

1 **Precise spatio-temporal control of rapid optogenetic cell ablation**
2 **with mem-KillerRed in Zebrafish**

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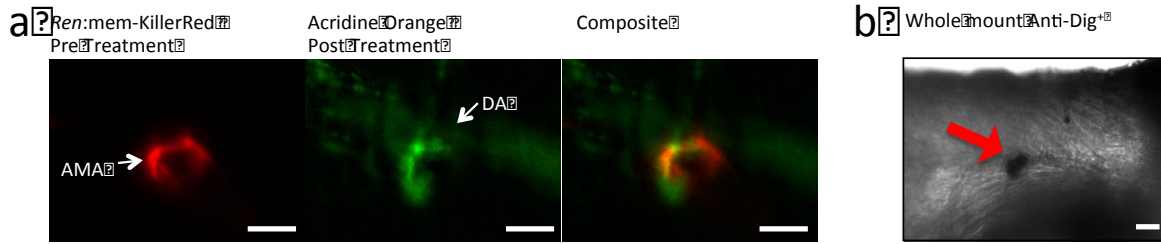
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27 **Supplementary Data**
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29 **Supplementary Figure 1:** Further staining to confirm cell death. (a) 3 dpf *ren:mem-KillerRed*
30 fish were incubated in acridine orange (2 ng/ml) for 30 mins in the dark, followed by two 5
31 mins washes and immediately imaged live on the SPIM. Treatment with the light-sheet was
32 performed (1 hr, 4 mW, depth-stack every 10s) and the acridine orange fluorescence signal
33 was collected post-treatment and a composite image was created. Treated KillerRed⁺ cells
34 can be seen to take up acridine orange readily. (b) A whole mount *in situ* protocol was
35 performed with an anti-Dig⁺ antibody, visualised using the colourant Nitro Blue Tetrazolium
36 using the brightfield camera on the SPIM. Dig⁺ cells are dark spots, as indicated with the red
37 arrow. Scale bars represent 30 μ m.

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40 **Supplementary Video 1:** SPIM-illuminated KillerRed⁺ targeting in live larval zebrafish.
41 Representative example of a 3 dpf *ren:mem-KillerRed;kdr:GFP* fish illuminated using the
42 SPIM light-sheet with a power of 3 mW (561 nm laser, 1 hr, depth stack acquired every 10 s).
43 Images were acquired axially every 1 μ m across the dorsal aorta (DA) and KillerRed⁺ AMA
44 region, across approximately 80 μ m in depth. Fish were embedded in 0.5% agar and
45 suspended between the perpendicularly-arranged objective lenses using FEP tubing. A
46 maximum intensity projection of the depth stacks was performed for each timepoint, and form
47 the frames of the time-lapse movie presented at 10 images per second. The scale bar
48 represents 30 μ m.

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