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RNAi	rde-1(+)	rde-1(-)	Mesoderm- rde
lin-3	13/17 Vul		
	4/17 Partial Induction	20/20 Normal	10/28 Vul
	1/56 Normal		51/62 Normal
lin-39	13/56 Partial induction	58/58 Normal	10/62 Partial induction
	42/56 Vul		1/62 Vul

gap-1(-)



WormMart	# Genes	% Conserved	<i>p</i> -value
Total genes in RNAi library	16,256		
Conserved genes in RNAi library	7,670	47.2	
Identified mesodermal genes	39		
Conserved mesodermal genes	31	79.5	0.00007

Figure S1 related to Figure 1 and Figure 3. Mesodermal RNAi in *C. elegans* identifies conserved factors. (A) Schematic of vulva development. Top: circles depict VPCs and anchor cell (AC). Gonad is in gray. Middle: during larval L3 stage, signaling pathways frequently mutated in human cancer induce VPCs to divide to form the 22 cells of the mature vulva, a DIC image of which is shown at the mid-larval L4 stage (bottom). A, anterior; P, posterior; V, ventral; D, dorsal. Reproduced / adapted with permission from Green et al., Development, 2007. (B) DIC (top) and fluorescence (bottom) images of larval L3 transgenic worms expressing GFP driven by tissue-specific promoters active in the anchor cell (arrowhead), somatic gonad, muscle, and vulval cells. Dashed lines indicate worm body boundaries. (C) RNAi sensitivity was enhanced by introducing the *rrf-3* mutation (*rrf-3(-*)). Introduction of the *rrf-3(-*) allele did not inadvertently alter RNAi resistance of rde-1(-) strains because rrf-3(-); rde-1(-) double mutants were viable upon RNAi-mediated depletion of two essential genes, pop-1 (TCF/LEF transcription factor) and *pie-1* (CCCH zinc finger protein). Embryonic lethality was evaluated in two independent *rrf-3(-); rde-1(-)* double mutant isolates, *H* and *K*. N2, wild-type. (D) Summary of Vulvaless phenotypes (Vul) after RNAi-mediated depletion of *lin-3* and *lin-39* in wild-type rde-1 worms (rde-1(+)), rde-1 mutants (rde-1(-)) and rde-1(-) mutants with mesoderm-specific RNAi (mesoderm-rde). (E) DIC and fluorescence images of wild-type gap-1 (gap-1(+)) and gap-1mutant (gap-1(-)) worms expressing a LET-60/RAS-dependent transcriptional reporter. Cell lineages are designated as P6.px (presumptive 1° cell fate) and P5.px and P7.px (presumptive 2° cell fate). Bottom panels are overexposed to better visualize CFP expression in P5.px and P7.px cells. (F) Percentage of genes with mammalian orthologs for all C. elegans genes included in the RNAi library or the 39 mesodermal genes identified by the RNAi screen. P=0.00007, Fisher's exact test.



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Chr.	RNAi	C. elegans	Human	O and Description				
#	Library ID	Gene	Ortholog	Gene Description				
	Chromatin Remodeling							
I	K07A1.11	rba-1	RBBP4	retinoblastoma binding protein 4				
11	ZK131.9	his-15	HIST2H2BF	histone cluster 2, H2bf				
	Y49E10.6	his-72	H3F3A/H3F3B	histone H3.3				
III	C07A9.3	tlk-1	TLK1/TLK2	tousled-like kinase 1/2				
IV	F55G1.10	his-61	HIST2H2AB	histone cluster 2, H2ab				
IV	T22D1.10	ruvb-2	RUVBL2	RuvB-like AAA ATPase 2				
V	F45F2.3	his-5	HIST1H4H	histone cluster 1, H4h				
V	C55A6.9	C55A6.9	PAF1	RNA polymerase II associated factor				
		Су	toplasmic Po	lyadenylation				
I	B0414.5	cpb-3	CPEB1	cytop. polyadenylation element binding protein 1				
V	F25G6.2	symk-1	SYMPK	symplekin				
			Translational	Regulation				
I	F26A3.2	ncbp-2	NCBP2	nuclear cap binding protein subunit 2, 20kDa				
	Y57A10A.30	ife-5	EIF4E	eukaryotic translation initiation factor 4E				
IV	F55F10.1	F55F10.1	MDN1	MDN1, midasin homolog (yeast)				
IV	Y51H4A.15	Y51H4A.15	TSR2	20S rRNA accumulation, homolog (S. cerevisiae)				
V	D1054.15	plrg-1	PLRG1	pleiotropic regulator 1 (PRL1 homolog, Arabidopsis)				
			Other Fur	nctions				
I	W02D9.1	pri-2	PRIM2	primase, DNA, polypeptide 2 (58kDa)				
II	K10G6.1	lin-31	FOXB1	Forkhead Box B1				
11	R53.3	egl-43	PRDM16	PR domain containing 16				
11	ZK430.1	toe-1	HEATR1	HEAT repeat containing 1				
П	C56E6.1	abcx-1	ABCG5	ATP-binding cass., sub-family G (WHITE), mem. 5				
- 111	F58A4.4	pri-1	PRIM1	primase, DNA, polypeptide 1 (49kDa)				
Ш	T26G10.1	T26G10.1	DDX47	DEAD (Asp-Glu-Ala-Asp) box polypeptide 47				
IV	C37F5.1	lin-1	ELK3	ETS-Domain Protein (SRF Accessory Protein 2)				
IV	H34C03.2	H34C03.2	USP4	ubiquitin specific peptidase 4				
V	F22F7.2	F22F7.2	SCCPDH	saccharopine dehydrogenase (putative)				
V	W02H5.4	W02H5.4	POGK	pogo transposable element with KRAB domain				
V	H14N18.1	unc-23	BAG2	BCL2-associated althanogene 2				
V	ZK856.10	ZK856.10	POLR3H	pol (RNA) III (DNA directed) polypep. H (22.9kD)				
X	F13D11.2	hbl-1	ZNF124	zinc finger protein 124				
X	K09E2.1	K09E2.1	COL6A6	collagen, type VI, alpha 6				
X	T27B1.2	ztf-19	PRDM10	PR domain containing 10				
Nematode-Specific								
IV	F38A5.6	F38A5.6	N/A	Unknown				
IV	F38E11.9	F38E11.9	N/A	Unknown				
IV	C43F9.4	C43F9.4	N/A	Unknown				
IV	VV02A2.8	WU2A2.8	N/A	Unknown				
V	Y45G12C.14	srj-19	N/A	serpentine receptor, class J				
V	ZK697.4	sri-27	N/A	serpentine receptor, class I				
V	20317.6	20317.6	N/A	Unknown				
Х	C24H10.1	C24H10.1	N/A	claudin homolog				

