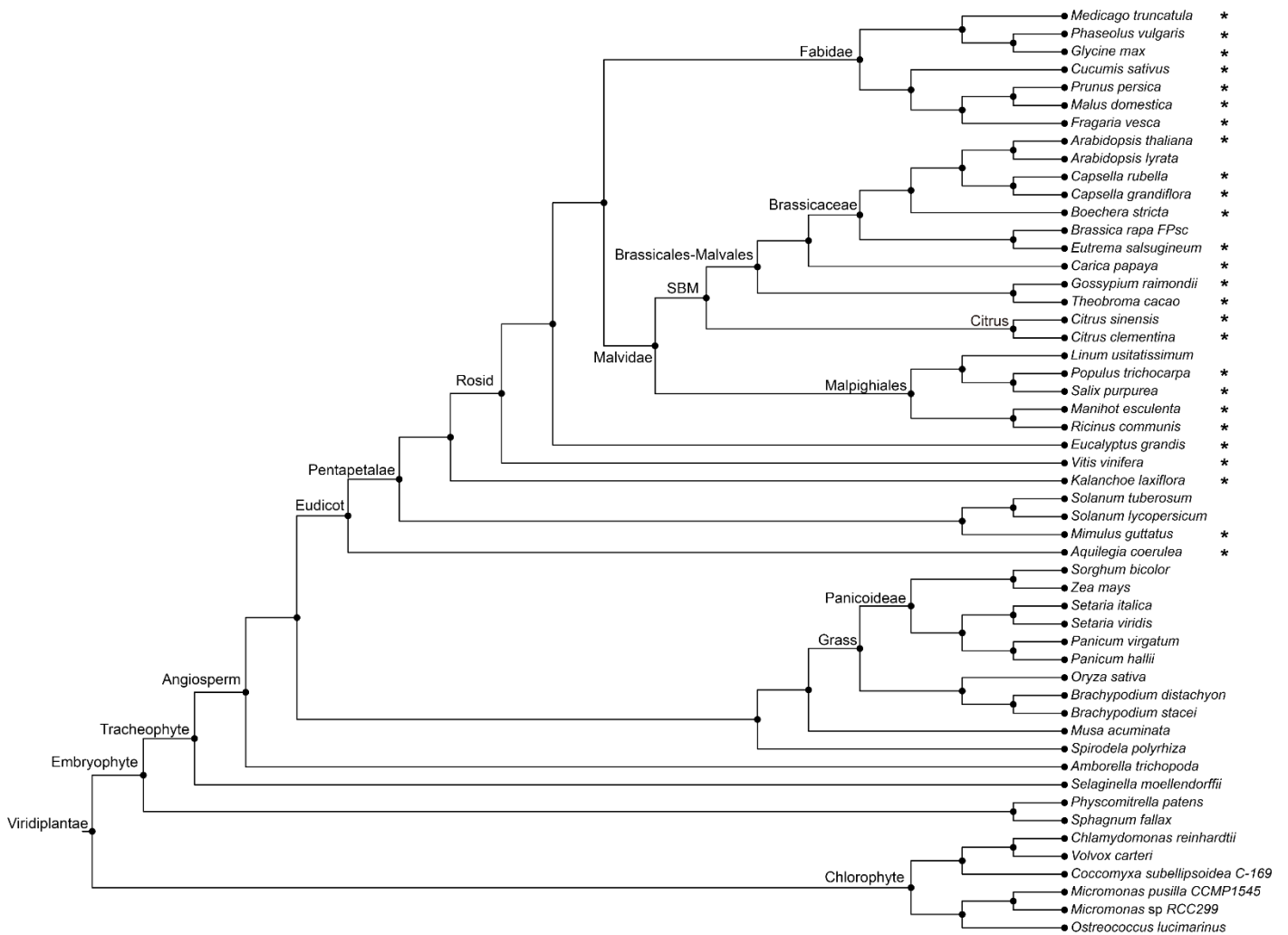
Supplementary Fig. S1. The phylogenetic tree for predicting the function of SmMYB36. The phylogenetic tree was constructed by maximum likelihood method of MEGA 6.06 based on the multiple sequence alignment using ClustalW method. The amino acid sequences of SmMYB36, ZmC1, VvMYB5b, PtMYB14 and all R2R3-MYBs of *Arabidopsis thaliana* and *Salvia miltiorrhiza* were used to construct the tree. SmMYB36 gathers together with these R2R3-MYBs whose function is regulating terpene or phenylpropanoid-derived compound biosynthesis.



