

Supporting Information Legends

Figure S1. Nuclear distribution of TE-lincRNAs and non-TE-lincRNAs in Arabidopsis (A), rice (B), and maize (C). Each chromosome was separated into 100 kb bins. The lines on top of each chromosome represent frequencies of TE-lincRNA (red) and non-TE-lincRNAs (green) in each bin. The heatmaps at the bottom of each chromosome represent the frequencies of TEs (red) and genes (blue) in each bin. **(D)** Pairwise correlations of densities of TE-lincRNAs, non-TE-lincRNAs, genes and TEs.

Figure S2. Length distribution of TE-lincRNAs and non-TE-lincRNAs identified from Arabidopsis (A), rice (B) and maize (C).

Figure S3. Exon numbers of TE-lincRNAs and non-TE-lincRNAs identified from Arabidopsis (A), rice (B) and maize (C).

Figure S4. Numbers of RNA motifs detected in TE-lincRNAs and non-TE-lincRNAs from Arabidopsis (A), rice (B) and maize (C).

Figure S5. Distribution of the distance of lincRNAs to their corresponding nearby genes in Arabidopsis (A), rice (B) and maize (C). X axis represents the relative distance of lincRNAs with respect to the coordinate of the 5' end (Sense) or 3' end (Antisense) of neighbouring genes.

Figure S6. Correlation of expression between lincRNAs and 10 closest genes and the example of lincRNA (A) and protein-coding gene co-expression network showing stress response (B and C). (A) Pearson's correlation of the expression between lincRNAs and 10 closest genes across 15 different samples. Red bar in the middle represents TE-lincRNAs, and black bar represents non-TE-lincRNAs. Grey colour in the heatmap represents non-significant correlation (p -value ≥ 0.05). (B) Expression pattern of lincRNAs and protein-coding genes in module/sub-network "paleturquoise" is shown in the top panel, and the bar plot in the bottom panel shows the eigengene values for different samples. Green represents "under-expressed" and red represents "over-expressed" in the heatmap. The "eigengene value" is defined as the first principal component of this module, so it can be considered as the representative of the gene expression profiles in this module. (C) Visualization of co-expression module/sub-network "paleturquoise". And label(s) in black colour represent protein-coding genes and label(s) in blue colour represent TE-lincRNA. The size of the node/label and edge weight is proportional to betweenness centrality.

Figure S7. Contribution of TEs to lincRNAs in Arabidopsis, rice and maize. (A) Distribution of numbers of TEs in TE-lincRNAs. **(B)** TE-lincRNAs with multiple TEs have longer sequences. **(C)** Distribution of coverage of lincRNAs covered by TEs in TE-lincRNAs. **(D)** Distribution of coverage of TEs overlapped by TE-lincRNAs. **(E)** Positional distribution of TEs in TE-lincRNAs with respect to their host TE-lincRNAs. X axis represents the relative coordinates of the 5' end of TEs compared to the 5' end of TE-lincRNAs, and the Y axis represents the relative coordinates of the 3' end of TEs compared to the 3' end of TE-lincRNAs. Therefore, each dot represents one TE-lincRNA. There is no dot located in the top left corner, because this region represents lincRNAs fully nested in TEs, which were filtered out by our pipeline.

Figure S8. Comparison of lincRNAs and protein-coding genes contributed by TEs in Arabidopsis, rice and maize. (A) Percentage of lincRNAs or protein-coding genes contributed by TEs. (B) Box plot showing difference of TE coverage in lincRNAs and protein-coding genes.

Figure S9. Mutated TE-lincRNA11195 alters sensitivity to ABA in seed germination and post-germination development in Arabidopsis. (A) Seed germination of WT, *lincRNA11195-1* and *lincRNA11195-2* mutants. Seeds (90 per genotype) were grown on half MS medium supplemented with different amounts of ABA. Seeds were vernalized at four degree for three days, and germinations of seeds were determined when they had grown for three days. Seeds were considered as germinated once radicle penetrated the seed coat. (B) Post-germination seedling development of WT, *lincRNA11195-1* and *lincRNA11195-2* under 0 or 1 μ M ABA. (C) Primary root length and fresh shoot weight of seedlings shown in B. Graphs are presented as the percentage relative to growth on control half MS media. ABA assays were performed by stratifying seeds at 4 °C for 3 days, seedlings were grown for eleven days on half MS medium supplemented with or without 1 μ M ABA. Error bars stand for standard deviation (n = 20).

Figure S10. Expression of Arabidopsis TE-lincRNA11195 in WT and an ABA-insensitive mutant *pyr1/pyl1/pyl4*. Transcription of an ABA responsive gene RD29A is also shown.

Figure S11. Mutated TE-lincRNA11195 alters sensitivity to salt in seed germination and post-germination seedling development in Arabidopsis. (A) Expression of lincRNA11195 and its neighbouring gene under different conditions. (B) Seed germination of WT, *lincRNA11195-1* and *lincRNA11195-2* mutants. Seeds (90 per genotype) were grown on half MS medium supplemented with different amounts of salt. (C) The percentage of green seedlings in B. Seedlings were grown on half MS medium with 150 mM NaCl for eleven days, and the number of seedlings with green cotyledons were measured. *: $p < 0.05$ (Two-sample Independent t test).

Figure S12. TE strengthens the ABA-responsive transcription to lincRNA11195 in Arabidopsis. The Col-0 lincRNA11195 DNA sequence with (lincRNA11195_C) or without (lincRNA11195_noTE) the LTR driven by its native promoter was introduced into *11195-1* (Right panel) and *11195-2* (Left panel), respectively. LincRNA11195 transcript levels after 12 h of 100 μ M ABA treatment were analysed by qRT-PCR, and WT used in this assay was that Col-0 plants transformed empty vector. The experiment was performed with T1 plants, and it was carried out with at least three biological replicates, and each replicate contained at least three seedlings.

Figure S13. TE-lincRNA11195 is detected only in Arabidopsis genus. PCR amplification of TE-lincRNA11195 was carried out in *Arabidopsis thaliana*, *Arabidopsis lyrata* and *Capsella rubella*. Template cDNA, RNA (no RT) or DNA were used in the reactions.

Figure S14. Sequence pairwise alignment and secondary structure prediction of TE-lincRNA11195 in *A. thaliana* and *A. lyrata*. (A) Sequence pairwise alignment of lincRNA11195 between *A. thaliana* and *A. lyrata*. Secondary structures of lincRNA11195 of *A. thaliana* and *A. lyrata* were shown in (B) and (C) respectively. Colour gradient represents base-pair probabilities.

Table S1. General information about RNA sequence libraries used in this study.

Table S2. Genomic coordinates of TE-lincRNAs in three species.

Table S3. Summary of Rfam RNA motifs detected in TE-lincRNAs and non-TE-lincRNAs from three species.

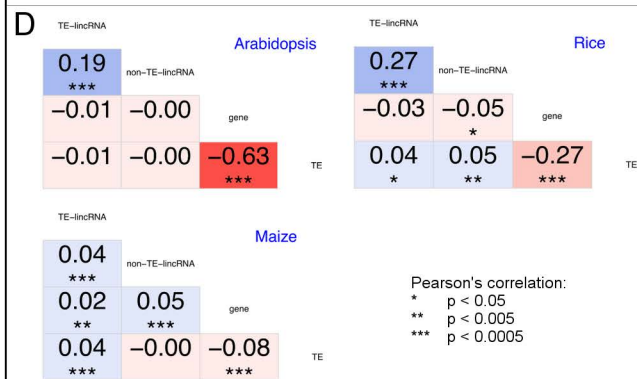
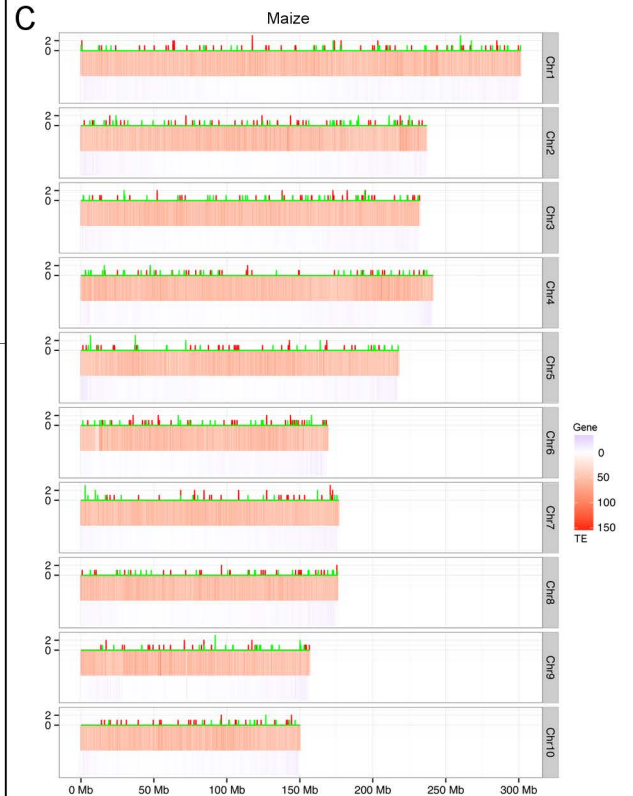
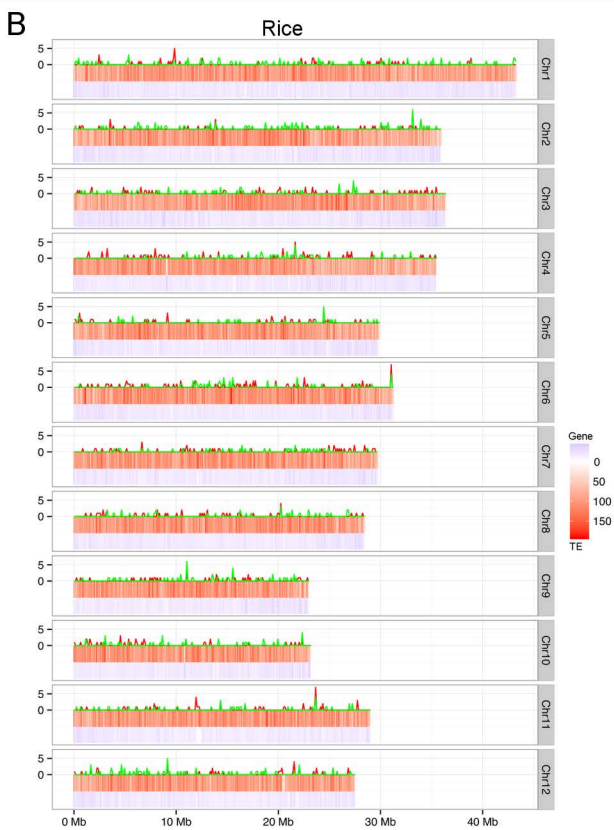
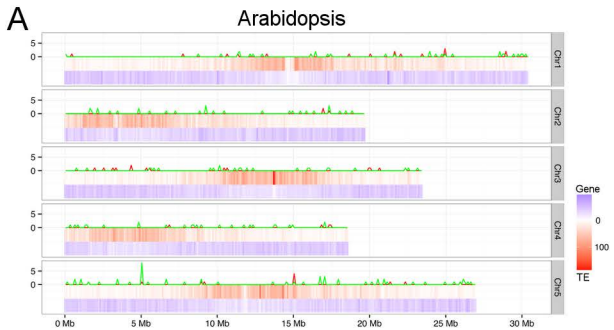
Table S4. Summary of co-expression network analysis in Arabidopsis.

