# SUPPLEMENTAL ONLINE MATERIAL for:

# Estimation of biofilm-specific reaction rates with application to bacterial urea hydrolysis

#### **Authors**

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#### **Strain and growth conditions**

*Escherichia coli* MJK2 (Connolly et al., 2013) was used as the model biofilm-producing ureolytically active organism in this study. MJK2 possesses a pJN105 plasmid that has been modified to contain the urease operon from *E. coli* DH5 $\alpha$ (pURE14.8) (Collins and Falkow, 1988). The urease-carrying plasmid, pMK001, contains an L-arabinose-inducible promoter and encodes for gentamycin resistance. MJK2 also possesses a mutant chromosomal *gfp* (green fluorescent protein gene) variant that can be used for imaging. The parent strain of MJK2 is *E. coli* AF504 which is a nalidixic acid resistant derivative of *E.coli* K12 strain MG1655 (Folkesson et al., 2008). The growth medium for MJK2 was Luria–Bertani (LB) medium containing 10 g/L tryptone, 5 g/L yeast extract, 10 g/L NaCl, supplemented with 50 mM (7.5 g/L) L-arabinose, 10 μM NiCl<sub>2</sub>, 10 μg/mL gentamycin and varying amounts of urea. The medium was adjusted to a pH of 6 with HCl.

100 mL liquid cultures were inoculated from frozen stock cultures at a concentration of 1.0 μL per mL in 250 mL Erlenmeyer flasks at 37ºC on horizontal shakers running at approximately 150 rpm. The urea concentration of the starter culture matched the experimental concentration in the tube reactors. 100 μL of culture was transferred into 100 mL of the same media after approximately 24 hours. Cells from the transfer culture were harvested after approximately 12 hours by centrifugation at 4200×g for 10 minutes at 5<sup>o</sup>C in 50 mL conical centrifuge tubes containing 40 mL of culture. Cells were suspended in sterile phosphate buffered saline (PBS) by vigorous vortexing. Cells were washed one more time by centrifugation and resuspension. PBS had final concentrations of 8 g/L NaCl, 0.61 g/L KH<sub>2</sub>PO<sub>4</sub>, 0.96 g/L K<sub>2</sub>HPO<sub>4</sub> and was adjusted to a pH of 7 with HCl. The cell suspension was adjusted to an optical density of 0.6 by diluting with additional PBS after the final cell wash. Optical density was measured on triplicate 100 μL samples in polycarbonate 96-well plates (light path length of 0.26 cm) with a BioTek Instuments (Winooski, VT, USA) Synergy HT Multi-Mode Microplate Reader, and the data were analyzed using Gen5 software. The cell suspensions were used as the inoculum for the biofilm growth experiments.

#### **References relating to the strain and growth conditions:**

- Collins CM, Falkow S. 1988. Genetic Analysis of an *Escherichia coli* Urease Locus: Evidence of DNA Rearrangement. *Journal of Bacteriology* 170:1041–1045.
- Connolly J, Kaufman M, Rothman A, Gupta R, Redden G, Schuster M, Colwell F, Gerlach R. 2013. Construction of two ureolytic model organisms for the study of microbially induced calcium carbonate precipitation. *Journal of Microbiological Methods* 94:290–299.
- Folkesson A, Haagensen JAJ, Zampaloni C, Sternberg C, Molin S. 2008. Biofilm Induced Tolerance towards Antimicrobial Peptides. *Plos One* 3:e1891.

#### **Biofilm Thin Section Calculations**

The following calculations were applied to the thresholded thin section images (e.g. Figure 2C) in order to estimate a representative biofilm thickness at discrete points within the tube.

	Pixel Size (p) $0.449965 \mu m/px$
Image Size $(X$ by $Y)$ 1940 by 1460 px	
Total Image Area (A) $573471.78 \text{ }\mu\text{m}^2$	
Visible Biofilm Arc Length $(S)$ 922.95 $\mu$ m	
Tube Inside Radius $(r)$ 800 $\mu$ m	

Table SI – Constants used in thickness calculations

The visible biofilm arc length was calculated by first calculating the known chord length (C) of the visible biofilm arc,

$$
C = X p = 872.61 \,\text{µm} \,. \tag{S1}
$$

Images were taken such that they were centered in the long dimension  $(X)$  of the image. The central angle of the biofilm arc  $(\theta)$  can then be calculated with a simple trigonometric relationship where

$$
\theta = 2 \sin^{-1} \left( \frac{0.5 \text{ C}}{\text{r}} \right) = 1.154 \text{ rad}, \qquad (S2)
$$

and finally the biofilm arc length can be calculated by

$$
S = r \theta = 922.95 \,\text{µm} \,. \tag{S3}
$$

Images were thresholded as stated in the main article and the image area occupied by biofilm  $(A_f)$  was determined in ImageJ. The calculated average biofilm thickness  $(L_f)$  for each image, as reported in Tables S2-S9, was then be calculated by

$$
L_f = \frac{A_f}{S} \tag{S4}
$$

It should be noted that this calculation is only valid for thin biofilms. Thin in this context means that the biofilm thickness is much smaller than the tube radius  $(L_f \ll r)$ . If the thin condition is not met, the visible area of the biofilm will be dependent on  $L_f$  due to the vertically (rather than radially) cut off biofilms at the edges of the image.

## **Urea Measurements**

Table SII – Urea measurements obtained in the study including those that were eliminated from the analysis.



Note: Samples "Out C" and "In" were taken as the representative concentrations for each tube.

## **Biofilm Thin Section Data**

Table SIII - Tube 1 thin section data.





Table SIV - Tube 2 thin section data.



Table SV - Tube 3 thin section data.



Table SVI - Tube 4 thin section data.



Table SVII - Tube 6 thin section data.



Table SVIII - Tube 7 thin section data.



Table SIX - Tube 8 thin section data.



Table SX - Tube 9 thin section data.