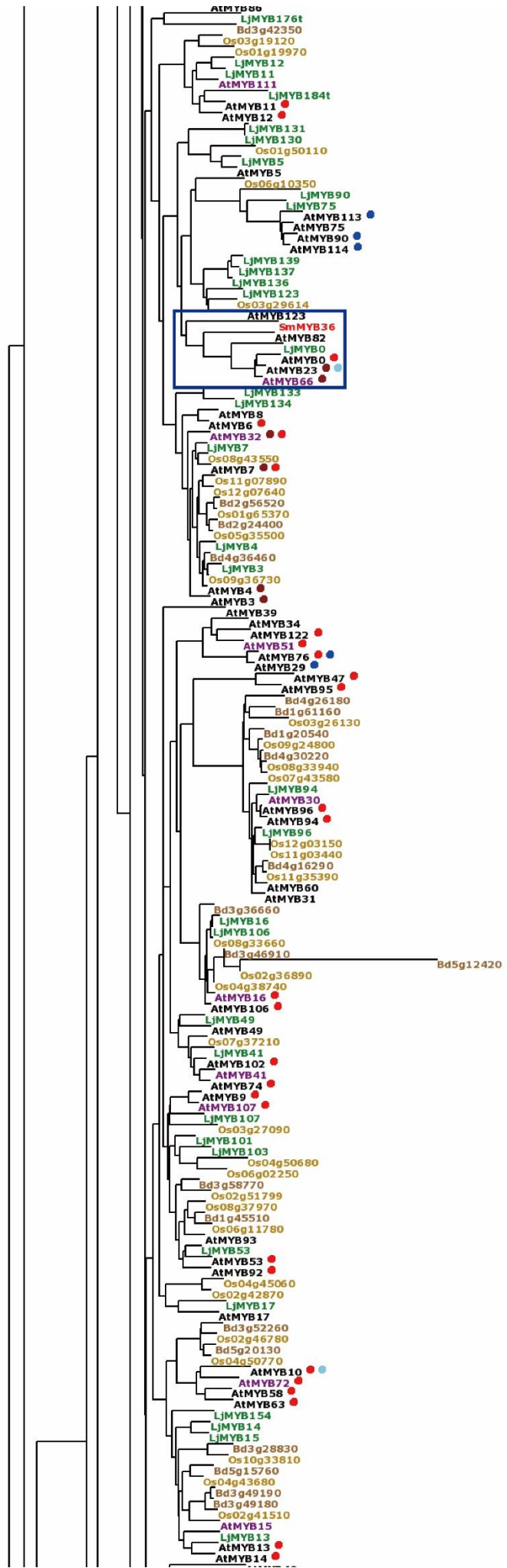
Supplementary Figure S2. (a) Bright field (upper lane) and red fluorescent field (bottom lane) of hairy roots of *Salvia miltiorrhiza*. Fluorescence was observed using a fluorescent microscope after harvest. (b) PCR identifications of hairy roots using *rolB*, *rolC*, *npt* and *SmMYB36* specific primers. The pictures show WT (infected by *Agrobacterium rhizogenes* strain ATCC15834), EV (infected by *Agrobacterium rhizogenes* strain ATCC15834 containing plasmid pK7FWG2R-EV), line 36-1, line 36-2, line 36-3, line 36-4 (infected by *Agrobacterium rhizogenes* strain ATCC15834 containing plasmid pK7FWG2R-SmMYB36) hairy roots from left to right. (c) Relative expression level of *SmMYB36* in transgenic hairy roots.

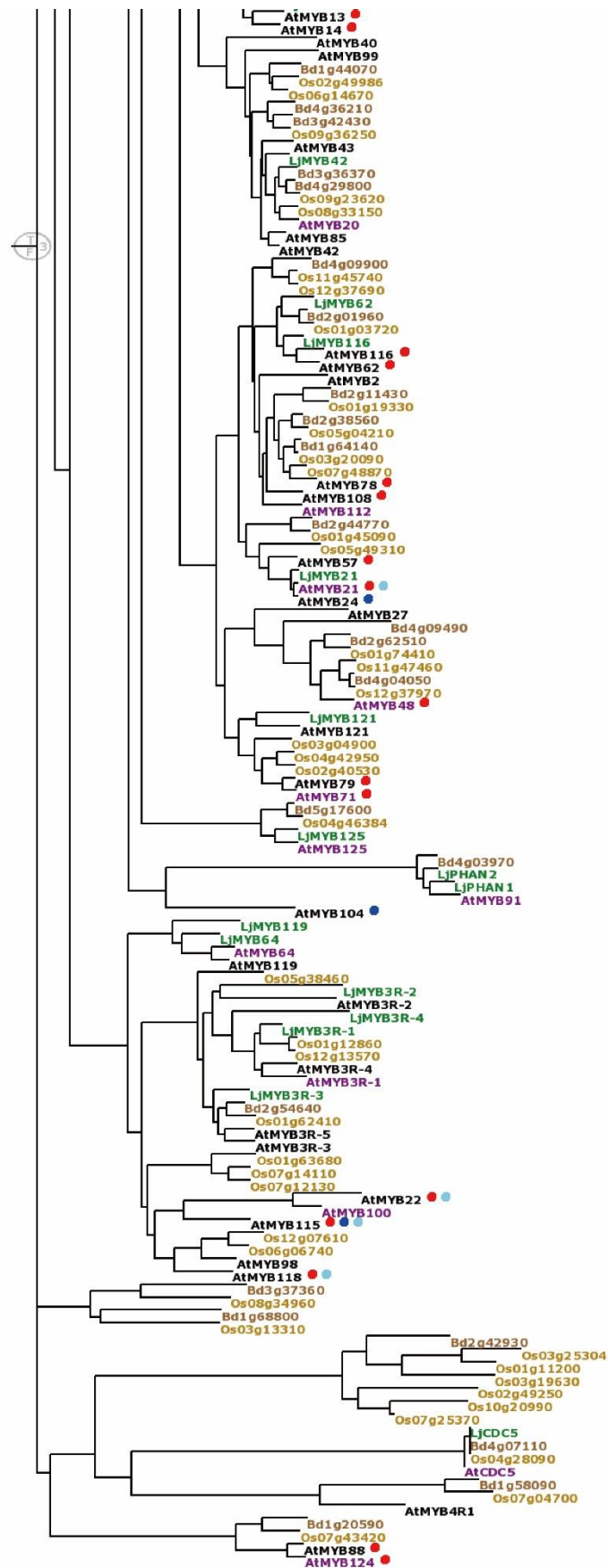


Supplementary Figure S3. The multiple sequence alignment and motif analysis of R2R3-MYB transcription factors.

