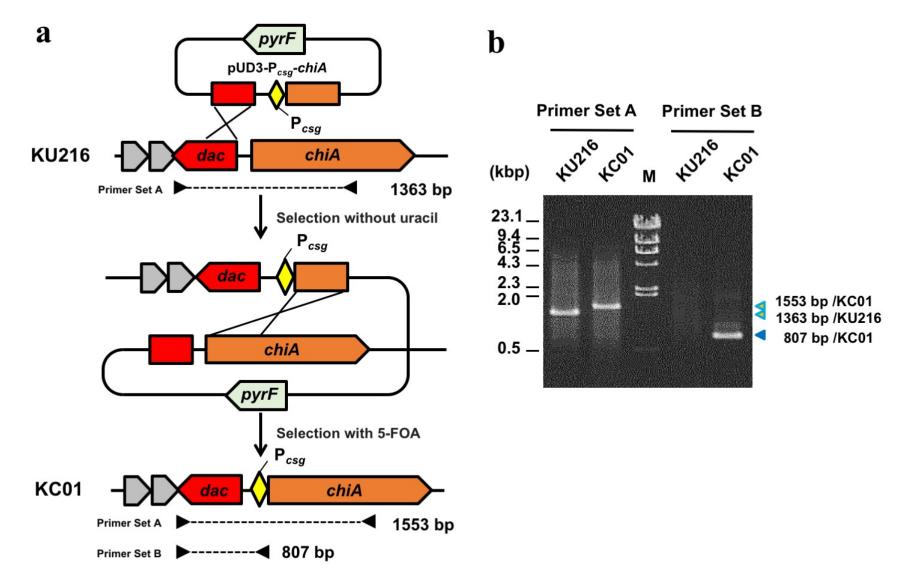
1	Supplemental Material
2	
3	Engineering of the hyperthermophilic archaeon Thermococcus
4	kodakarensis for chitin-dependent hydrogen production
5	
6 7 8 9	Mehwish Aslam <sup>1</sup> , Ayumi Horiuchi <sup>1</sup> , Jan-Robert Simons <sup>1,2</sup> , Savyasachee Jha <sup>1,2</sup> , Masahiro Yamada <sup>1</sup> , Toru Odani <sup>1</sup> , Rikako Fujimoto <sup>1</sup> , Yasuyuki Yamamoto <sup>1</sup> , Ryoma Gunji <sup>1</sup> , Tadayuki Imanaka <sup>2,3</sup> , Tamotsu Kanai <sup>1,2</sup> , Haruyuki Atomi <sup>1,2*</sup>
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Supplementary Table S1. Specific growth rates of engineered strains of *T. kodakarensis* in ASW-YT-Pyr medium

Strains	Specific growth rate (h <sup>-1</sup> )
KOD1	$0.47 \pm 0.03$
KU216	$0.45 \pm 0.04$
KC01	$0.39 \pm 0.04$
KC04	$0.40 \pm 0.05$
KU216∆t	$0.37 \pm 0.02$
KC04∆t	$0.34 \pm 0.03$
KC04∆tM1	$0.42 \pm 0.05$

The data represents the average of four independent cultures and are shown with standard deviations.



**Fig. S1 Construction of a ChiA overexpression strain of** *T. kodakarensis*. (a) A general strategy to construct strain KC01 *via* single-crossover insertion/pop-out recombination. A strong promoter ( $P_{csg}$ ) was placed just before *chiA* ORF (b) Insertion of  $P_{csg}$  was confirmed by PCR using Primer Set A (attached in *dac* or *chiA*) and Primer Set B (attached in  $P_{csg}$  promoter or *dac*). Analysis by Primer Set A shows an amplified product (1363bp) in KU216, while a longer product (of 1553bp) was amplified in in KC01 as a result of  $P_{csg}$  insertion. Analysis by Primer Set B does not give any product in the case of KU216, while a product (of 807bp) is amplified for KC01, indicating the presence of  $P_{csg}$  next to *dac*.

