Commensal Gut Microbiota Immunomodulatory Actions in Bone Marrow and Liver

have Catabolic Effects on Skeletal Homeostasis in Health

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Supplementary Figure S1: Femur length and cortical bone analysis. (**a-c**) 11 to 12 week-old male SPF & GF mice were euthanized; femur harvested for micro-CT analysis (n=4/gp). (**a**) Micro-CT analysis of femur length. (**b,c**) Micro-CT analysis of femur mid-diaphysis cortical bone: (**b**) Ct.Ar/Tt.Ar = cortical area fraction; (**c**) Ct.Th = cortical thickness. Data are expressed as mean ± SEM.



Supplementary Figure S2: Osteoclast-precursor (OCP) differentiation assays. (a–c) 11 week-old male SPF & GF mice were euthanized; bone marrow harvested; hematopoietic progenitor cells (HPCs) isolated. Magnetic cell sorting was applied to separate CD11b^{neg} HPCs, which were then stimulated in culture (primed with CSF1) to enrich for CD11b^{neg} osteoclast-precursor (OCP) cells having high osteoclastic potential. CD11b^{neg} OCP cultures were then stimulated with control (CSF1 alone) or treatment (CSF1 & RANKL) media for 3, 5 and 7 days. Cytomorphometric cellular differentiation endpoints were analyzed in TRAP stained CD11b^{neg} OCP cultures at day-3/5/7; TRAP+ cell with ≥3 nuclei considered an osteoclast. (a) Day-3 TRAP stain assay (n=4/gp). Representative images (200X) of CD11b^{neg} OCP cultures stimulated with control (CSF1) media for 3 days. (b) Day-5 TRAP stain assay (n=4/gp). Representative images (100X) of CD11b^{neg} OCP cultures stimulated with control (CSF1) media for 5 days. (c) Day-7 TRAP stain assay (n=4/gp). Representative images (100X) of CD11b^{neg} OCP cultures stimulated with control (CSF1) media for 7 days.



Supplementary Figure S3: Commensal microbiota *in vivo* regulation of pattern-recognition receptor (PRR) signaling in the liver. (a,b) 11 to 12 week-old male SPF & GF mice were euthanized; livers harvested for gene expression assays. RNA was isolated from livers (n=4/gp), and NanoString analysis was carried out to assess pattern-recognition receptor (PRR) signaling. (a) Analysis of toll-like receptor (TLR) and nucleotidebinding oligomerization domain (NOD) – like receptor levels: *Tlr1*, *Tlr2*, *Tlr3*, *Tlr4*, *Tlr5*, *Tlr6*, *Tlr7*, *Tlr8*, *Tlr9*, *Nod1*, *Nod2* RNA. (b) *Myd88*, *Tirap(Mal)*, and *Irak4* RNA assessed as critical regulators of TLR2 mediated signal transduction; *Ticam1(Trif)* RNA assessed as a critical regulator of TLR3 mediated signal transduction. Data were normalized to the geometric means of spiked-in positive controls and internal control genes. Absolute quantification of RNA expressed as normalized RNA counts. Data reported as mean \pm SEM. *p<0.05 vs. SPF; ***p<0.001 vs SPF.



Supplementary Figure S4: Bone marrow T-cell hematopoiesis. (**a**,**b**) 11 week-old gender matched (2 male and 2 female per group) SPF & GF mice were euthanized; femoral whole bone marrow cells were isolated and stained for flow cytometry analysis (n=4/gp). (**a**) % CD3⁺CD4⁺CD8⁻ (helper) T-cells. (**b**) % CD3⁺CD4⁻CD8⁺ (cytotoxic) T-cells. Percentages are expressed relative to total marrow cells. Data are reported as mean ± SEM.



Supplementary Figure S5: Bone marrow effector CD8⁺T-cell intracellular cytokine analysis. (**a**,**b**) 11 week-old gender matched (2 male and 2 female per group) SPF & GF mice were euthanized; femoral whole bone marrow was plated overnight for cytokine activation (PMA, Ionomycin, Monensin); cells were isolated and stained for flow cytometric analysis of intracellular cytokine expression (n=4/gp). (**a**) % CD3⁺CD4⁻CD8⁺IFNγ⁻ IL17a⁺ (CD8⁺IL17a⁺) cells. (**b**) % CD3⁺CD4⁻CD8⁺IFNγ⁺IL17a⁻ (CD8⁺IFNγ⁺) cells. Percentages are expressed relative to CD3⁺CD4⁻CD8⁺ cells. Data reported as mean ± SEM.



Supplementary Figure S6: Schematic summarizing study findings and proposed Gut-Liver-Bone

Axis.