Supporting Information

An ¹¹¹In-Labeled PLA-PEG Nanoparticle for Imaging PSMA-Expressing Tissues

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1. Reaction scheme to synthesize **DOTA-PEG-Alkyne** and **In-DOTA-PEG-Alkyne**

LC-MS spectra for **DOTA-PEG-alkyne** (blue) and **In-DOTA-PEG-alkyne** (red)

2. Stability studies of stable **In-DOTA-PEG-alkyne** (MW: 729.21) by LC-MS) in different experimental condition used for formulation of nanoparticles (click chemistry) formulation

3. Calculation radiolabeling yield and specific activity of 111 In-labeled nanoparticles

Four separate spin-columns were used to purify the nanoparticles to remove lowmolecularweight impurities and free ¹¹¹In-DOTA-PEG-alkyne from the nanoparticle formulation before performing the biological experiments. A table was prepared to show the calculation of radiolabeling yield and specific activity.

5. HPLC peak for stable (cold) **In-DOTA-PEG-alkyne** (λ =220 nm)

6. HPLC peak for unlabeled DOTA-PEG-alkyne (λ =220 nm). Peak at 12.8 min did not show any mass at ESI-MS. (λ =220 nm, top) and (λ =254 nm, bottom).

7. Supporting Figures for Tissue imaging

Figure S1*. Ex vivo* NIRF imaging of two mouse pairs each receiving either TNP or UNP as indicated. Mice were sacrificed and imaged 72 h after nanoparticle administration. Tumor types are as indicated. Heterogeneity of tumor uptake is apparent by lack of tumor uptake in either line in UNP mouse 2. All images are scaled to the same acquisition parameters.

Figure S2. *In vitro* epifluorescence micrograph of cultured PSMA negative PC-3 flu cells containing TNPs. This magnification shows the intracellular distribution of TNPs (red) with PSMA (green, absent), tubulin (white) and nuclei (blue). White arrows indicate some co-localization of TNPs with tubulin (white at the centrosome), possibly due to passive endocytosis. Scale bar = 10μ m.

Figure S3. *In vitro* epifluorescence micrograph of cultured PSMA+ PC-3 PIP cells containing UNPs. This magnification shows the intracellular distribution of TNPs (red) with PSMA (green), tubulin (white) and nuclei (blue). White arrows indicate some co-localization of TNPs with tubulin (white at the centrosome), possibly due to passive endocytosis. Scale bar = 10 µm.

Figure S4.

Figure S4. *In vitro* epifluorescence micrograph of cultured PSMA negative PC3 flu cells containing UNPs. This magnification shows the intracellular distribution of UNPs (red) with PSMA (green, absent), tubulin (white) and nuclei (blue). White arrows indicate some colocalization of TNPs with tubulin (white at the centrosome), possibly due to passive endocytosis. Scale bar = 10μ m.

Figure S5. Flow cytometry analysis of PSMA surface expression in PSMA(+) PC-3 PIP & PSMA(-) PC-3flu cell lines. The dot plot (left panel) and the histogram (right panel) show that PC-3 PIP has a uniformly high level of PSMA expression where as PC-3flu is negative.

Supplementary Figure 6. *In vivo* NIRF imaging of TNP-IRDye and UNP-IRDye distribution in tumor bearing mice at 72 h post-injection. Four athymic nude mice bearing PC-3 PIP (red arrows) and PC-3 flu (yellow arrows) xenografts were injected as indicated with either fluorescent TNP or UNP. Images at 72 h show TNP accumulation around both PIP and flu xenografts as well as around knee joints. UNP accumulation had largely cleared from both tumors except from a \sim 4 mm diameter PIP tumor within the UNP 1 mouse. Color bar indicates relative fluorescence units. PC-3 PIP tumor (red arrow), PC-3 flu tumor (yellow arrow)