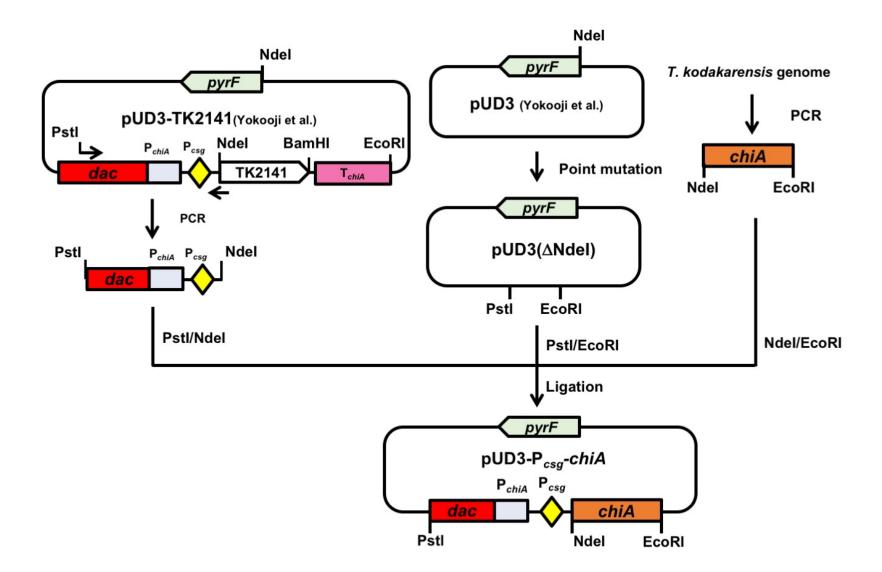


Fig. S2 Construction of the *chiA* overexpression vector pUD3-P<sub>csg</sub>-chiA.

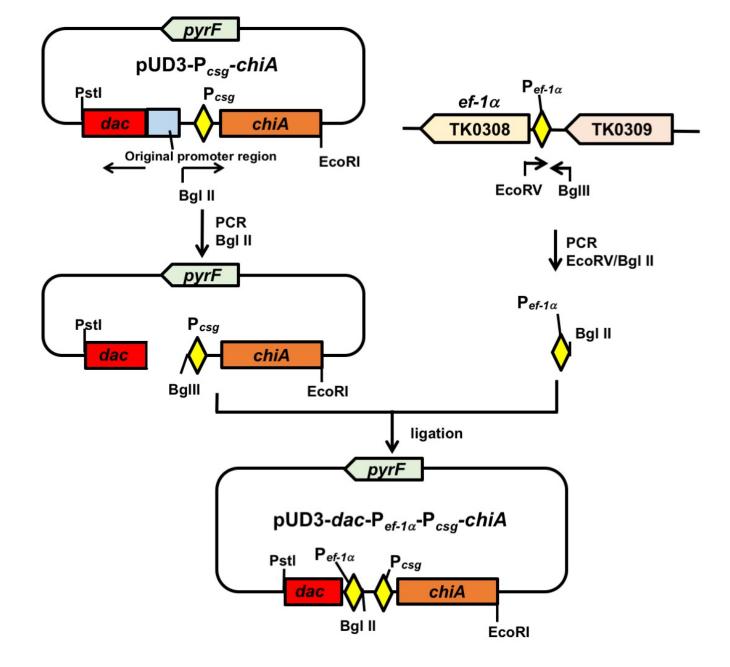


Fig. S3 Construction of the *dac* and *chiA* overexpression vector pUD3-*dac*-P<sub>ef-1a</sub>-P<sub>csg</sub>-chiA.

