

SUPPORTING INFORMATION

Hole Hopping through Tryptophan in Cytochrome P450

Maraia E. Ener,[†] Harry B. Gray,* and Jay R. Winkler*

Beckman Institute, California Institute of Technology, Pasadena, California 91125, United States

[†]Current address:

Department of Chemistry
Yale University
225 Prospect St.
P.O. Box 208107
New Haven CT 06520

Corresponding Authors

*Harry B. Gray. *Jay R. Winkler

Additional Experimental Details

Laser transient spectroscopy. Excitation pulse energies were in the range of 5-10 mJ/pulse. Samples were held in cuvettes with 1- cm optical path lengths and the absorbance at the 480-nm excitation wavelength was approximately 0.15.

The transient kinetics shown in manuscript Figures 3 and 4 are normalized to the prompt bleach at 440 nm. This bleach signal corresponds to formation of the $^*Ru^{2+}$ excited state. By normalizing to the initial excited state concentration, the signals for the different proteins reflect the relative yields of subsequent intermediates that form.

Sample spectra. Absorption spectra of $[Ru(bpy)_2(lAphen)]^{2+}$, CYP102A1, $Ru_{C97}(CYP102A1)W96$, and $pMeODMA^{*+}$ have been reported previously.^{1,2}

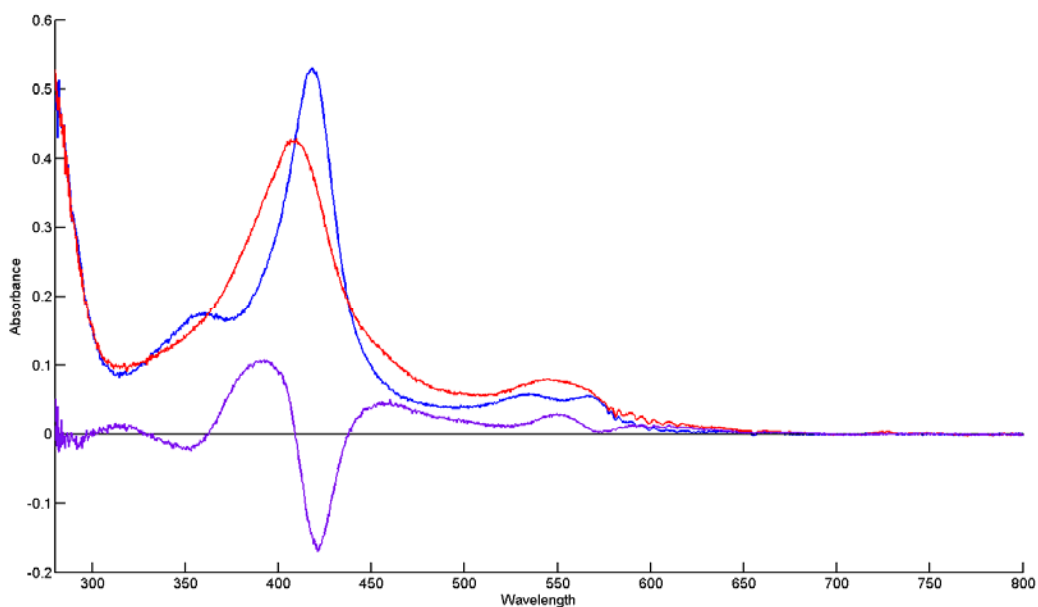


Figure S1. Spectra of ferric (blue) and ferrous (red) wild-type P450-BM3, and the ferrous - ferric difference spectrum (purple).

Electron Transfer Rate Calculations

Electron-transfer rate constants were estimated using the following semiclassical expression:³

$$k_{\text{ET}} = (10^{13} \text{ s}^{-1}) \times e^{-\beta(r-r_0)} \times e^{-\frac{(\Delta G^\circ + \lambda)^2}{4\lambda k_B T}}$$

The following parameters were used for all estimated rate constants in Table S1: $\beta = 1.1 \text{ \AA}^{-1}$; $r_0 = 3 \text{ \AA}$; $\lambda = 0.8 \text{ eV}$; $k_B = 8.6165 \times 10^{-5} \text{ eV K}^{-1}$; $T = 295 \text{ K}$.

Table S1. Estimated electron transfer rate constants

Reaction	ΔG° , eV	r , \AA	k_{ET} , s^{-1}	$\tau_{\text{ET}} = k_{\text{ET}}^{-1}$
$\text{Ru}_{\text{C97}}^{3+}\text{-Fe}^{3+}\text{P} \rightarrow \text{Ru}_{\text{C97}}^{2+}\text{-Fe}^{3+}\text{P}^{*\bullet}$	-0.2	20.8	3.8×10^2	2.7 ms
$\text{Ru}_{\text{C97}}^{3+}\text{-W96} \rightarrow \text{Ru}_{\text{C97}}^{2+}\text{-W96}^{*\bullet}$	-0.1	11.88	1.4×10^6	0.7 μs
$\text{W96}^{*\bullet}\text{-Fe}^{3+}\text{P} \rightarrow \text{W96}\text{-Fe}^{3+}\text{P}^{*\bullet}$	-0.1	7.15	2.5×10^8	4 ns
$\text{Ru}_{\text{C97}}^{3+}\text{-Fe}^{3+}\text{P} \rightarrow \text{Ru}_{\text{C97}}^{2+}\text{-Fe}^{2+}\text{P}$	-0.9	19.5	1.2×10^5	8.6 μs
$\text{Ru}_{\text{C97}}^{3+}\text{-Fe}^{3+}\text{P} \rightarrow \text{Ru}_{\text{C97}}^{2+}\text{-Fe}^{2+}\text{P}$	-0.9	23.8	1.0×10^3	1 ms

References

- [1] Ener, M. E., Lee, Y. T., Winkler, J. R., Gray, H. B., and Cheruzel, L. (2010) Photooxidation of cytochrome P450-BM3, *Proc. Natl. Acad. Sci. USA* 107, 18783-18786.
- [2] Sassoon, R. E., Gershuni, S., and Rabani, J. (1992) Photochemical generation and consequent stabilization of electron-transfer products on separate like-charged polyelectrolytes, *J. Phys. Chem.* 96, 4692-4698.
- [3] Winkler, J. R., and Gray, H. B. (2014) Electron Flow through Metalloproteins, *Chem. Rev.* 114, 3369-3380.