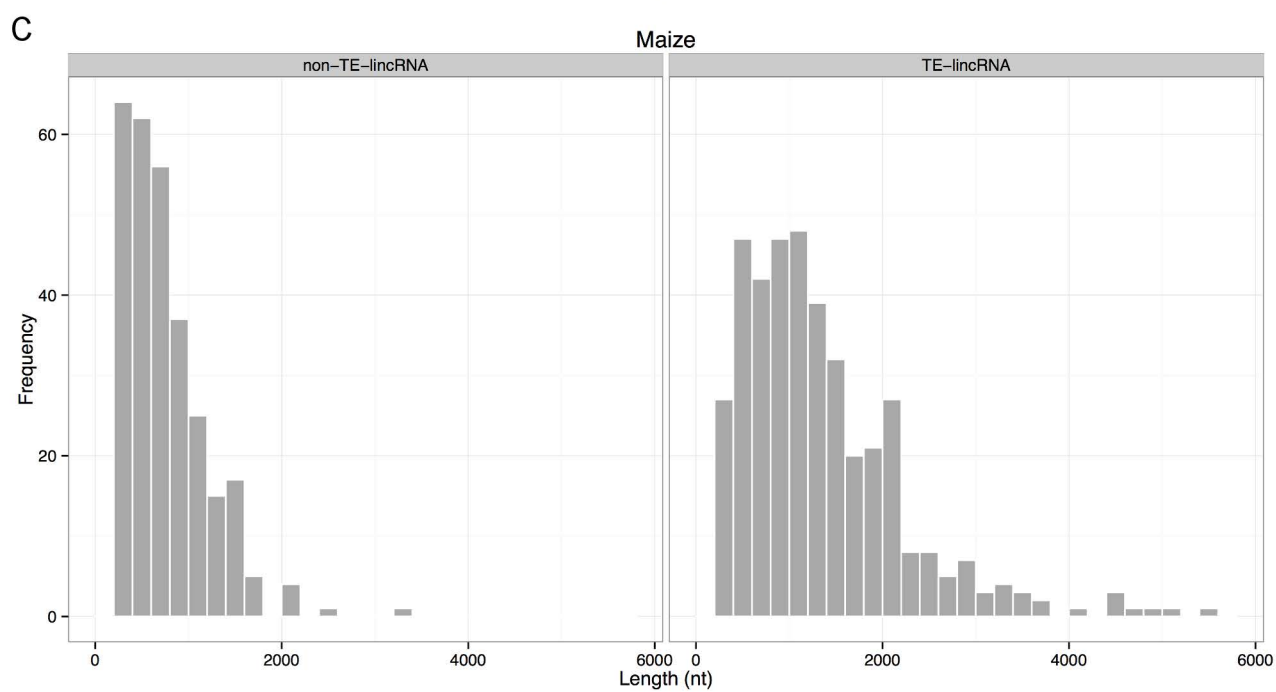
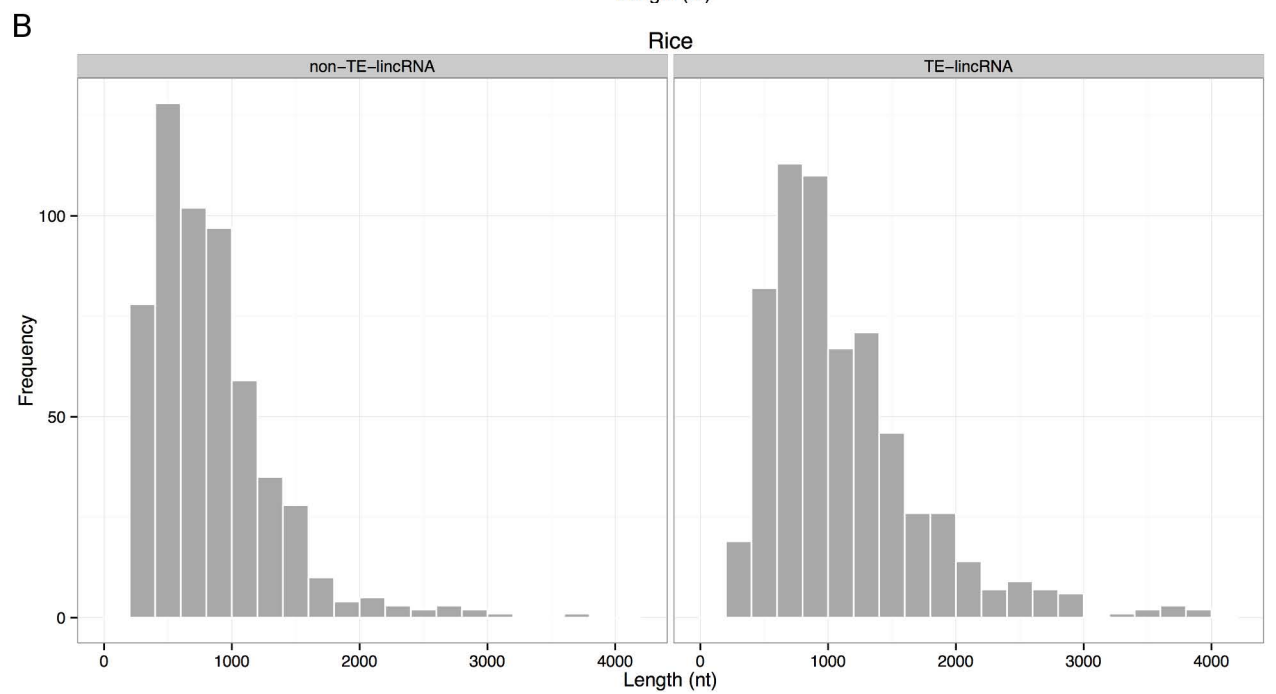
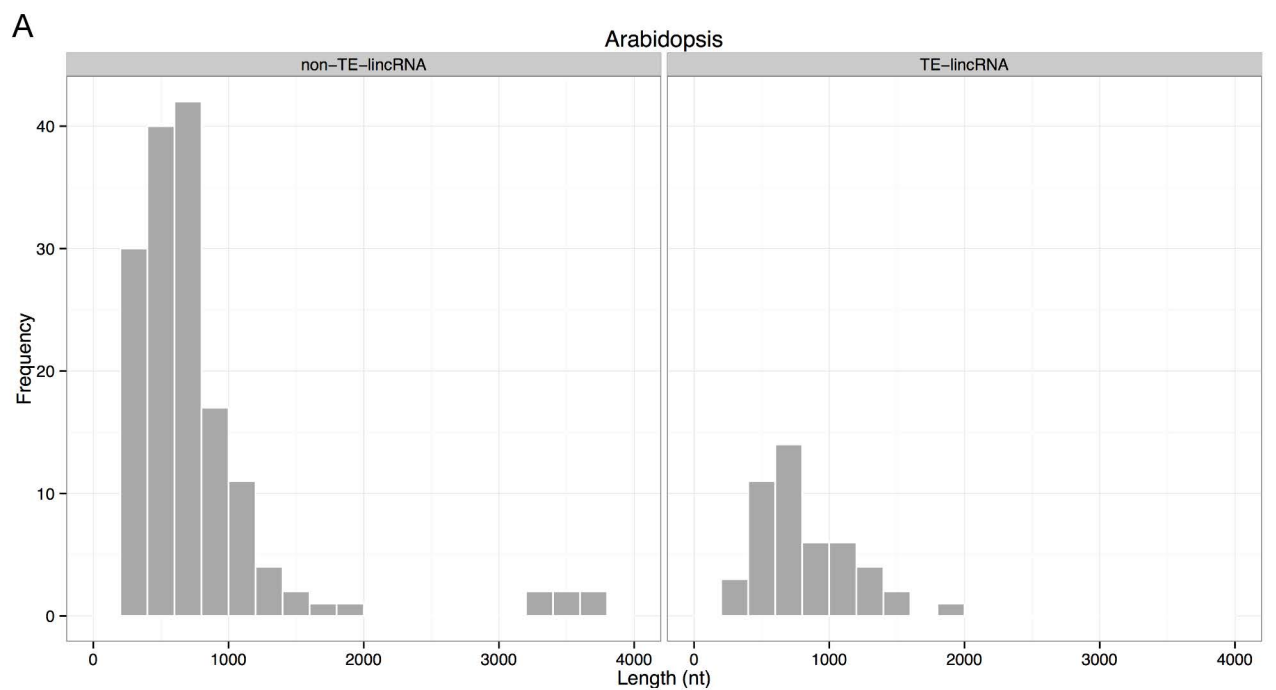
Table S5. Statistics of different TE families contributing to lincRNAs in three species.