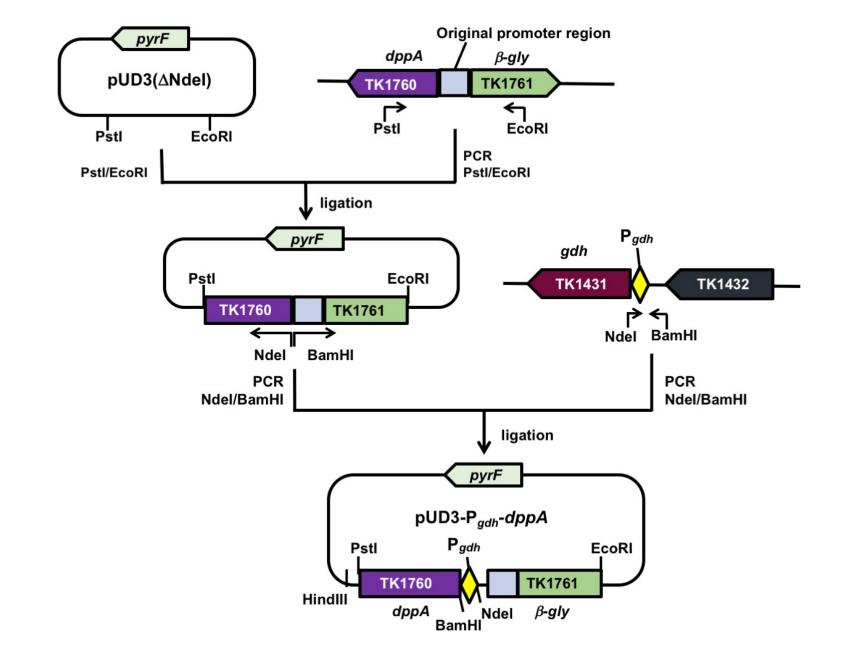
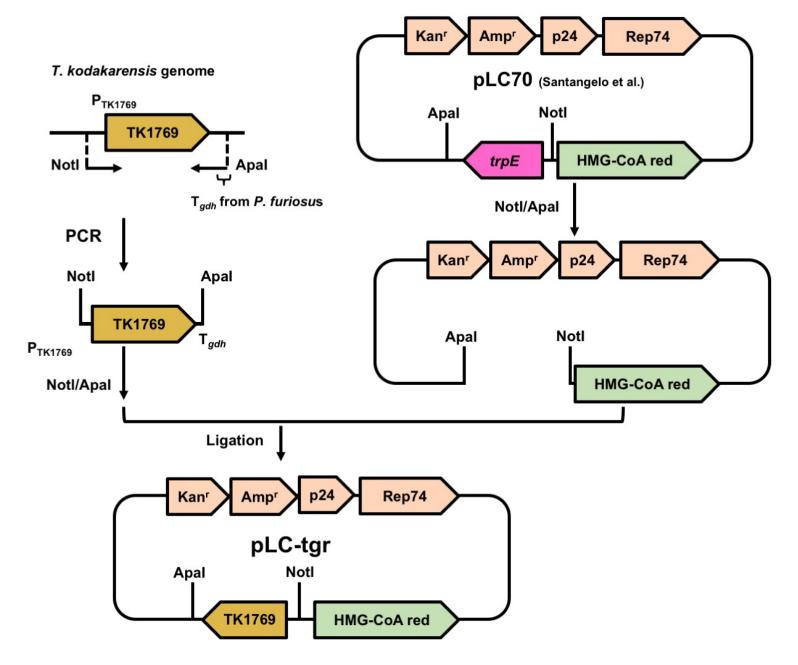


Fig. S4 Construction of the plasmid pUD3- $P_{gdh}$ -dppA for overexpression of ABC transporter genes, glmD and glmA.



**Fig. S5 Construction of the plasmid pLC-tgr for complementation of** *tgr.* The region containing *trpE* in pLC70 was replaced by TK1769 (*tgr*). HMG-CoA reductase was used as a selectable marker for transformation.

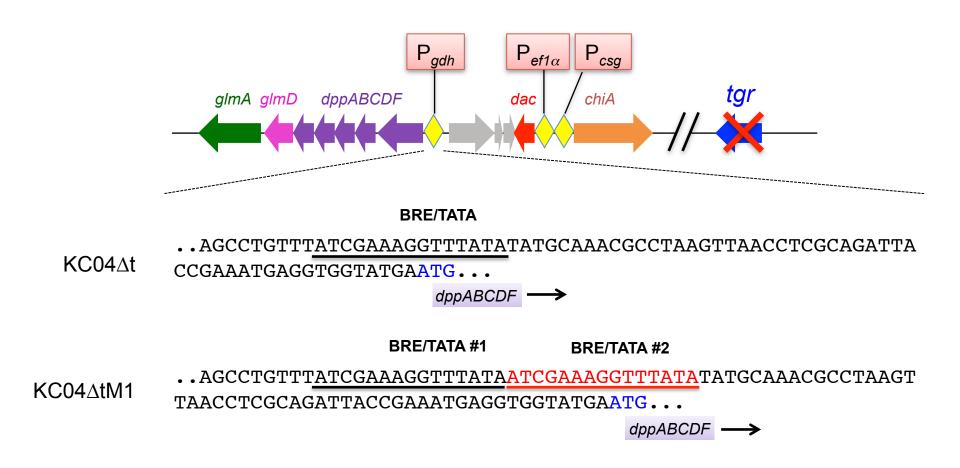
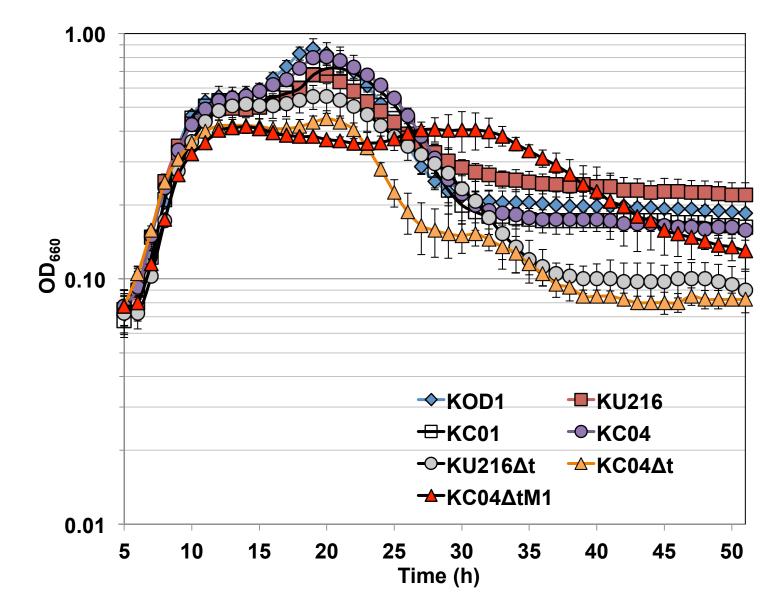