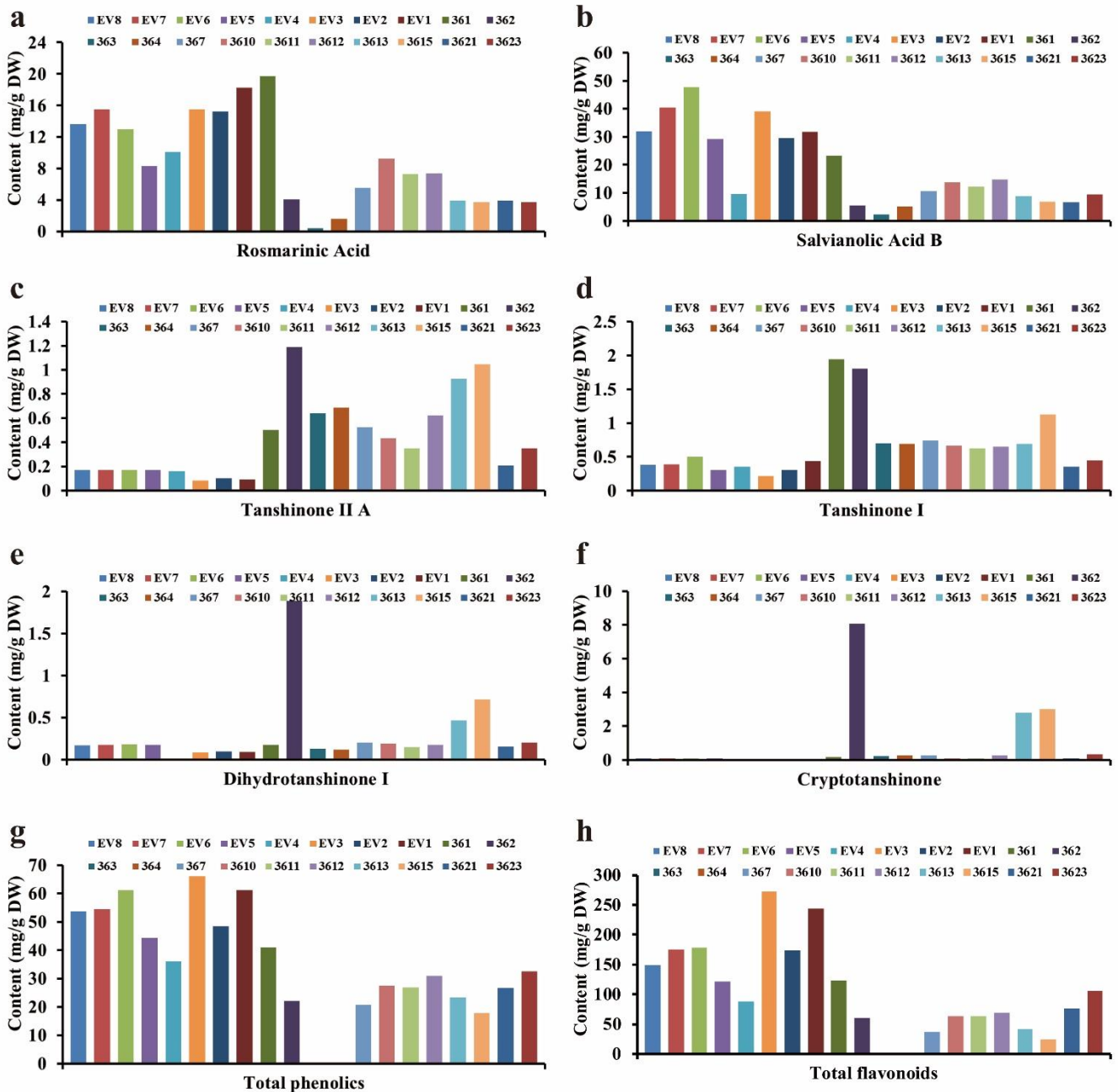


Supplementary Figure S4. The species phylogenetic tree which was slightly modified from the tree representation of the species in Phytozome v11.0. The species were emphasized (*) which contains the predicted orthologous genes of SmMYB36.

