## Supplementary Information

Figures S1 to S7 and Tables S1 and S2

Evaluation of *ATM* heterozygous mutations underlying individual differences in radiosensitivity using genome editing in human cultured cells

Ekaterina Royba<sup>1</sup>, Tatsuo Miyamoto<sup>1</sup>, Silvia Natsuko Akutsu<sup>1</sup>, Kosuke Hosoba<sup>1</sup>, Hiroshi Tauchi<sup>2</sup>, Yoshiki Kudo<sup>3</sup>, Satoshi Tashiro<sup>4</sup>, Takashi Yamamoto<sup>5</sup> and Shinya Matsuura<sup>1\*</sup>

 <sup>1</sup>Department of Genetics and Cell Biology, Research Institute for Radiation Biology and Medicine, Hiroshima University, Hiroshima 734-8553, Japan
<sup>2</sup>Department of Biological Sciences, Faculty of Science, Ibaraki University, Mito 310-8512, Japan
<sup>3</sup>Department of Obstetrics and Gynecology, Graduate School of Biomedical Sciences, Hiroshima University, Hiroshima 734-8551, Japan
<sup>4</sup>Department of Cellular Biology, Research Institute for Radiation Biology and Medicine, Hiroshima University, Hiroshima 734-8553, Japan
<sup>5</sup>Department of Mathematical and Life Sciences, Graduate School of Science, Hiroshima University, Higashi-Hiroshima 739-8526, Japan

\*Correspondence: e-mail: shinya@hiroshima-u.ac.jp



**Figure S1. Three independent CBMN assays in primary fibroblasts from an A-T-affected family. (a), (c) and (e)** Three graphs showing the results of three independent CBMN assays. Percentage of IR-induced MN formation in fibroblasts from all members of the A-T-affected family (>1000 BN cells, >50 BN cells only in A-T patient fibroblasts). **(b) (d) and (f)** The graphs from (a), (c) and (e), respectively, with magnification of the Y-axis including the percentage of IR-induced MN formation in fibroblasts derived from A-T heterozygous carriers and unaffected individuals.



## Figure S2. CRISPR-ObLiGaRe method generated ATM-edited hTERT-RPE1 cell lines with a uniform genetic background.

(a) The CRISPR/ObLiGaRe-mediated genome-editing strategy. *ATM* gene exon 11 and the targeting vector containing an *hsvTK-2A-Neo<sup>r</sup>* selection cassette were cleaved in hTERT-RPE1 cells using a set of Cas9-nuclease and the sgRNA targeting for *ATM* gene exon 11. The linearized targeting vector was integrated into *ATM* gene exon 11 in an NHEJdependent manner. The orientation of the targeting vector upon insertion could not be controlled. For genotyping of the targeted site in positive drug-selected clones, PCR genotyping and direct sequencing were combined. (b) PCR genotyping step: the integration of a drug-resistant cassette was evaluated by PCR using the primer pairs A and B as shown in (a). *ATM* exon 11 without a gene cassette was amplified by PCR using primer pair C denoted in (a). An agaroseelectrophoresis image showed the presence or absence of the insertion of a targeting vector into *ATM* gene exon 11 in the parental RPE1 cell, *ATM<sup>-/-</sup>* clone 2 and *ATM<sup>+/-</sup>* clone 2. (c) Direct sequencing step: Insertions/deletions in an allele without a drug-resistant cassette were directly checked by Sanger sequencing. *ATM<sup>-/-</sup>* clone 2 contained a 1-bp insertion (c.1653 insT, p.V551X) at a CRISPR/Cas9-mediated DSB site. (d) Schematic representation of the final genotype in *ATM*-edited hTERT-RPE1 cell clones generated by the CRISPR/ObLiGaRe method. For details, see Table 1.



Figure S3. Three independent CBMN assays in ATM-edited hTERT-RPE1 cell clones.

(a), (c) and (e) Three graphs showing the results of three independent CBMN assays. Percentage of IR-induced MN formation in *ATM*-edited cell clones (>1000 BN cells, >50 BN cells only in  $ATM^{-/-}$  cell clones). (b), (d) and (f) The graphs from (a), (c) and (e), respectively, with magnification of the Y-axis including the percentage of IR-induced MN formation in  $ATM^{+/-}$  and  $ATM^{+/+}$  cell clones.



