

Supplemental Information, Figure S1. Conserved *miR-132* is necessary for brain vascular integrity. (A) Conservation of *miR-132* from *Drosophila* to Human. The red box shows the seed region of *miR-132*. (B) Whole-mount *in situ* hybridization showing that *miR-132* is highly expressed in the brain region of zebrafish embryos at 3 days post-fertilization (dpf). (C) Schematic showing the design of *miR-132* MO and *miR-132* loop MO. Target-1, *miR-132* MO targeting site; target-2, *miR-132* loop MO targeting site. The sequence of mature *miR-132* is in red. (D) Efficiency of MOs in downregulating *miR-132* expression in zebrafish embryos. The experiments were repeated 12 times. (E) Death rate of 1-dpf embryos injected with Ctrl MO or *miR-132* MO. The experiments were repeated 7 times. (F) Malformation rate of 3-dpf embryos injected with Ctrl MO and *miR-132* MO. The experiments were repeated 7 times. (G) Representative TEM images of a control embryo and a hemorrhagic *miR-132* morphant showing the accumulation of blood cells (red arrows) in the brain ventricle in *miR-132* morphant. (H) Representative projected confocal images showing 10-kDa dextran leakage from brain blood vessels in a 3-dpf Tg(Flk1:eGFP) *miR-132* morphant. The regions outlined by the white rectangles were enlarged. (I and J) Representative projected confocal time-lapse images showing that red fluorescent bead (100 nm in diameter) leaked from brain blood vessels in a 3-dpf Tg(Flk1:eGFP) control zebrafish (I) and *miR-132* morphant (J). In (J), beads leaked from two functional vessels (white circles) which already carried blood flow. Scale bar, 10 μm (G), 100 μm and 25 μm (insert) (H), 100 μm (left) and 50 μm (right) (I and J). Error bars, SEM. n.s., no significant; * $P < 0.05$, ** $P < 0.01$ (one-way ANOVA with post-hoc Tukey's multiple comparison test for (D); unpaired two-tailed Student's *t* test for (E) and (F)).