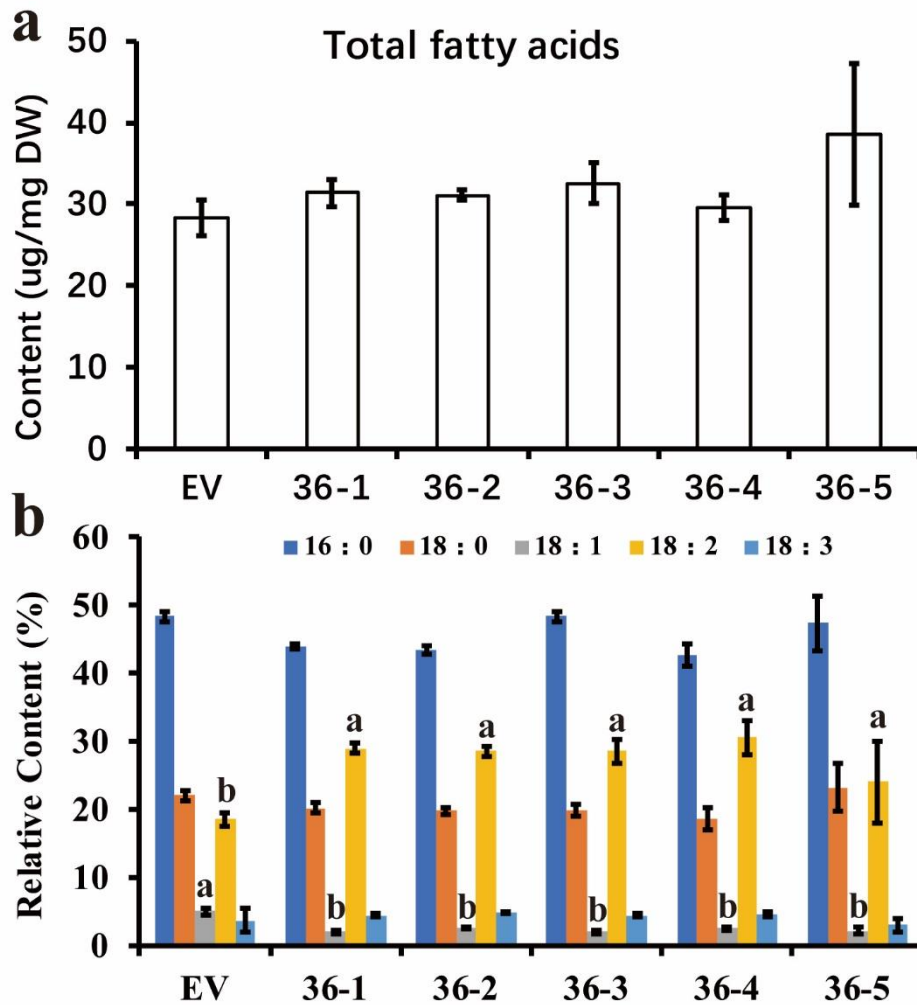




Supplementary Figure S5. The phylogenetic tree of R2R3-MYB transcription factors based on IT3F. The phylogenetic tree was constructed by submitting the amino acid sequence of SmMYB36 to the R2R3-MYB protein family of *Arabidopsis thaliana*, *Oryza sativa* L., *Brachypodium distachyon* and *Lotus japonicas* in IT3F website (<http://jicbio.nbi.ac.uk/IT3F/>). The red text represents the query sequence (SmMYB36) and the R2R3-MYBs of *A.thaliana* were outlined by blue box which have the closest relationships with SmMYB36.



Supplementary Fig. S6. The preliminary experiment results of twelve *SmMYB36*-overexpressing lines (without replicates) and eight empty-vector control lines (without replicates). (a) The content of rosmarinic acid in hairy roots. (b) The content of salvianolic acid B in hairy roots. (c) The content of tanshinone II A in hairy roots. (d) The content of tanshinone I in hairy roots. (e) The content of dihydrotanshinone I in hairy roots. (f) The content of cryptotanshinone in hairy roots. (g) The content of total phenolics in hairy roots. The content of line 363 and 364 was not detected. (h) The content of total flavonoids in hairy roots. The content of line 363 and 364 was not detected.



Supplementary Fig. S7. (a) The content of total fatty acids in hairy roots of *S. miltiorrhiza*. (b) The relative contents of palmitic acid (C16:0), stearic acid (18:0), oleic acid (C18:1), linoleic acid (C18:2), linolenic acid (C18:3) in hairy roots of *S. miltiorrhiza*. The GC analysis have three biological repeats of transgenic lines and each biological repeat has three technological repeats. The metabolite contents were shown by their means \pm SD.

