Title: Breast Tissue Ablation with Irreversible Electroporation in Rabbits: a safety and feasibility Study

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	ITEM	RECOMMENDATION
Title	$1 (Y \sqrt{N})$	Provide as accurate and concise a description
		of the content of the article as possible.
Abstract	2 (Y √/ N)	Provide an accurate summary of the
		background, research objectives, including
		details of the species or strain of animal
		used, key methods, principal findings and
		conclusions of the study.
NITRODUCTION		conclusions of the study.
INTRODUCTION	2 (77 //27)	
Background	$3 (Y \sqrt{N})$	a. Include sufficient scientific background
		(including relevant references to previous
		work) to understand the motivation and
		context for the study, and explain the
		experimental approach and rationale.
		b. Explain how and why the animal species
		and model being used can address the
		scientific objectives and, where appropriate,
		the study's relevance to human biology.
Objectives	4 (Y √/ N)	Clearly describe the primary and any
objectives		secondary objectives of the study, or specific
		hypotheses being tested.
METHODO		hypotheses being tested.
METHODS	7 (N. //NI)	
Ethical statement	$5 (Y \sqrt{N})$	Indicate the nature of the ethical review
		permissions, relevant licences (e.g. Animal
		[Scientific Procedures] Act 1986), and
		national or institutional guidelines for the
		care and use of animals, that cover the
		research.
Study design	6 (Y √/ N)	For each experiment, give brief details of the
		study design including:
		a. The number of experimental and control
		groups.
		b. Any steps taken to minimise the effects of
		subjective bias when allocating animals to
		treatment (e.g. randomisation procedure) and
		1 7
		when assessing results (e.g. if done, describe
		who was blinded and when).
		c. The experimental unit (e.g. a single

		animal, group or cage of animals).
		,
		A time-line diagram or flow chart can be
		useful to illustrate how complex study
		designs were carried out.
Experimental	$7 (Y \sqrt{N})$	For each experiment and each experimental
procedures		group, including controls, provide precise
		details of all procedures carried out.
		For example:
		a. How (e.g. drug formulation and dose, site
		and route of administration,
		anaesthesia and analgesia used [including
		monitoring], surgical procedure, method of
		euthanasia). Provide details of any specialist
		equipment used, including supplier(s).
		b. When (e.g. time of day).
		c. Where (e.g. home cage, laboratory, water
		maze).
		d. Why (e.g. rationale for choice of specific
		anaesthetic, route of administration, drug
		dose used).
Experimental	8 (Y √/ N)	a. Provide details of the animals used,
animals	0 (1 1/11)	including species, strain, sex,
ammais		developmental stage (e.g. mean or median
		age plus age range) and weight (e.g. mean or
		median weight plus weight range).
		b. Provide further relevant information such
		as the source of animals,
		international strain nomenclature, genetic
		modification status (e.g. knock-out or
		transgenic), genotype, health/immune status,
		drug or test naïve, previous procedures, etc.
Housing and	$9 (Y \sqrt{N})$	Provide details of:
husbandry		a. Housing (type of facility e.g. specific
		pathogen free [SPF]; type of cage or
		housing; bedding material; number of cage
		companions; tank shape and material etc. for
		fish).
		b. Husbandry conditions (e.g. breeding
		programme, light/dark cycle, temperature,
		quality of water etc for fish, type of food,
		access to food and water, environmental
		enrichment).
		c. Welfare-related assessments and
		interventions that were carried out prior to,
		merventions that were carried out prior to,

		during, or after the experiment.
Sample size	10 (Y √/ N)	a. Specify the total number of animals used
Sampre Size		in each experiment, and the number of
		animals in each experimental group.
		b. Explain how the number of animals was
		arrived at. Provide details of any sample size
		calculation used.
		c. Indicate the number of independent replications of each experiment, if relevant.
Allocating animals	$11 (Y \sqrt{N})$	a. Give full details of how animals were
to experimental		allocated to experimental groups, including
groups		randomisation or matching if done.
		b. Describe the order in which the animals in
		the different experimental groups were
		treated and assessed.
Experimental	12 (Y √/ N)	Clearly define the primary and secondary
outcomes		experimental outcomes assessed (e.g. cell
		death, molecular markers, behavioural
		changes).
Statistical methods	13 (Y √/ N)	a. Provide details of the statistical methods
Statistical filethous	13 (1 1/11)	used for each analysis.
		b. Specify the unit of analysis for each
		dataset (e.g. single animal, group of animals,
		single neuron).
		c. Describe any methods used to assess
		whether the data met the assumptions of the
		statistical approach.
RESULTS		
Baseline data	14 (Y √/ N)	For each experimental group, report relevant
		characteristics and health status of animals
		(e.g. weight, microbiological status, and drug
		or test naïve) prior to treatment or testing
		(this information can often be tabulated).
Numbers analysed	15 (Y √/ N)	a. Report the number of animals in each
		group included in each analysis. Report
		absolute numbers (e.g. 10/20, not 50%).
		b. If any animals or data were not included in
		the analysis, explain why.
Outcomes and	16 (Y √/ N)	Report the results for each analysis carried
estimation		out, with a measure of precision (e.g.
· 		standard error or confidence interval).
Adverse events	17 (Y √/ N)	a. Give details of all important adverse
1 Idvoise events	1/(1 1/11)	events in each experimental group.
		b. Describe any modifications to the

		experimental protocols made to reduce adverse events.
DISCUSSION		
Interpretation/ scientific implications	18 (Y √/ N)	a. Interpret the results, taking into account the study objectives and hypotheses, current theory and other relevant studies in the literature. b. Comment on the study limitations including any potential sources of bias, any limitations of the animal model, and the imprecision associated with the results. c. Describe any implications of your experimental methods or findings for the replacement, refinement or reduction (the 3Rs) of the use of animals in research.
Generalisability/ translation	19 (Y √/ N)	Comment on whether, and how, the findings of this study are likely to translate to other species or systems, including any relevance to human biology.
Funding	20 (Y √/ N)	List all funding sources (including grant number) and the role of the funder(s) in the study.

Signature: <u>Liqi Bi</u>

May 25, 2017