Fig. S6 Comparison in sequences in  $P_{gdh}$  regions of KC04 $\Delta$ t and KC04 $\Delta$ tM1. A 15bp repeat sequence (underline) was found in BRE/TATA region of  $P_{gdh}$  in strain KC04 $\Delta$ tM1 (shown as BRE/TATA #1 and BRE/TATA #2), which is not present in strain KC04 $\Delta$ t (the parental strain).



**Fig. S7. Growth curve of** *T. kodakarensis* **strains in ASW-YT-Pyr medium.** Batch cultivation of each strain was carried out at 85°C. Error bars represent the standard deviations of four independent growth experiments. We observed a tendency that strains with a higher number of overexpressed genes exhibited lower specific growth rates.

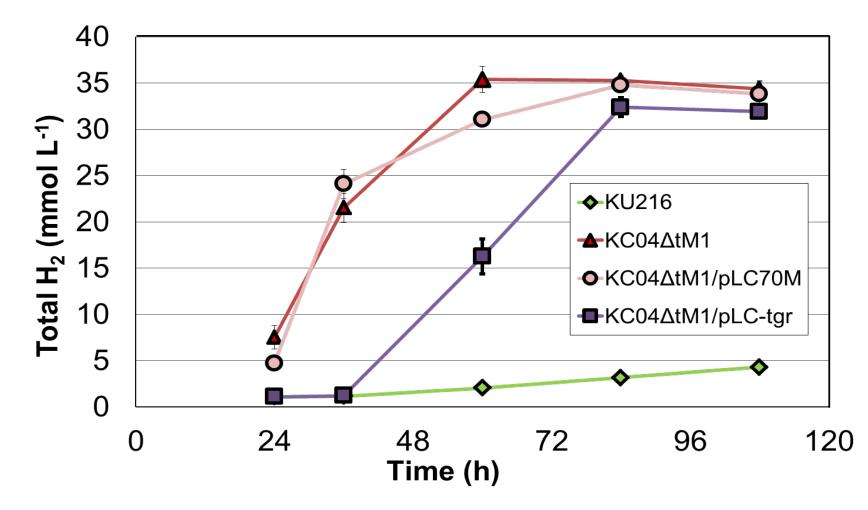


Fig. S8 Effect of *tgr* gene reintroduction on chitin-dependent hydrogen (H<sub>2</sub>) production in KC04 $\Delta$ tM1. KU216 is shown in *green* and *diamonds*, while KC04 $\Delta$ tM1 is shown in *red* and *triangles*. KC04 $\Delta$ tM1 complemented with pLC70M is shown in *pink* and *circles*, while KC04 $\Delta$ tM1 complemented with pLC-tgr is shown in *purple* and *squares*. Cultivation was performed with ASW-VMT-SC medium at 85°C. Total H<sub>2</sub> (mmol L<sup>-1</sup>) represents the amount of molecular hydrogen present in the headspace of a culture bottle per mL of culture volume (media volume was 15 mL). Error bars represent standard deviations of three independent cultivations. Reintroduction of Tgr gene to KC04 $\Delta$ tM1 results in a delay in the H<sub>2</sub> production.