Supplementary Table S1. Primers and probes used in the research

Primer Name	Primer Sequence (5' to 3')	Description
SmMYB36 F1	GGGGACAAGTTTGTACAAAAAAGCAGGCTTAATG GCGAGTGATGCATCTCT	The primers were used to generate the pDONR207-SmMYB36 entry vector.
SmMYB36 R1	GGGGACCACTTTGTACAAGAAAGCTGGGTATCATT CATCCTCGTCGAGTTC	
SmMYB36 F2	CCGCTCGAGATGGCGAGTGATGCATCT	The primers were used to generate the pA7-GFP-SmMYB36 vector.
SmMYB36 R2	GGACTAGTGATTCATCCTCGTCGAGTTCAAGAAA	
SmMYB36 F3	ATTTGGAGAGGACTCCGGTATT	The primers were used to identify positive hairy roots.
SmMYB36 R3	AATCCAACCCAGAGGCAGAG	
SmMYB36 F4	CCTCAACAGGTGCGGTAAGAGTT	The primers were used to quantitative analysis of SmMYB36.
SmMYB36 R4	TTCTTCCTCAAGTGAGCATTCCAGT	
ro1B F	GCTCTTGCAGTGCTAGATTT	The primers were used to confirm transgenic hairy roots.
ro1B R	GAAGGTGCAAGCTACCTCTC	
ro1C F	CTCCTGACATCAAACCTCGTC	The primers were used to confirm transgenic hairy roots.
ro1C R	TGCTTCGAGTTATGGGTACA	
NPT F	ACGTTGTCACTGAAGCGGGAAGG	The primers were used to confirm transgenic hairy roots.
NPT R	GGCGATACCGTAAAGCACGAGGAA	
SmMYB36 F5	GTTATTTCAATTGGAGAGGACTCCG	The primers were exogenous SmMYB36 specific primers and used to confirm transgenic hairy roots.
SmMYB36 R5	CCAGTAGTTCTTGATCTCGTTGTC	
SmMYB36 F6	CGCGGATCCATGGCGAGTGATGCATCT	The primers were used to generate the pET32a-SmMYB36 vector
SmMYB36 R6	CCGCTCGAGTTCATCCTCGTCGAGTTCAAG	
C4H1-MBS1-F	TCTTTAAACTCCA ACT GATATGATGTTTCG	The primers were used to generate the specific C4H1-MBS1 probe of <i>C4H1</i> promoter for EMSA.
C4H1-MBS1-R	CGAACATCATATC AGTT GGAGTTTTAAAGA	
C4H1-MBS3-F	GTTTAAAAGACATA ACT GAGTTATCGAACA	The primers were used to generate the specific C4H1-MBS3 probe of <i>C4H1</i> promoter for EMSA.
C4H1-MBS3-R	TGTTCGATA ACTCAGTT ATGTCTTTTAAAC	
4CL2-MRE-F	TTTGTAGAGTT GAACCTAA AGATGATGGGTG	The primers were used to generate the specific 4CL2-MRE probe of <i>4CL2</i> promoter for EMSA.
4CL2-MRE-R	CACCCATCATCT TTAGGTT CAACTCTACAAA	
HPPR-MRE-1-F	TTTTGTGGGACAA ACCTAAAA AAGAACTGAA	The primers were used to generate the specific HPPR-MRE probe of <i>HPPR</i> promoter for EMSA.
HPPR-MRE-1-R	TTCAGTTTCTTT TTAGGTT TGTCCACAAAA	
HPPR-MRE-2-F	AATGATTGATTT AACTAAT CATTGTGAAACC	The primers were used to generate the specific

HPPR-MRE-2-R	GGTTTCACAATGATTAGGTTAAATCAATCATT	HPPR-MRE probe of <i>HPPR</i> promoter for EMSA.
DFR-MBS3-F	AGCTAGCTAATTA ACT GAAAAACAATATT	The primers were used to generate the specific DFR-MBS3 probe of <i>DFR</i> promoter for EMSA.
DFR-MBS3-R	AATATTGTTTTT CAGTTA ATTAGCTAGCT	
DXR-MRE-F	CCCCTTTTACAAA ACCTA ATTTCCCCCAT	The primers were used to generate the specific DXR-MRE probe of <i>DXR</i> promoter for EMSA.
DXR-MRE-R	ATGGGGGGAAAT TAGGTTT TGAAAAGGGG	
MCT-MBS2-1-F	AGGAAACCCTC ACGGTC ACCTTAACATTCA	The primers were used to generate the specific MCT-MBS2 probe of <i>MCT</i> promoter for EMSA.
MCT-MBS2-1-R	TGAATGTTAAGGT GACCGT GAGGGTTTCCT	
MCT-MBS2-2-F	AAAACAGGCAT CGGTC AGTAAGTCCGCAT	The primers were used to generate the specific MCT-MBS2 probe of <i>MCT</i> promoter for EMSA.
MCT-MBS2-2-R	ATGCGGACTT ACTGACCG GATGCCTGTTTT	
MCT-MBS3-1-F	TATCATCTTCGGT AACTG ATACTCAAGGAT	The primers were used to generate the specific MCT-MBS3 probe of <i>MCT</i> promoter for EMSA.
MCT-MBS3-1-R	ATCCTTGAGTAT CAGTTAC CGAAGATGATA	
MCT-MBS3-2-F	AACTAGATGAAT AACTGG TAGCAATTACC	The primers were used to generate the specific MCT-MBS3 probe of <i>MCT</i> promoter for EMSA.
MCT-MBS3-2-R	GGTAATTGCTACC AGTTA ATTCATCTAGTT	
CMK-MBS1-F	TGAAGAATGGTG CAACTG CAATCAGTAAAA	The primers were used to generate the specific CMK-MBS1 probe of <i>CMK</i> promoter for EMSA.
CMK-MBS1-R	TTTTACTGATT GCAGTTG CACCATTCTTCA	
CMK-MBS3-F	CTCCTAATTATATA AACTGT CTCTGTATTAT	The primers were used to generate the specific CMK-MBS3 probe of <i>CMK</i> promoter for EMSA.
CMK-MBS3-R	ATAATACAGAGAC AGTTAT ATAATTAGGAG	
CMK-MRE-1-F	TCCACACTACAAA ACCTAAC CCTAACCGTA	The primers were used to generate the specific CMK-MRE probe of <i>CMK</i> promoter for EMSA.
CMK-MRE-1-R	TACGGTTAGGGT TAGGTTT TGTAGTGTGGA	
CMK-MRE-2-F	AGAATGGAATTT AACCTAA ATTAGTGTTC	The primers were used to generate the specific CMK-MRE probe of <i>CMK</i> promoter for EMSA.
CMK-MRE-2-R	TGAACACTAAT TAGGTTA AATTCCATTCT	
GGPPS1-MBS2-F	TTAAAATCGTT ACGGTC ATACCACGGAAAC	The primers were used to generate the specific GGPPS1-MBS2 probe of <i>GGPPS1</i> promoter for EMSA.
GGPPS1-MBS2-R	GTTCCGTGGTAT GACCGT AACGATTTAA	
GGPPS1-MRE-F	TTTTTCGATAGT AACCTA ATTTTCTGTAGTG	The primers were used to generate the specific GGPPS1-MRE probe of <i>GGPPS1</i> promoter for EMSA.
GGPPS1-MRE-R	CACTACAGAAA ATTAGGTT ACTATCGAAAA	