Figure S2 related to Figure 2. Genome-wide RNAi screen identifies conserved mesodermal factors that suppress ectopic epithelial cell division. (A) Experimental workflow of the genome-wide mesoderm-specific RNAi screen. (B) The 39 genes identified by mesoderm-specific RNAi screen are grouped based on annotated functions. Chr., chromosome.





Figure S3 related to Figure 4. In vivo deletion of Tlk1/2 in mouse mammary fibroblasts increases the proliferation of adjacent epithelial cells. (A) Examples of sections from Fspcre:  $Tlk1^{F/F}$ :  $Tlk2^{F/F}$  ( $Tlk1^{\Delta/\Delta}$ :  $Tlk2^{\Delta/\Delta}$ ) mice immune-stained with Ki67 or smooth muscle actin (SMA) and counterstained with hematoxylin (blue) (top, bar =  $100 \mu m$ ; bottom, bar =  $50 \mu m$ ). (B) Representative images of sections of inquinal mammary gland from *Tlk1<sup>F/F</sup>:Tlk2<sup>F/F</sup>* mice (control),  $Tlk1^{\Delta/\Delta}$ ,  $Tlk2^{\Delta/\Delta}$  or  $Tlk1^{\Delta/\Delta}$ ;  $Tlk2^{\Delta/\Delta}$  mice immuno-stained for cleaved caspase-3 (CC3: brown) and counterstained with hematoxylin (blue) (bar = 50  $\mu$ m). Boxes indicate area of magnification in adjacent panel (bar =  $20 \mu m$ ). (C) Number of CC3 positive cells per duct (dots) of tissue described in (B). Red bar, mean; black bars, S.D.; pooled counts from 2 mice of each genotype; n.s., not significant. P value: Student's t-test. (D) Left: Bright field images of whole mount inquinal mammary glands from 10-week-old Tlk1<sup>F/F</sup>:Tlk2<sup>F/F</sup>(control). Fsp-cre:Tlk1<sup>F/F</sup>  $(Tlk1^{\Delta/\Delta})$  and  $Tlk2^{\Delta/\Delta}$  mice stained with carmine red (bar = 2000 µm). Boxes indicate area of magnification in adjacent panel (bar =  $100 \,\mu$ m). Middle: H&E stained sections of opposite inguinal gland (bar = 50  $\mu$ m). Right: representative images of sections of inguinal mammary glands immuno-stained for Ki67 (brown) and counterstained with hematoxylin (blue) (bar = 50  $\mu$ m). Boxes indicate area of magnification in adjacent panel (bar = 20  $\mu$ m). (E) Percent Ki67 positive epithelial cells per 20X field (dots) of tissue sections described in (A). Red bar, median; black bars, interquartile range; pooled counts from 6 control mice or 3 and 5 mice for the  $Tlk1^{\Delta/\Delta}$ and  $Tlk2^{\Delta/\Delta}$  groups respectively. P value: Mann-Whitney test of medians. (F) Quantification of phospho-ERK (pERK) positive epithelium from both cytoplasmic and nuclear signal per 20X field (dots). Red bar, median; black bars, interguartile range; pooled counts from 3 control or 5  $Tlk1^{\Delta/\Delta}$ :  $Tlk2^{\Delta/\Delta}$  mice. P value: Mann-Whitney test of medians.