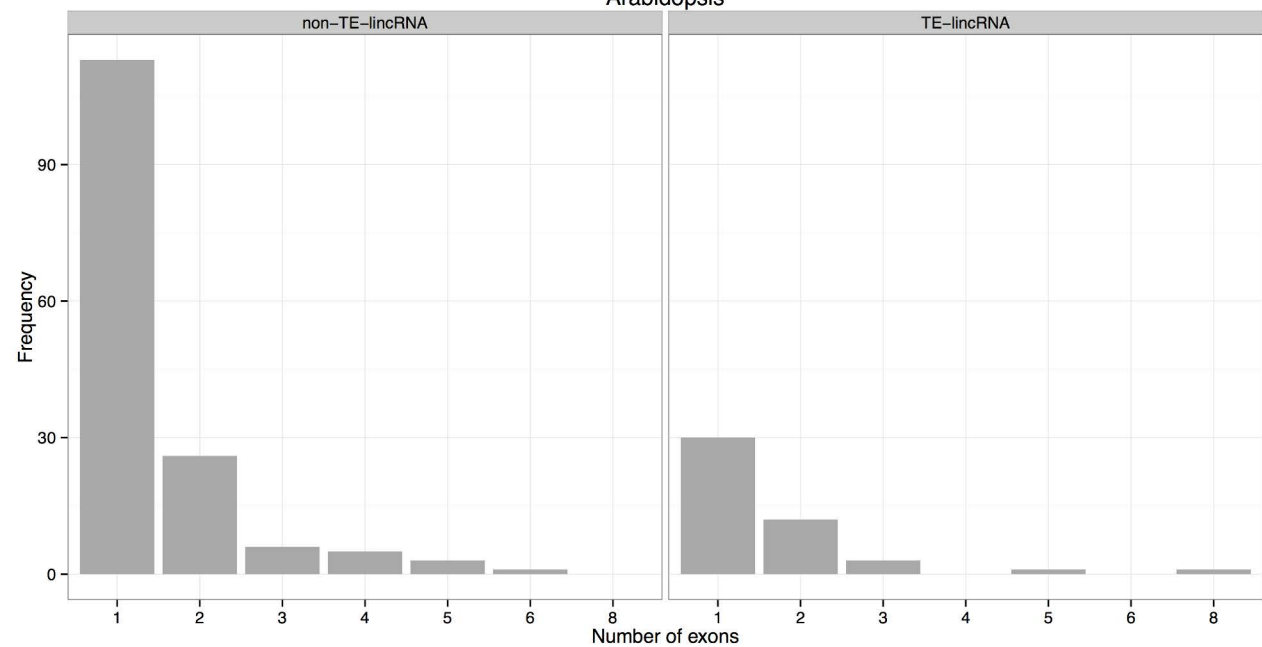
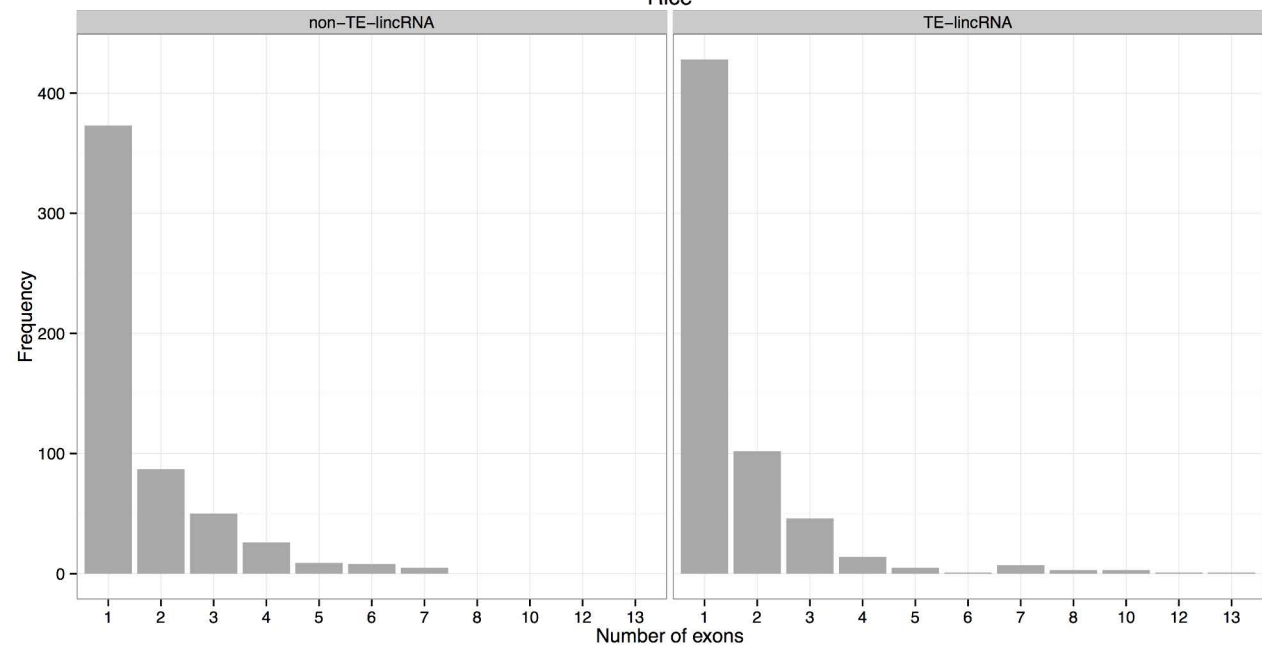
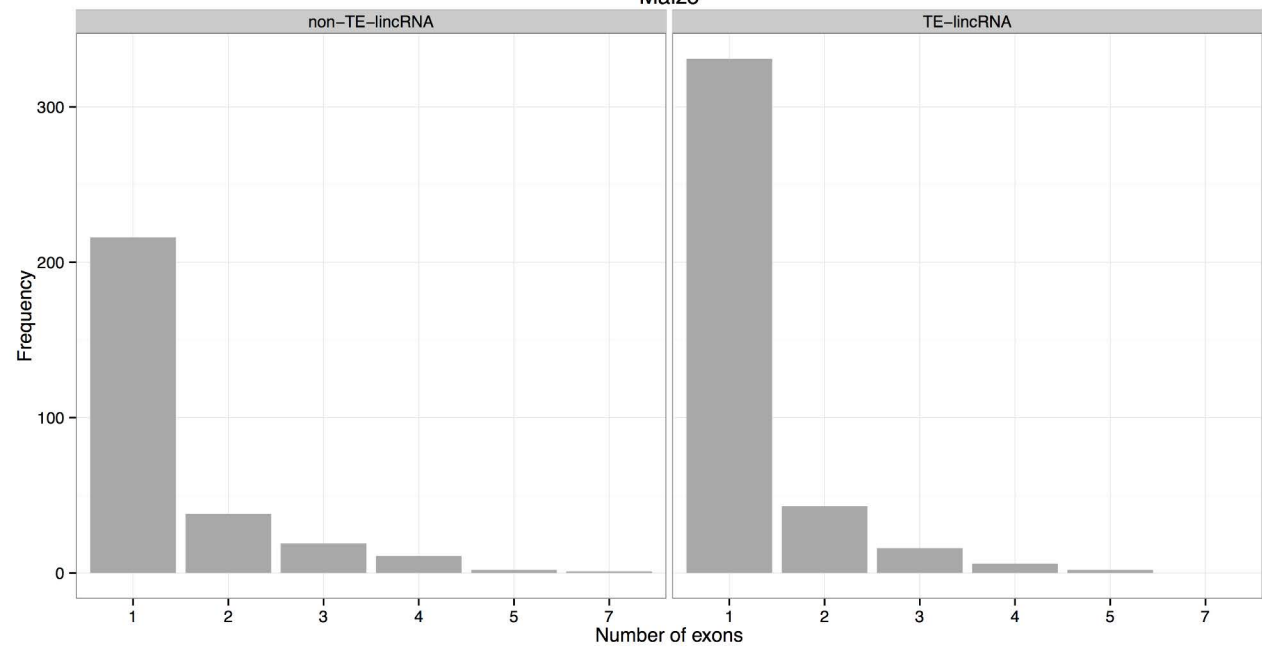
Table S6. Summary of differential gene expression between wild type and mutant *TE-linc11195-2*.

Table S7. Non-Mendelian inheritance of *ddm1* induced lincRNAs.

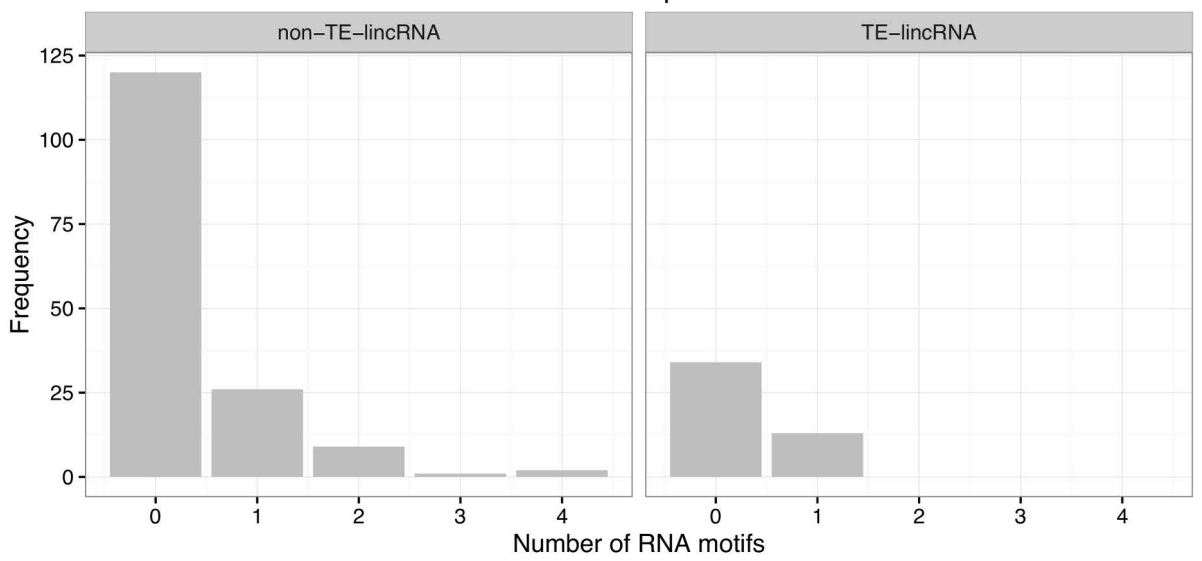
Table S8. Primers used in this study.