IPPI-MBS1-F	CCGCCGCCGCCACA ACT GTAATGGTGCTTT	The primers were used to generate the specific IPPI-MBS1 probe of <i>IPPI</i> promoter for EMSA.
IPPI-MBS1-R	AAAGCACCATTACAGTTGTGGCGGCGGCGG	
HMGS1-MBS3-F	GTGTGTGCTGTT TA ACTGATCCGCTATTTTC	The primers were used to generate the specific HMGS1-MBS3 probe of <i>HMGS1</i> promoter for EMSA.
HMGS1-MBS3-R	GAAATAGCGGATCAGTTAAACAGCACACAC	
MBS I -1-F	AAAAA ACCGTTA	The primers were used to generate the core MBS I probe for EMSA.
MBS I -1-R	TAACGGTTTTTTT	
MBS I -2-F	AAAAA ACCGTTA	The primers were used to generate the core MBS I probe for EMSA.
MBS I -2-R	TAACCGTTTTTTT	
MBS II -F	AAAAG TTAGTTA	The primers were used to generate the core MBS II probe for EMSA.
MBS II -R	TAACTAACTTTT	
MBSF1	CAACTG	The primers were used to generate the core MBS1 probe for EMSA.
MBSR1	CAGTTG	
MBSF2	CGGTCA	The primers were used to generate the core MBS2 probe for EMSA.
MBSR2	TGACCG	
MBSF3	TAACTG	The primers were used to generate the core MBS3 probe for EMSA.
MBSR3	CAGTTA	
MREF	AACCTAA	The primers were used to generate the core MRE probe for EMSA.
MRER	TTAGGTT	
6CKF	GATT CG	The primers were used to generate the control probe for the core probe (MBS1) for EMSA.
6CKR	CGA ATC	
7CKF	AGAA AGC	The primers were used to generate the control probe for the core probe (MRE) for EMSA.
7CKR	GCTTT CT	
12CKF	AGTTTT CTTGAA	The primers were used to generate the control probe for three core probes (MBS I -1, MBS I -2, MBS II) for EMSA.
12CKR	TTCAAGAAA ACT	

Note: The **bold** bases represent the MYB-related core elements (MBS1, CAACTG; MBS2, CGGTCA; MBS3, TAACTG; MRE, AACCTAA; MBS I , AAAAAAC(C/G)GTTA; MBS II , AAAAGTTAGTTA).

Supplementary Table S2. The conserved motifs of subgroup 4, 5, 6 and 15

Amino Acid Sequence	Motif Name	Subgroup	Reference
[D/E]Lx2[R/K]x3Lx6Lx3R	bHLH binding motif	4, 5, 6, 15	1,2
Lx3GIDPxTHRPI	C1 motif	4	3
LNL[E/D]L	C3/EAR/ERF motif	4	2,4
DNEI	DNEI motif	5	5
IRTKA[I/L]RC	Sg5 motif	5	6
DEDWLRxxT	Motif 5	5	7
KPRPR[S/T]F	Motif 6	6	8
VNNL[M/T][N/D]GDNMWLE	-	6	9
ANDV	ANDV motif	6	10
WVxxDxFELsXL	-	15	7
-W-(X ₁₉)-W-(X ₁₉)-W-.....-F/I/L/M-(X ₁₈)-W-(X ₁₈)-W-	-	4, 5, 6, 15	8

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- 3 Kranz, H. D. *et al.* Towards functional characterisation of the members of the R2R3-MYB gene family from *Arabidopsis thaliana*. *Plant Journal for Cell & Molecular Biology* **16**, 263-276 (1998).
- 4 Ohta, M., Matsui, K., Hiratsu, K., Shinshi, H. & Ohme-Takagi, M. Repression domains of class II ERF transcriptional repressors share an essential motif for active repression. *The Plant cell* **13**, 1959-1968 (2001).
- 5 Nesi, N., Jond, C., Debeaujon, I., Caboche, M. & Lepiniec, L. The *Arabidopsis* TT2 gene encodes an R2R3 MYB domain protein that acts as a key determinant for proanthocyanidin accumulation in developing seed. *The Plant cell* **13**, 2099-2114 (2001).
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- 9 Kranz, H. D. *et al.* Towards functional characterisation of the members of the R2R3-MYB gene family from *Arabidopsis thaliana*. *Plant Journal for Cell & Molecular Biology* **16**, 263-276 (1998).
- 10 Lin-Wang, K. *et al.* An R2R3 MYB transcription factor associated with regulation of the anthocyanin biosynthetic pathway in Rosaceae. *BMC plant biology* **10**, 1-17 (2010).