## Chromosomal aberrations produced after irradiation in G1/ early S phase

**Figure S4. Evaluation of unrepaired DNA DSBs from chromosomal aberrations detected by PNA-FISH analysis.** Since the pattern of chromosomal aberrations is dependent on the number of unrepaired DSBs, we considered the process of formation of each such aberration to quantify unrepaired DSBs. Scale bars: 2 µm.



Figure S5. Centromere/telomere PNA-FISH analysis in the primary fibroblasts from the A-T-affected family. (a), (c) and (e) Raw data for three independent trials in the primary fibroblasts. (b), (d) and (f) from (a), (c) and (e), respectively, with magnification of the Y-axis for three independent trials in the fibroblasts from A-T heterozygous carrier and normal individuals.



Figure S6. Centromere/telomere PNA-FISH data in *ATM*-edited hTERT-RPE1 cell lines. (a), (c) and (e) Raw data for three independent trials in *ATM*-edited hTERT-RPE1 cell lines. (b), (d) and (f) Raw data from (a), (c) and (e), respectively, with magnification of the Y-axis for three independent trials in  $ATM^{+/-}$  and  $ATM^{+/+}$  cell clones.



Figure S7. Full-scan of western blots and agarose gel electrophoresis. (a) Raw data of Fig. 1a and Fig. 2a captured using an automated capillary-based western blotting system. (b) Raw image of agarose gel electrophoresis in Fig. S2b.

Fibroblast cell line ID (Coriell Institute)	Donor —	ATM gene mutation (Transcript# ATM-201, ENST00000278616.8)			Genter
		ATM exon#	DNA sequence change	Amino acid change	Genotype
GM03487	Male, 8 y.o., proband	Exon 9 Exon 56	c.1141ins4 c.8266A>T	[\$381X] [K2756X]	ATM <sup>≁</sup>
GM03488	Male, 41 y.o., father	Exon 9	c.1141ins4	[S381X]	<i>ATM</i> <sup>+/-</sup>
GM03489	Female, 37 y.o., mother	Exon 56	c.8266A>T	[K2756X]	<i>ATM</i> <sup>+/-</sup>
GM03490	Female, 16 y.o., sister	Exon 9	c.1141ins4	[S381X]	<i>ATM</i> <sup>+/-</sup>
GM03491	Female, 15 y.o., sister	-	Wild type	Wild type	<i>ATM</i> <sup>+/+</sup>
GM03492	Male, 7 y.o., brother	-	Wild type	Wild type	<i>ATM</i> <sup>+/+</sup>

Table S1. Genetic information about ATM gene in six members of the A-T-affected family examined in this study.

C. II. K. J. D.	ATM exon#	Allele 1 (Obligare-mediated Neo <sup>R</sup> cassette integration)	Allele 2 (CRISPR/Cas9-mediated indel mutations) (Transcript # ATM-201, ENST00000278616.8)		
Cell line ID			DNA sequence change	Amino acid change	Genotype
$ATM^{\checkmark}$ clone 1	Exon 11	Neo+(reverse orientation)	c.1653insT	[V551X]	ATM <sup>≁</sup>
<i>ATM</i> <sup>-/−</sup> clone 2	Exon 11	Neo+(forward orientation)	c.1653insT	[V551X]	ATM <sup>≁-</sup>
$ATM^{+/-}$ clone 1	Exon 11	Neo+(reverse orientation)	Wild type	Wild type	ATM <sup>+/-</sup>
ATM <sup>+/-</sup> clone 2	Exon 11	Neo+(reverse orientation)	Wild type	Wild type	ATM <sup>+/-</sup>
ATM <sup>+/-</sup> clone 3	Exon 11	Neo+(reverse orientation)	Wild type	Wild type	ATM <sup>+/-</sup>
ATM <sup>+/+</sup> clone 1	-	Neo- (wild type) plus Neo+(random integration)	Wild type	Wild type	<i>ATM</i> <sup>+/+</sup>
ATM <sup>+/+</sup> parental	-	Neo- (wild type)	Wild type	Wild type	<i>ATM</i> <sup>+/+</sup>

Table S2. Genetic information of ATM-edited hTERT-RPE1 cell clones.