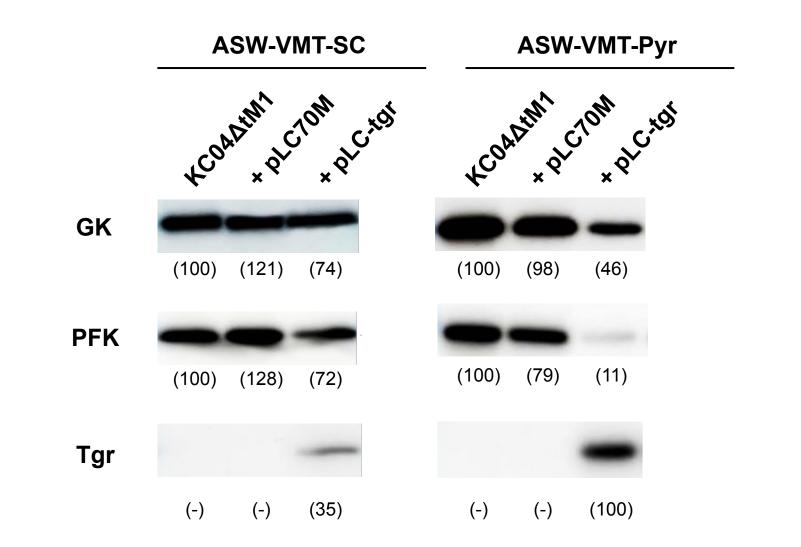


Fig. S9 Expression levels of GK, PFK and Tgr detected by Western blot analyses. Western blot analysis of GK, PFK and Tgr was performed with cell-free extracts from KC04 $\Delta$ tM1, and KC04 $\Delta$ tM1 strains complemented with pLC70M or pLC-tgr. Cells were cultivated at 85°C in ASW-VMT-SC or ASW-VMT-Pyr media. Numbers in parentheses indicate band intensities (%) relative to that of KC04 $\Delta$ tM1 cultivated in the same medium condition (which is defined as 100%). In the case of quantification of Tgr, the amount of KC04 $\Delta$ tM1 harboring pLC-tgr cultivated in ASW-VMT-Pyr medium was set as 100%. Tgr gene introduction to KC04 $\Delta$ tM1 represses expressions of GK and PFK in ASW-VMT-Pyr medium, while the effects of repression are much lower in ASW-VMT-SC medium.

**ASW-VMT-Pyr** 

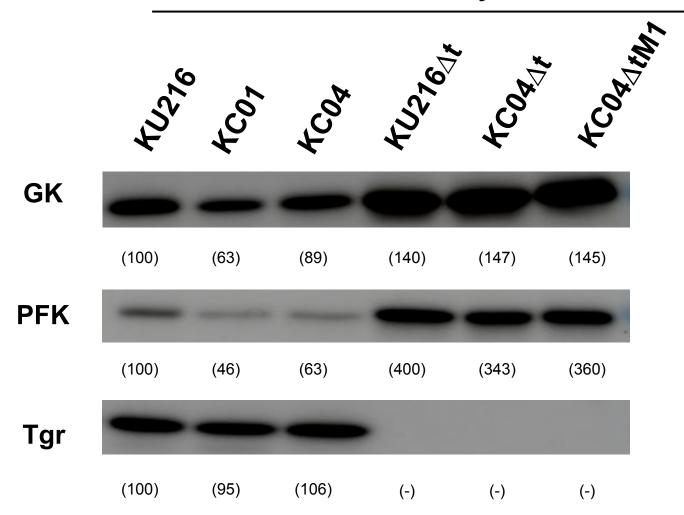


Fig. S10 Expression levels of GK, PFK and Tgr detected by Western blot analyses. Western blot analysis of GK, PFK and Tgr was performed with cell-free extracts from KU216, KC01, KC04, KU216 $\Delta$ t, KC04 $\Delta$ t and KC04 $\Delta$ tM1 strains. Cells were cultivated at 85°C in ASW-VMT-Pyr media. Equal amount (3 µg) of protein was loaded on each lane for GK and PFK, while 10 µg of protein was loaded in the case of Tgr. Numbers in parentheses indicate band intensities (%) relative to that of KU216 (which is defined as 100%). Expression levels of GK and PFK are much higher in the Tgr gene knockout strains (KU216 $\Delta$ t, KC04 $\Delta$ t and KC04 $\Delta$ tM1) compared to those of strains that harbor the Tgr gene.