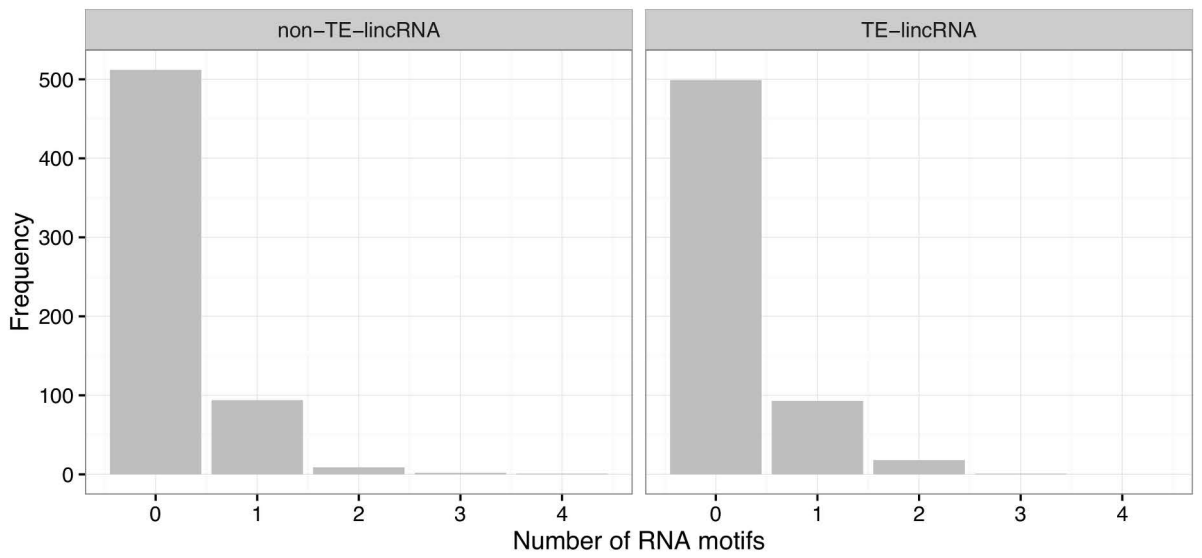


A**Arabidopsis****B****Rice****C****Maize**

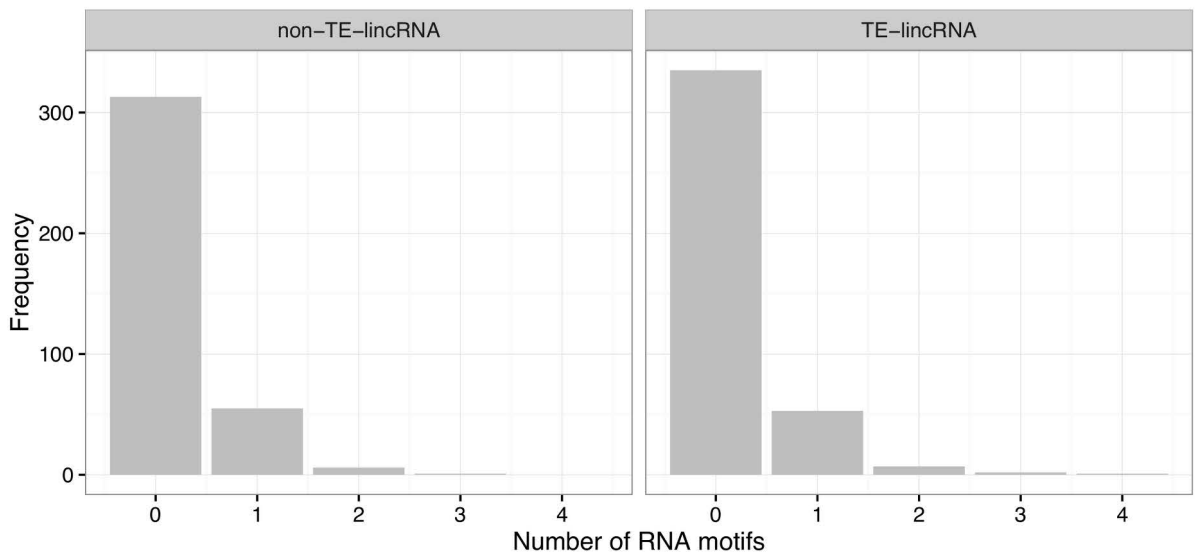
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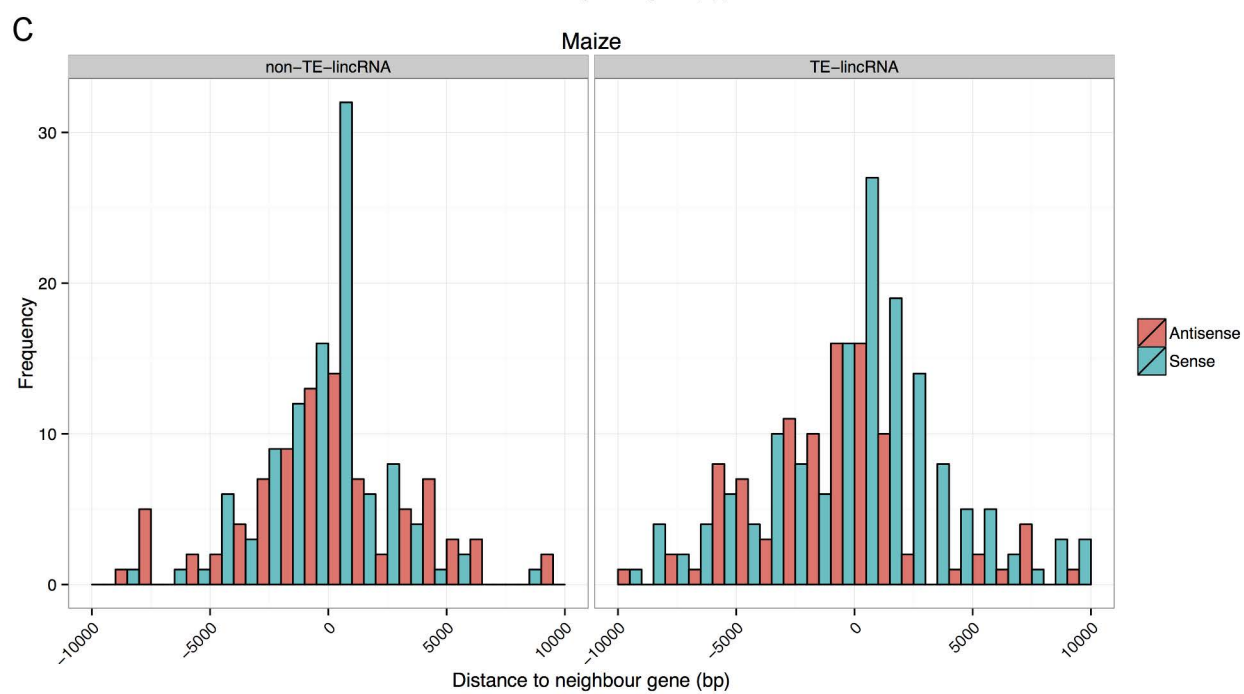
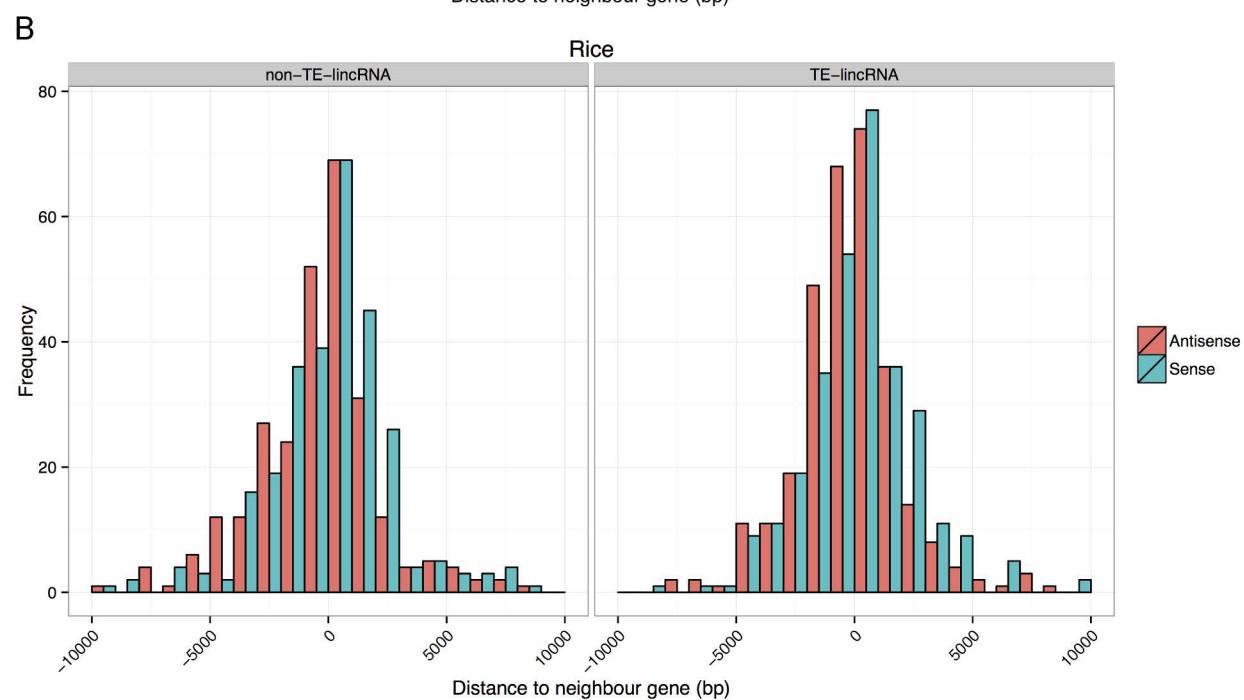
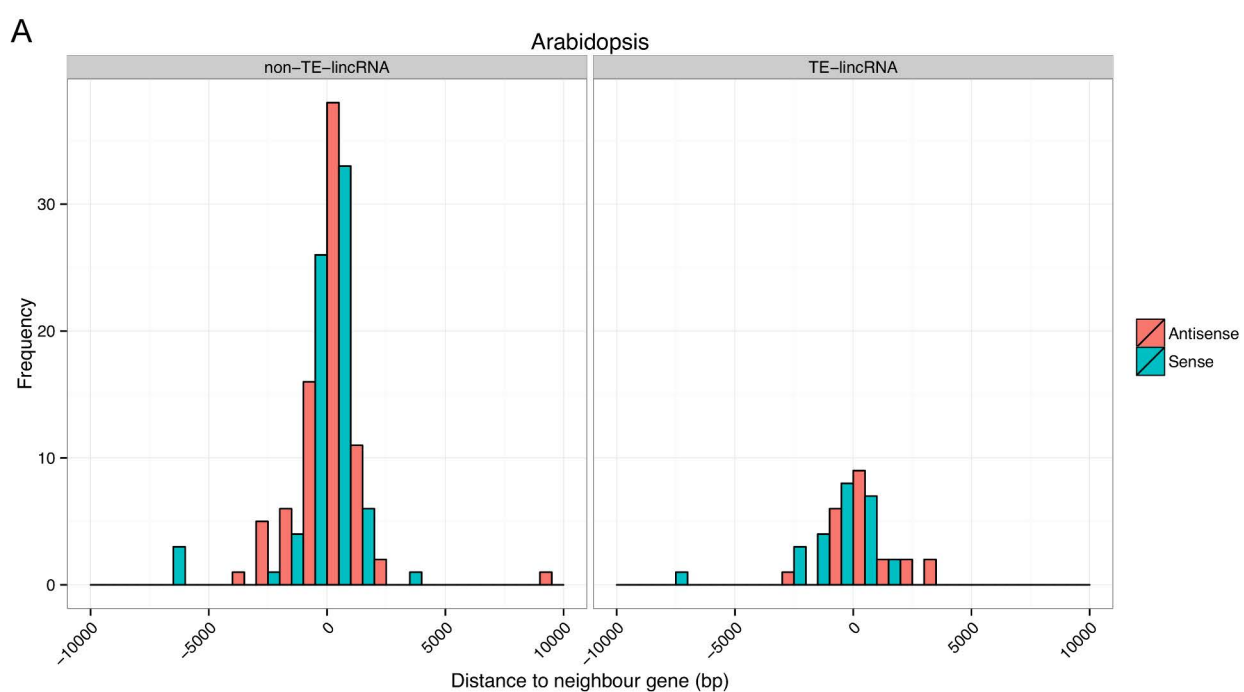