Supplementary Table S3. The promoter sequences of pathway genes which contains MYB-related elements

Gene Name	Promoter Sequence
SmC4H1	<p>TGTGAGTTAATGTTTTACCTTGGAATAAAAACAGCTACTTTTTTAAATAATAAACTCTCT GCCTGAATAAAGATTTGTGCAATATCCACACTTTTCTTCTTCAAATAAGCACCTTGTTT TGCTTTTGACTTTTTAAATTTGCATCGCAGTTTGAAAGAAGAAAAATAGACTGTTAGCT AGAAAGTGAACCTAACTTTACAAAATACTTGGAGGAAATGTCAGCGGAATTAGTTTA TCTCTTAAATAACATTTATACTACTCCATTTTTTATTTATTTTTTGTCTATCCTACTTATG ACGAAAATAGAAAAGTTAATTGTTATTTGAAGATGTGTTTGGTATAGATTTAATGTCGTA ATTCAGTTGCATTGAGTACTTTTGACTAAGTTTTGTTGAATAAATCCGACACATAAGATA TTGACGATGTTCTAGATTGGGCTAATTGTCCTAAATAGGAACTAGTACCCTAATTTGTAA TTACAATACAATTATCAAATATTCAAATTATAGACCATCTATTATTTTACAATTACACC ATTCCAAATTTTTTAAACAGACAAGACAAGCTATACTAAATTGGAACTATTAATAAAA AGATTGAATTATAATTACATTGTTTTGTGATTACAATTTATGATTAGTCTATGTTTTAAATA GTTGGCCCTTTCAAATAATGACGATCTCAACTACAACTTGTCAAAATCATGAGGTGG TTAGACATATATAGAAGGAGCATTTTACATATCAAATTCATGAGAGTGGCTCCAGAC ATAAATTTGAAAACAATTAATACCA<u>ACCTAA</u>TTATAAATGTGTAATTAATAAATACTTC AAATCTTGACTGGTTTAATCGTGAGAAGATTTACAAAACGTTGTGCTAGTATATGATATT GACTTAATATGTCTAGCGAACGACTAATTAAACTCAATTTTTTCAAATTGTTAATTTTATT ACTTATTGGCGTTTACCACGATTGATTATATGCATGATTGAGTATTTTTATTCTAAAGAGT AGGATTTCTCCAATTTACTTTTTCTATGGGATAAGAATCAAATAAAATTAGGTTGAATG ATAAAAATAATAAAGTATCTTAAAATATTTTCGAAGTTTCAATCCATTAATAATAAGACA TAGAGTCTATGTGTCTATGAAATGAACGACTTAAGAACGAGTAATAAATAATTAGATAA CTTCTTACAAATCCATCTCCAACCTACACGATACATTAAGTCCAATTTTCGTGAGTAAATG TATATCTCGGTAAGAGACAAAAGTTTTTTTTAATTTTTAAAATCAAATCTAGGTCNCTTT TTATAGTGTGTGCGCATTGTAATCGTGGTGCAGTAGTTGTCACAAATTTGTAAGGCGG GGAAGCACCGAATTTTTTTTTATCAAGAAATACACGTTTTTCATTGAGAGTTAGTTGTAG CACAAGCAATCCTACAACGTTTTTACAAAATATATCCATTATATAATACTCCCTTCGTCC ACCAAAGATATGCCACAATTTCTTTTTTCGTCCGTTACAAAAAATATATCACATTCATT TTTAGTAATAGAGTCTACACCATTTCATTACATTTAAAATGAGATCCTTACTCCACTAC CAACTTCACTCACATCTTATTAAAACTCGTGCCGAAAGTAAAGTGTGCATATTTTTTGTG GACGGAGGGAGTAGGTTATATTTTTCAAAGCCTCCAGCCATTCTATATCTGATGTGTCGT TCTCAATAATCTTCATTCTTTTTTCTAATTTCTACAATTATTCATTTTTAATAATTATTA ATATACATATATTTTAAAAACTTTCATTTTCAGTGTCTTTTATGTTTGTAAATTCACCTT TTGGAAAATGATTTCTTTAATTGTTATTATTCTAATTATTCATTAATATTTATCAATCAAAT AATTAATTAGCACGTTAATATTAATTACATTAATAAATTTTTATTTTCTATACTATTTTTTCA CTATAAATATAGAAATTTGCAACCAATTTTCATCCACATTAAGTTCTTTATGCATCTTCAT TCGCTTTTCTATCCAGCGACTCAAAATGCTATAATATATTGTATTATGTGGTAGCAAAT AAGATGATTCTAAAAGTCTCAAGAGAAAAAGAATTTTTTAAAATAAAGAAATCAAG GATTCTTATTGAATCAGAAGCTTCTCTCAACTTTTTTTTTTTTTTTTTTTTGGAGAGTGAA GCAGAACTCTTAGTTGCTTACATTGTAACGAAGTACTCCTTCCGTCTCAAGAAAAT TGCATATTTCTTTTTGAGTTATCCCAACGAAAGTGGTGCATTTTTTTATAGCAAAAATTA ATAAATTTAAAACTTAATTGACAAAATAAATGCACCCTTATTTTTTTTTAAATTAATTA AATTAACACACACTCTCTCTCTTCTATCTTCCCCATGTCTCTCTAAAACAATCCATTTGG ATGAAAGAACAAAAGCAGCCACCACCGTGGCCGCTGCCGCCCTCATCTCTGGCGC CTCCCCTGCCGTGCTGGAGCCGCGAGTTGACGTCGTCTTCCCCCGCCGTGCTAGGGT TTCGATTTGGGGTGGGTGCAGGGAGCAGTTGGGGATTGGGGCTTGGCGATGCCAGCG GCCTCAGCTACAGGTCACGGACGAGAAAGGTTAGAAAAGCAGATAAGGTGGTTTTTTG CGGCGATTTATCAAAAATCGAAAGAGAAGAAATTATGAAGTTTAAAAGTTAGGGAT TTTGTATATGTGTTTCGATTTGAGAGATTGTCATTTAAAGTAGCGGTGGCTGAAAGTG</p>