**Figure S4 related to Figure 4 and Figure 5.** *Fsp-cre* transgene expression in various tissues. (A) Organs and glands were isolated from the fsp –cre line possessing only the RosaloxP reporter, fixed and stained for conditional β-galactosidase reporter activity in situ. Staining of the stomach is from intestinal bacteria as evidenced by the control while minimal staining was observed in the liver, lung, and intestine with most if not all staining from connective tissue and associated with the vasculature. (B) In situ X-gal staining of skin from a Fsp-cre;Rosa<sup>+/loxP</sup> mouse, that was then paraffin embedded and sectioned. -cre expression was detected in every cell type in the skin: Adipocytes, Smooth Muscle, Epithelium and Stroma. (C) Higher (10x) magnification of wholemount mammary glands from Figure 4J from mice with the indicated genotypes; cross sections stained for β-galactosidase and counterstained with nuclear fast red (NFR, bottom panels). Str, stromal fibroblasts; lu, lumen; epi, epithelial cells.





Cytolplasmic Nuclear

Figure S5 related to Figure 5. In vivo deletion of Sympk in mouse mammary fibroblasts increases proliferation of adjacent ductal epithelium. (A) Left: bright field images of whole mount inquinal mammary glands from 10-week-old Svmpk<sup>F/F</sup> mice (control) or Fsp-cre:Svmpk<sup>F/F</sup> (Sympk<sup> $\Delta/\Delta$ </sup>) mice stained with carmine red to emphasize ductal branching (bar = 1000 µm). Boxes indicate area of magnification in adjacent panel (bar = 100 µm). Middle: H&E stained sections of opposite inquinal gland (bar = 50  $\mu$ m). Right: representative images of sections of inguinal mammary gland immuno-stained for Ki67 (brown) and counterstained with hematoxylin (blue) (bar = 50  $\mu$ m). Boxes indicate area of magnification in adjacent panel (bar = 20  $\mu$ m). (B) Percent Ki67 positive epithelial cells per 20X field (dots) of tissue sections described in (A). Red bar, median; black bars, interguartile range; pooled counts from 6 control mice or 7 mice for the Fsp-cre group. P value: Mann-Whitney test of medians. (C&E) Representative images of sections of inguinal mammary gland from 10-week (C) or 6-month-old (E)  $Sympk^{F/F}$  or  $Sympk^{\Delta/\Delta}$ mice immuno-stained for cleaved caspase-3 (CC3; brown) and counterstained with hematoxylin (blue) (bar = 50  $\mu$ m). Boxes indicate area of magnification in adjacent panel (bar = 20  $\mu$ m). (D&F) Number of CC3 positive cells per duct (dots) of tissue described in (C) or (E) respectively. Red bar, mean; black bars, S.D.; pooled counts from 2 mice of each genotype. P value: Student's t-test. (G) Quantification of phospho-ERK (pERK) positive epithelium from both cytoplasmic and nuclear signal per 20X field (dots). Red bar, median; black bars, interguartile range; pooled counts from 4  $Sympk^{F/F}$  or 5  $Sympk^{\Delta/\Delta}$  6-month-old mice. P value: Mann-Whitnev test of medians. n.s., not significant.



**Figure S6 related to Figure 6. The human ortholog signature fails to distinguish breast tumor epithelium from adjacent normal epithelium.** Heatmap displaying RNA expression of the 33 human orthologs in normal (n=14) and tumor epithelium (n=40) from breast cancer patients. Each sample was given a rank for each gene and ordered among the experimental samples according to the rank-sums. The boundaries (dashed lines) represent the 95% distribution limits for rank-sums of 10,000 randomly generated samples. The P value indicates the ability of the gene signature to distinguish normal from tumor epithelium in breast cancer patients. Epi., epithelium. See also Table S4.





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Mutation status of breast cancer cell lines (x = no mutation)					
	RTK	Ras	Raf	NF1	
	DDR2 (V824A)				
	missense				
	EPHB6 (G403D)				
	missense				
	ERBB2 (G152fs)			(Q1033R)	
MDA-468	frame-shift	x	x	missense	
	EPHA5 (L1009P)				
	missense				
	ERBB4 (Y1242C)				
MCF-7	missense	х	x	x	
T47D	х	х	х	х	
	PDGFRA (Y172F)	KRAS (G13D)	BRAF (G464V)	(L466fs)	
MDA-231	missense	missense	missense	frame-shift	
BT474	х	х	х	x	
MCF10A	х	х	х	x	

## Figure S7 related to Figure 7. Characterization of cells used in human co-culture

**experiments (A)** 22 human orthologs were knocked down by lentirviral shRNA transduction as determined by real-time quantitative PCR analysis. Data are shown as mean percent residual gene expression ± standard deviation relative to non-silencing shRNA and normalized with *GAPDH* expression. **(B)** Fold increase in cell number at day 4 of culturing for breast cancer cell lines alone or co-cultured with shRNA control (shRNA-scr.) infected fibroblasts (Fib.). n=3 independent experiments with each experiment performed in triplicate. Data are presented as average ±SEM. See also Table S3. **(C)** Mutation status of breast cancer cell lines from Broad-Novartis Cancer Cell Line Encyclopedia (CCLE) https://portals.broadinstitute.org/ccle/home.