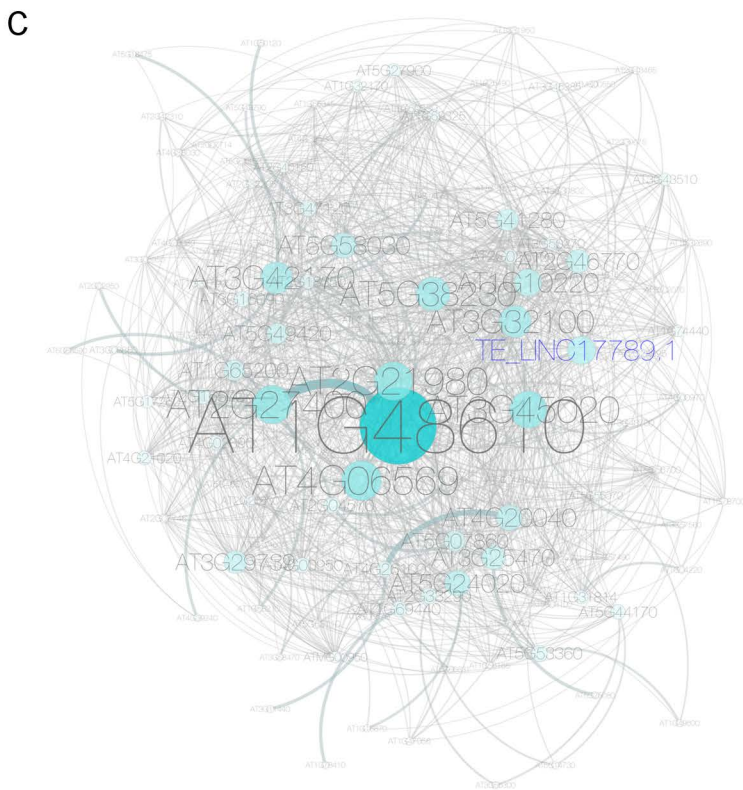
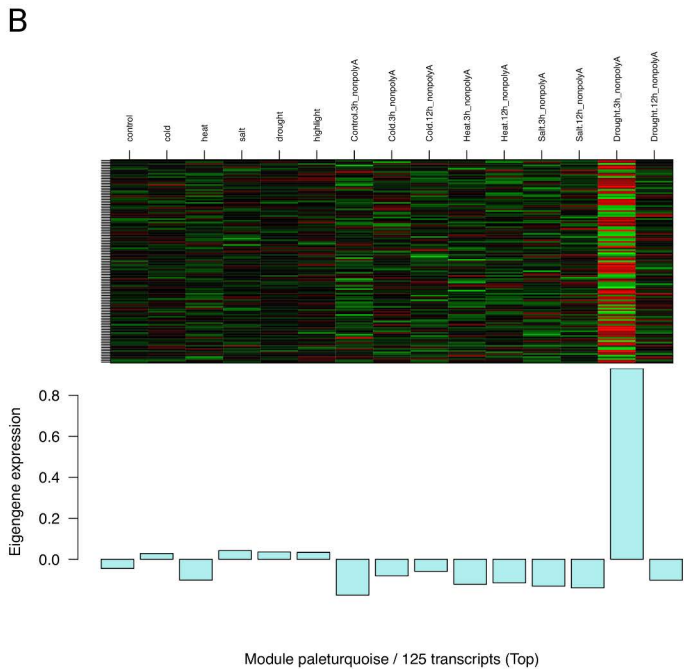
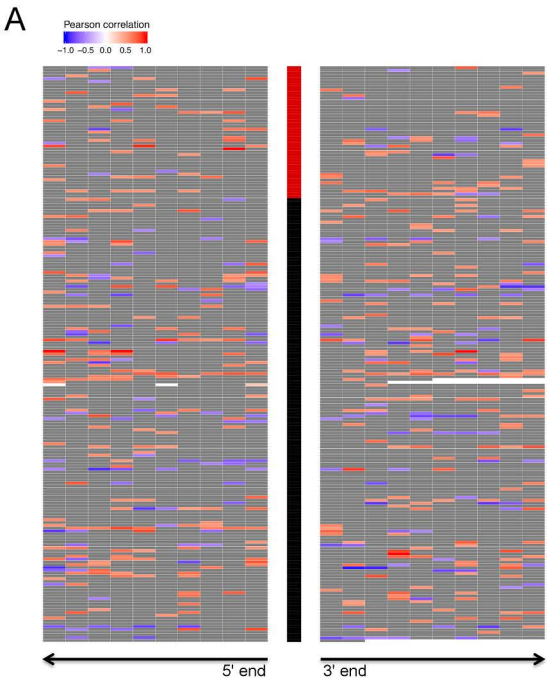
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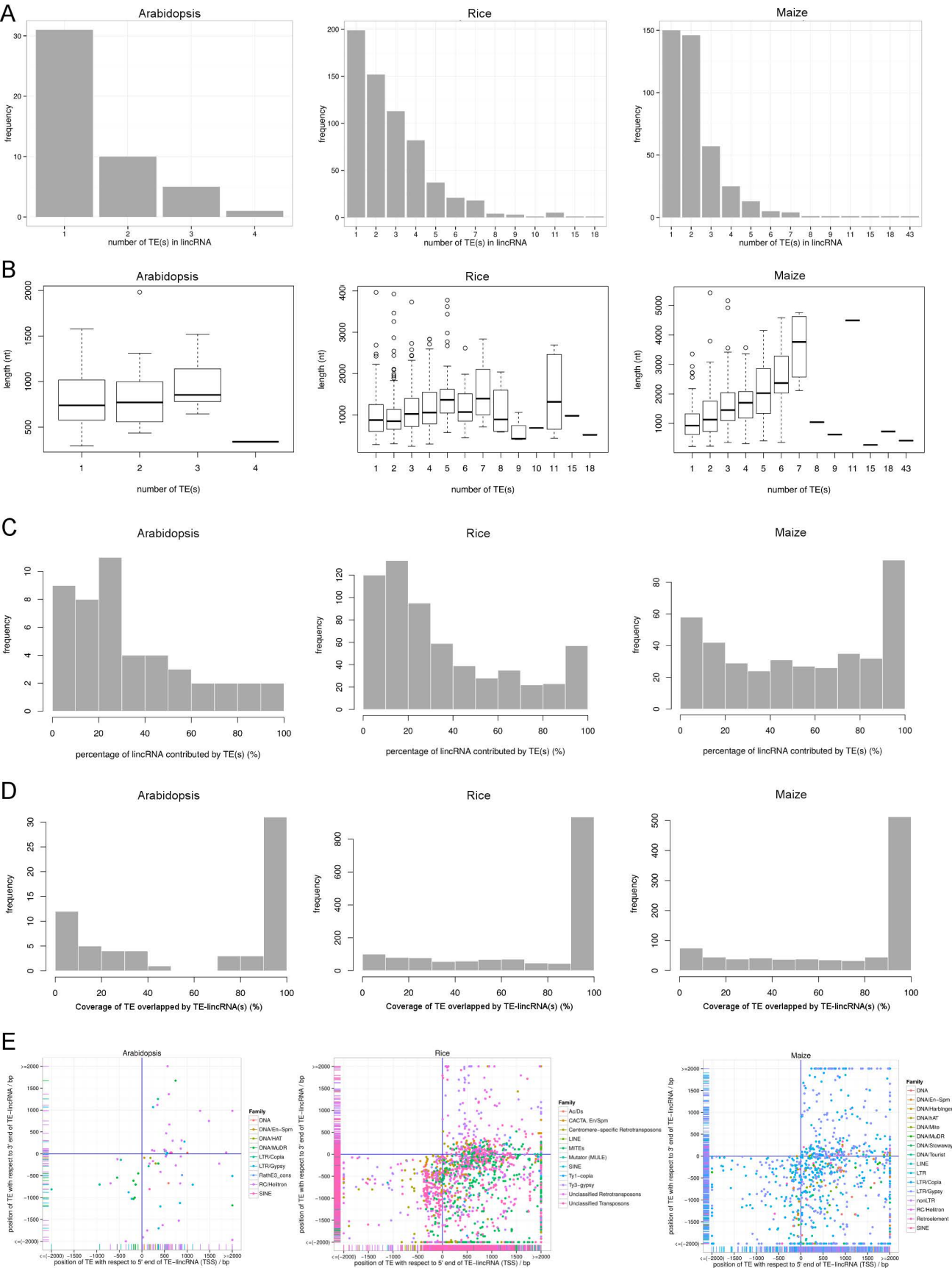


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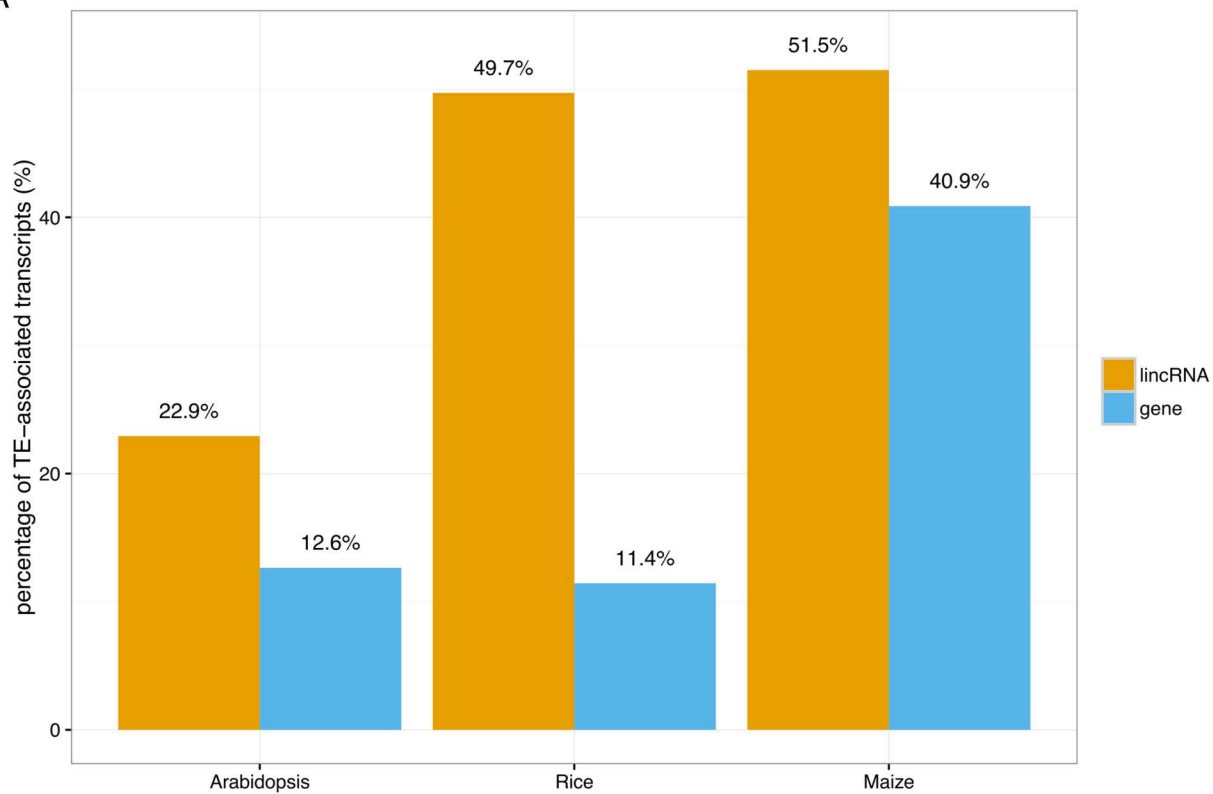