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A

Sm4CL2

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A

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TTCAACACACACCCTTTTAAATTTTAAATCATATTTATCGGTGACTATATTAGACTCTACC
TCAAGTGAGGCAAGCTCATCCGCAGCAGCC

Note: The **bold** bases represent the MYB-related core elements (MBS1, CAACTG; MBS2, CGGTCA; MBS3, TAACTG; MRE, AACCTAA; MBS I , AAAAAAC(C/G)GTTA; MBS II , AAAAGTTAGTTA). The underlined bases were synthesized for EMSA.

Supplementary Table S4. The homologous genes of SmMYB36 in different species

Organism	Defline
<i>Physcomitrella patens</i>	Pp3c1_4970V3.2
<i>Sphagnum fallax</i>	Sphfalx0111s0046.1
<i>Selaginella moellendorffii</i>	109587
<i>Amborella trichopoda</i>	evm_27.model.AmTr_v1.0_scaffold00024.222
<i>Musa acuminata</i>	GSMUA_Achr11T23500_001
<i>Spirodela polyrhiza</i>	Spipo0G0167800
<i>Brachypodium distachyon</i>	Bradi1g60106.1
<i>Brachypodium stacei</i>	Brast02G211100.1
<i>Oryza sativa</i>	LOC_Os01g50110.1
<i>Panicum hallii</i>	Pahal.D02873.1
<i>Panicum virgatum</i>	Pavir.J00444.1
<i>Setaria italica</i>	Seita.4G086300.1
<i>Setaria viridis</i>	Sevir.7G295300.1
<i>Sorghum bicolor</i>	Sobic.001G340900.1
<i>Zea mays</i>	GRMZM2G000818_T01
<i>Aquilegia coerulea</i>	Aquca_011_00570.1
<i>Kalanchoe laxiflora</i>	Kalax.0006s0265.1
<i>Mimulus guttatus</i>	Migut.A01129.1
<i>Solanum lycopersicum</i>	Solyc10g055410.1.1
<i>Solanum tuberosum</i>	PGSC0003DMT400034372
<i>Eucalyptus grandis</i>	Eucgr.D02099.1
<i>Vitis vinifera</i>	GSVIVT01006275001
<i>Linum usitatissimum</i>	Lus10033438
<i>Manihot esculenta</i>	Manes.02G041300.1
<i>Populus trichocarpa</i>	Potri.006G221200.1
<i>Ricinus communis</i>	28226.m000839
<i>Salix purpurea</i>	SapurV1A.1360s0110.1
<i>Citrus sinensis</i>	orange1.1g028843m
<i>Citrus clementina</i>	Ciclev10009521m
<i>Citrus clementina</i>	Ciclev10026498m
<i>Carica papaya</i>	evm.model.supercontig_46.144
<i>Gossypium raimondii</i>	Gorai.004G196800.1
<i>Theobroma cacao</i>	Thecc1EG015933t1
<i>Arabidopsis lyrata</i>	488320
<i>Arabidopsis thaliana</i>	AT5G40330.1
<i>Boechera stricta</i>	Bostr.1460s0063.1
<i>Brassica rapa</i>	Brara.J01999.1
<i>Capsella grandiflora</i>	Cagra.3166s0078.1
<i>Capsella rubella</i>	Carubv10001990m
<i>Eutrema salsugineum</i>	Thhalv10014715m
<i>Cucumis sativus</i>	Cucsa.340790.2
<i>Fragaria vesca</i>	mrna07418.1-v1.0-hybrid
<i>Glycine max</i>	Glyma.05G061900.1
<i>Malus domestica</i>	MDP0000226215
<i>Medicago truncatula</i>	Medtr4g100720.1
<i>Phaseolus vulgaris</i>	Phvul.003G222400.1

<i>Prunus persica</i>	Prupe.5G065500.1
<i>Chlamydomonas reinhardtii</i>	Cre16.g677382.t1.1
<i>Volvox carteri</i>	Vocar.0011s0012.1
<i>Coccomyxa subellipsoidea</i>	12896
<i>Micromonas pusilla</i>	39026
<i>Micromonas sp</i>	73784
<i>Ostreococcus lucimarinus</i>	8749

Notes: The analysis was based on the Phytozome database (<https://phytozome.jgi.doe.gov/pz/portal.html>) and NCBI BLAST tool (https://blast.ncbi.nlm.nih.gov/Blast.cgi?CMD=Web&PAGE_TYPE=BlastHome). The **bold** represents the predicted orthologous genes of SmMYB36.