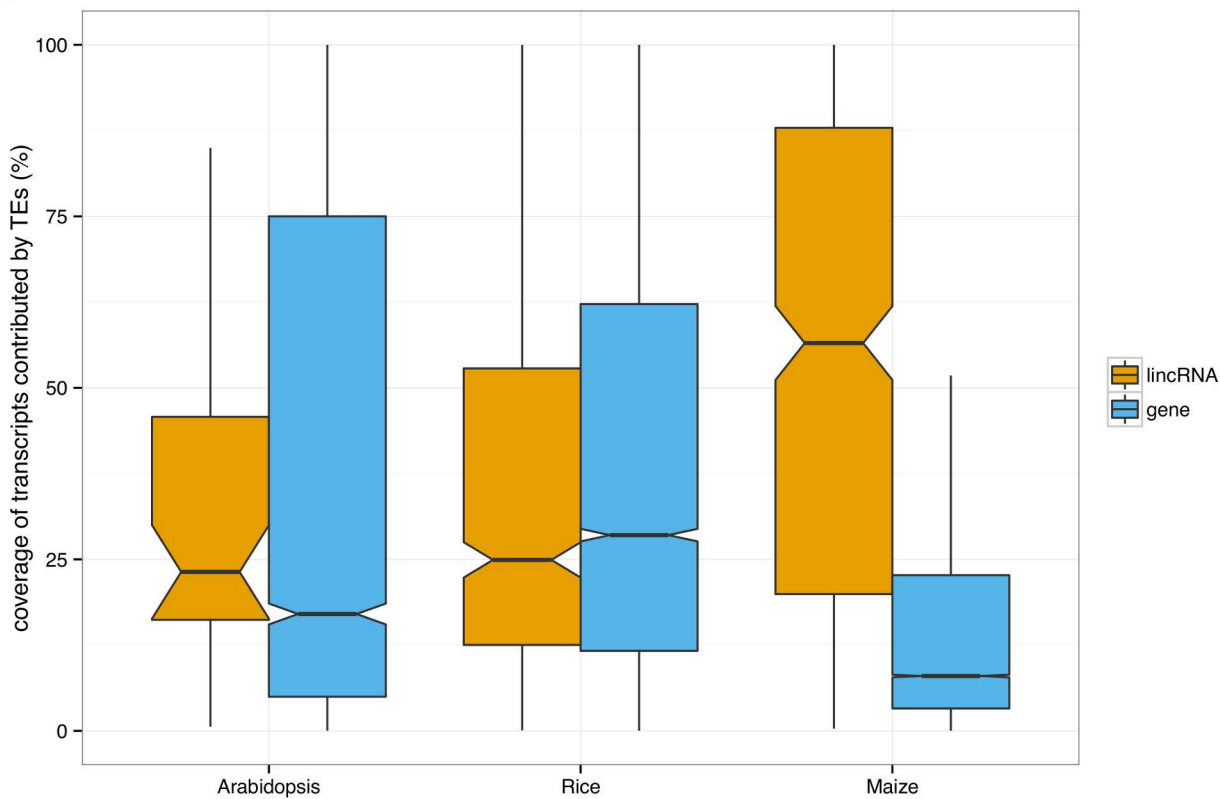




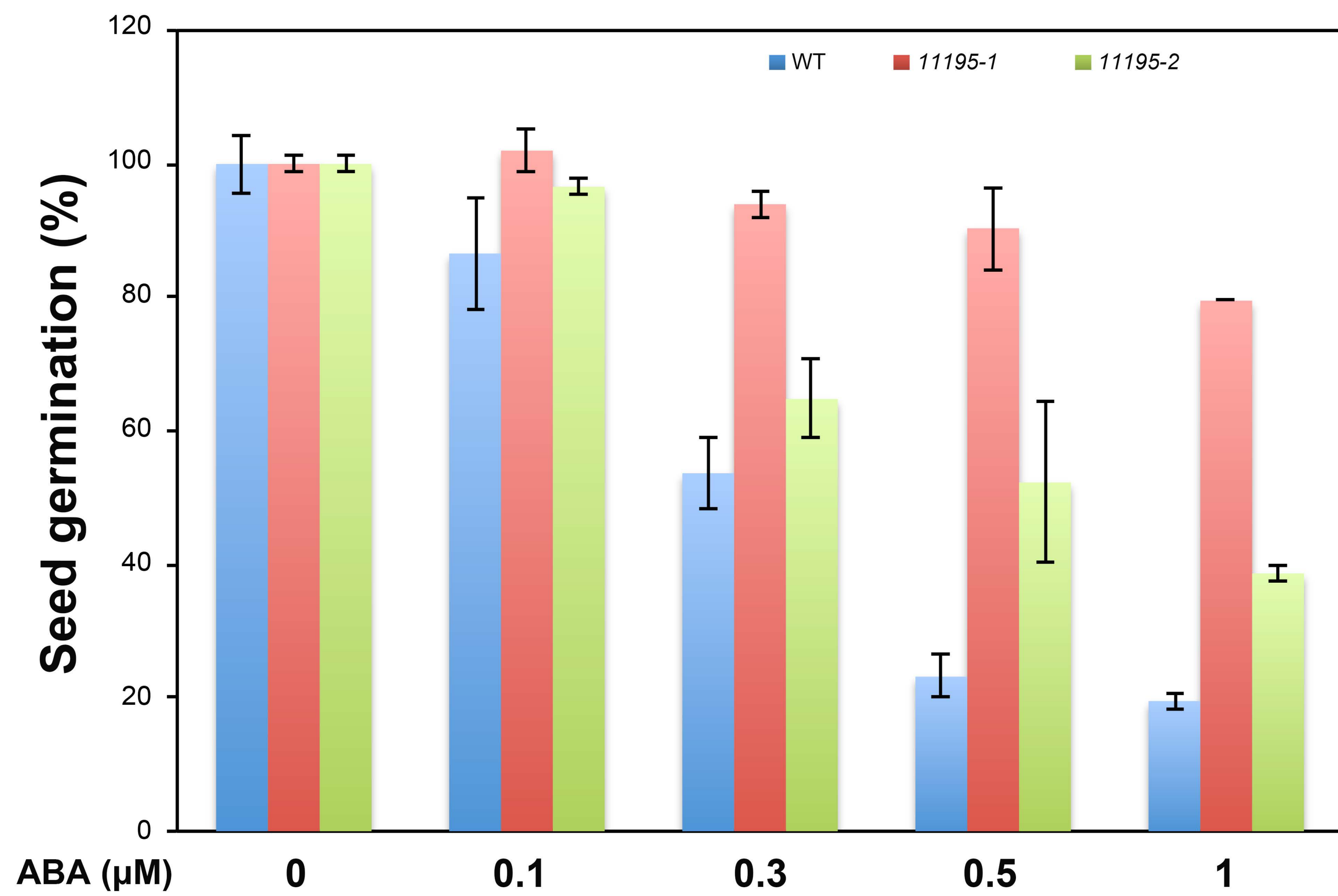
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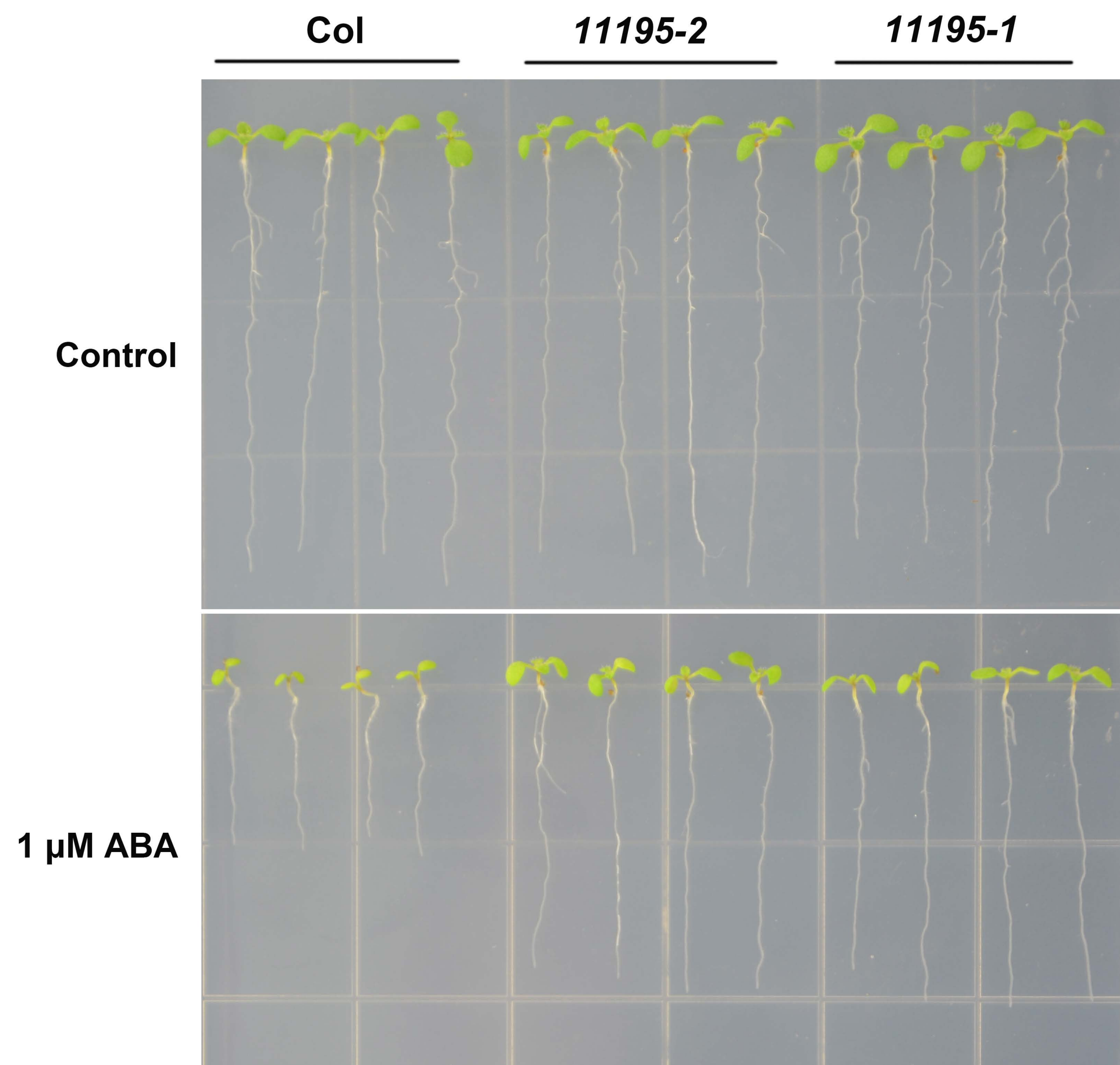
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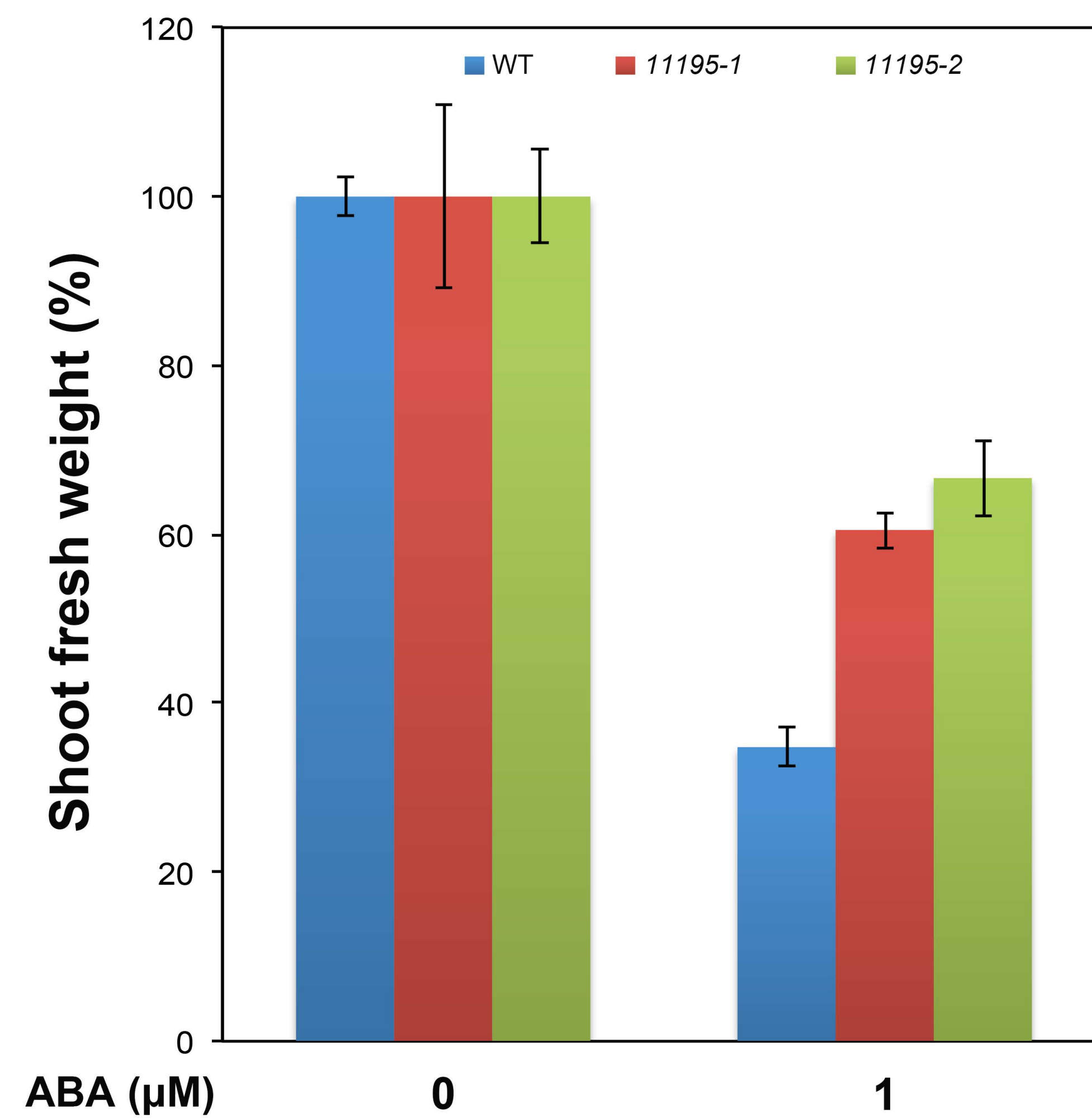
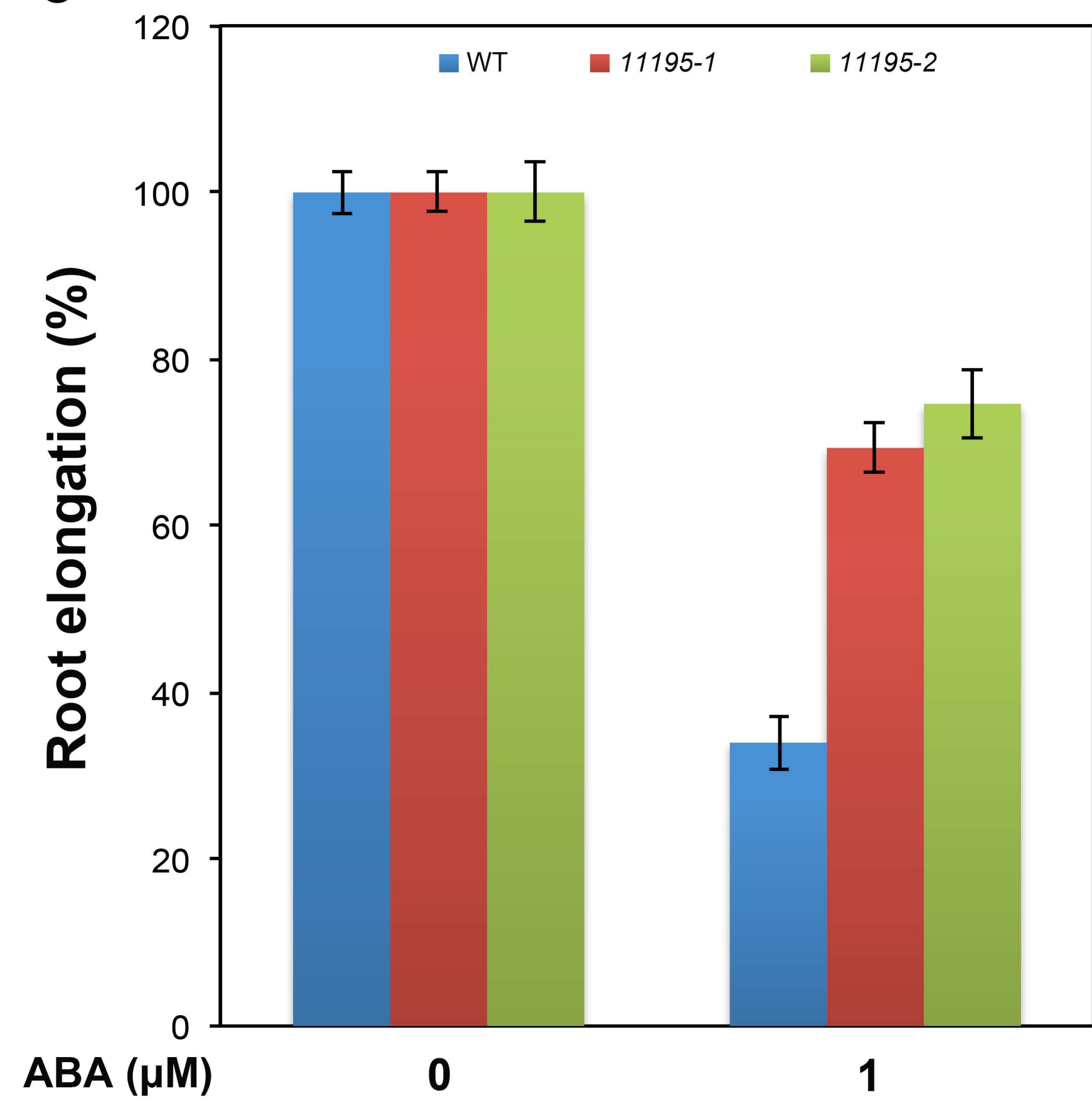
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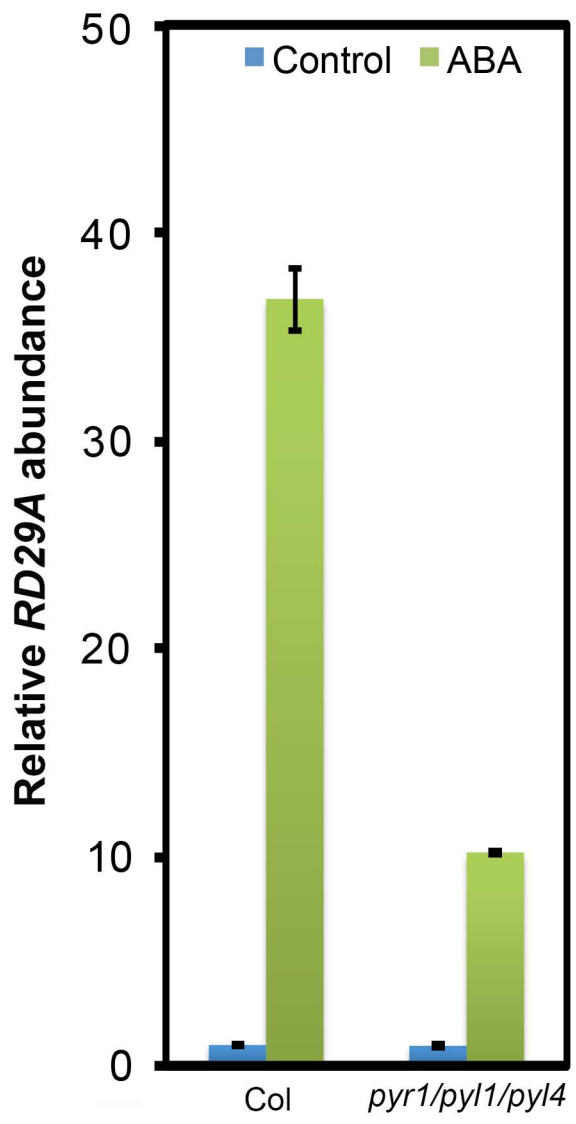
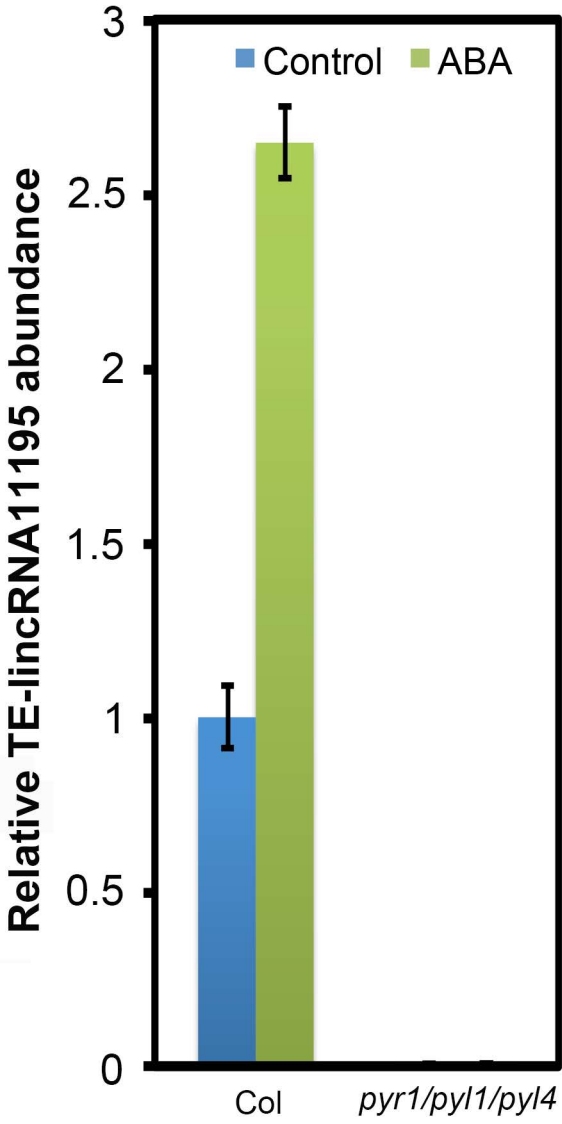


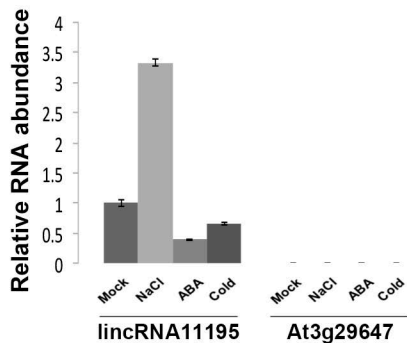
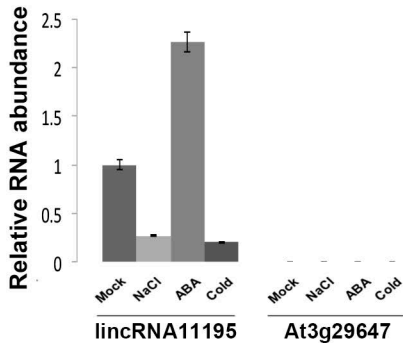
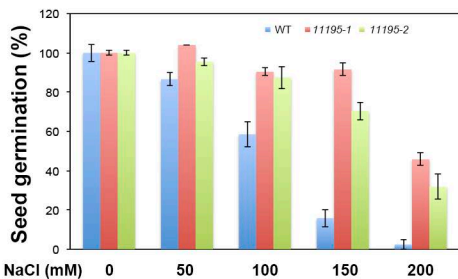
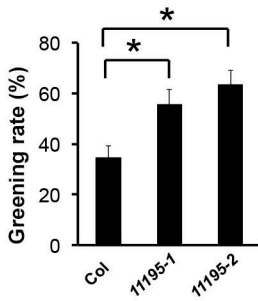
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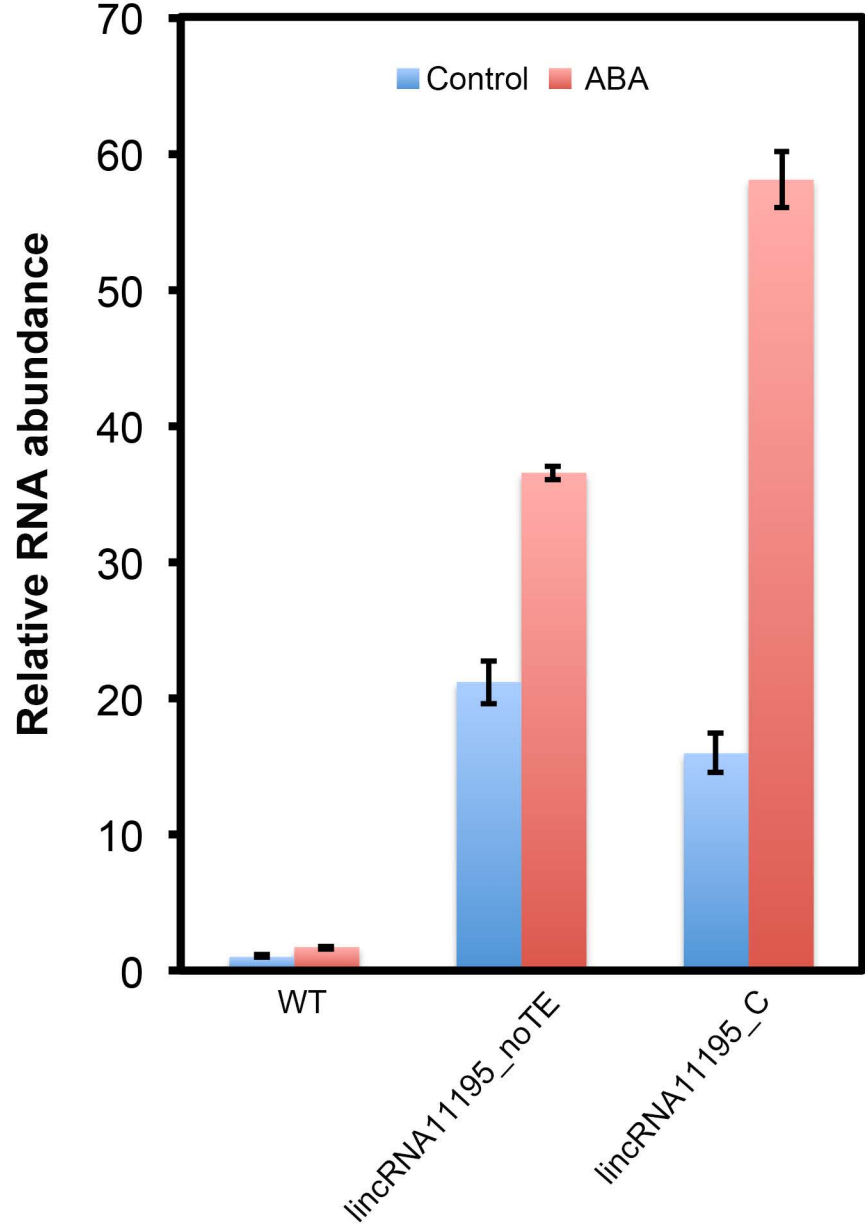
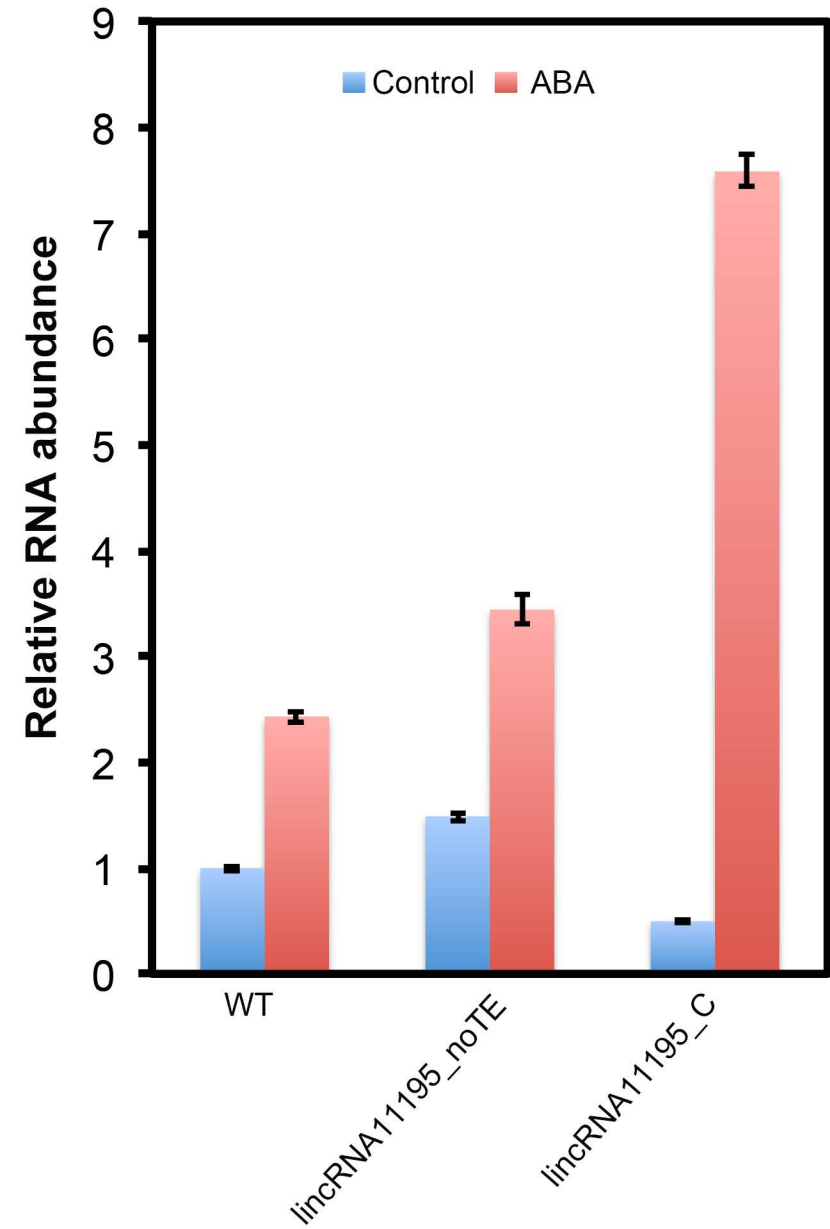


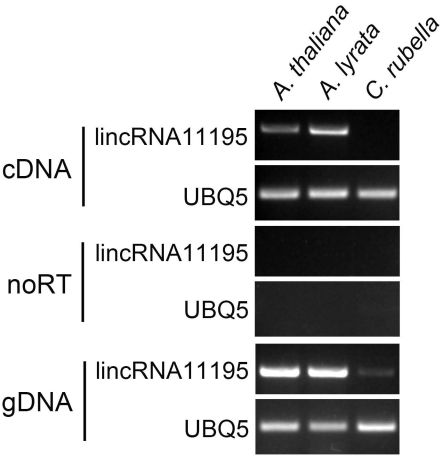
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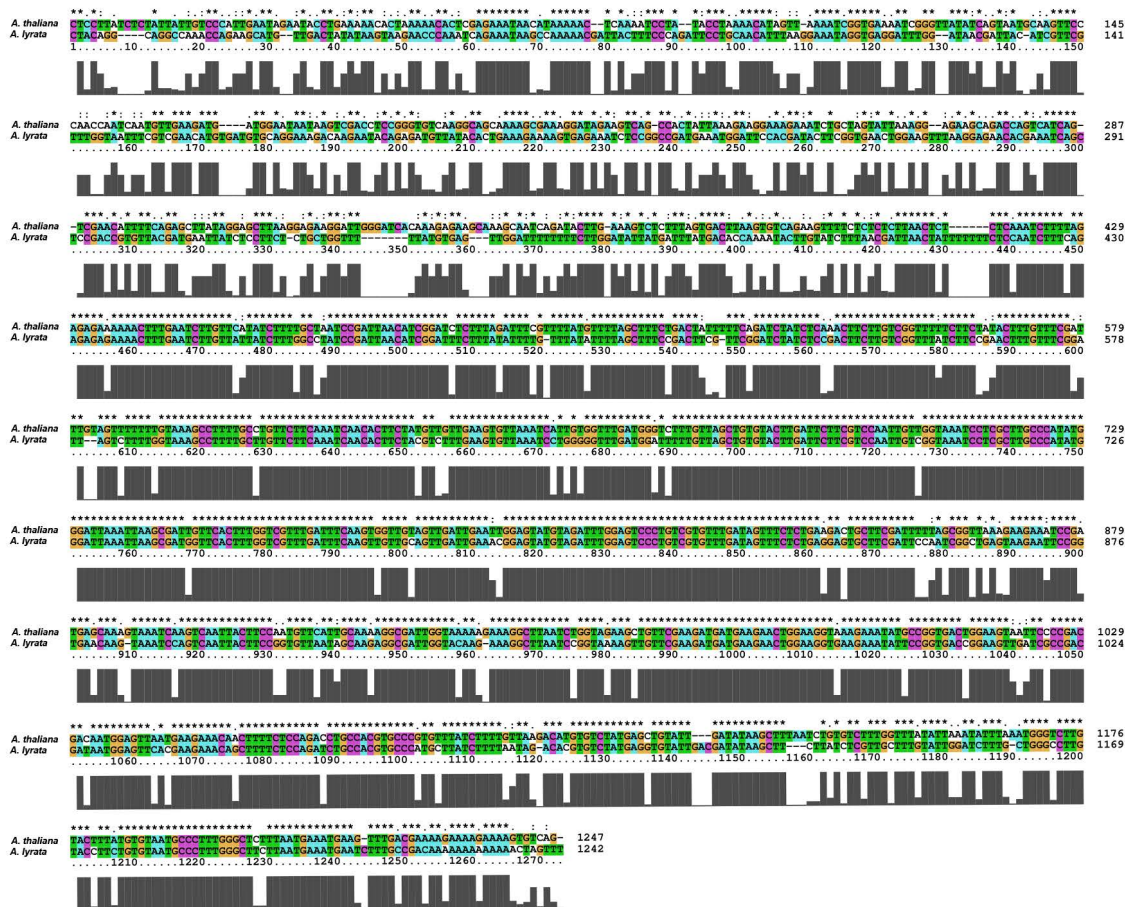


A**1 hour treatment****12 hour treatment****B****C**

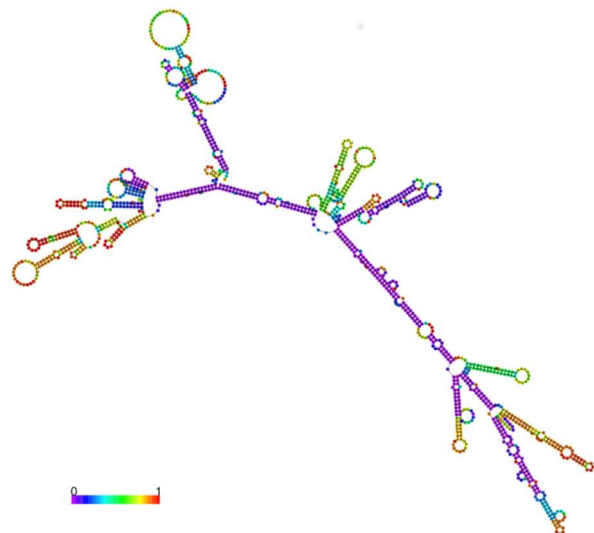




A



B



C

