

**Interleukin 37 promotes angiogenesis through TGF- $\beta$  signaling****One-sentence summaries:** IL-37 promotes angiogenesis via TGF- $\beta$  signaling

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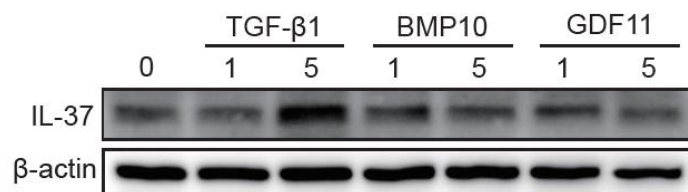
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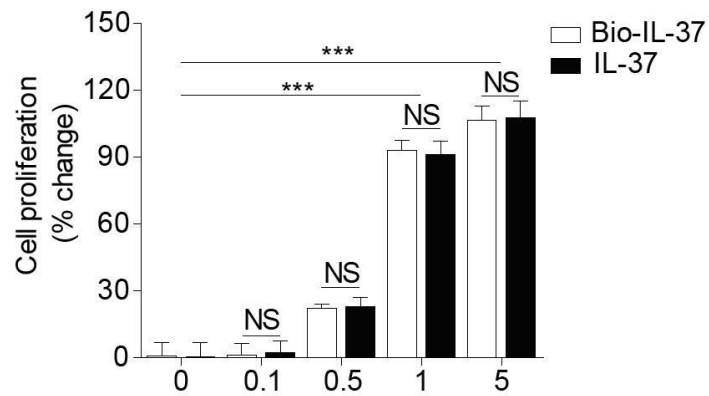
or

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## SUPPLEMENTAL MATERIAL

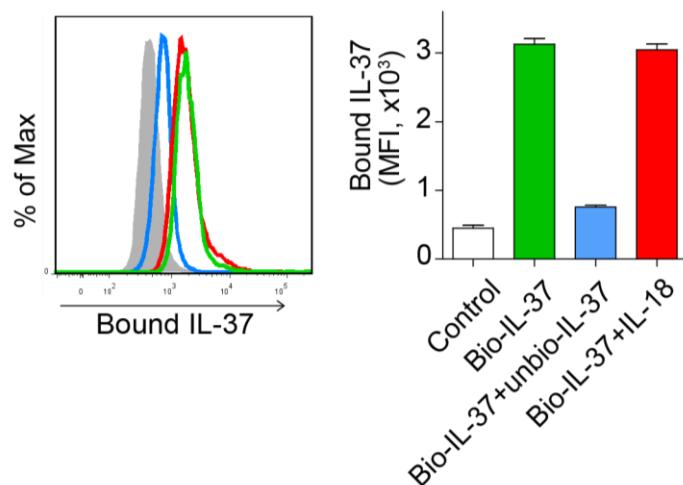


**Figure S1. The effect of TGF- $\beta$  family members on IL-37 expression.** HUVECs were pre-starved under serum-free conditions without supplemented growth factors overnight and then treated with indicated concentrations of factors (TGF- $\beta$ 1, BMP10 and GDF11) for 24 hours. IL-37 expression was examined by Western blot. Blots are representative of three experimental replicates.

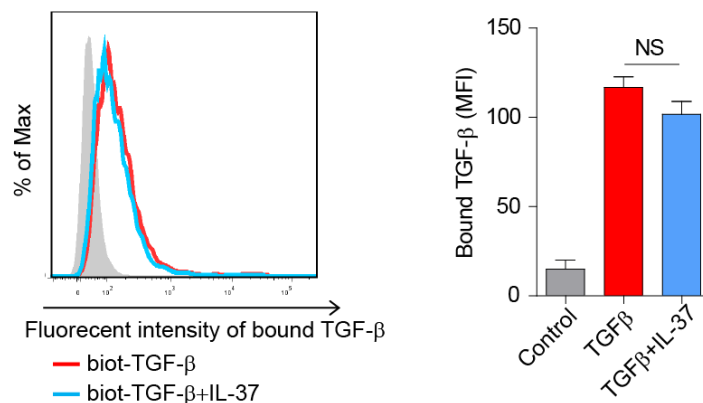


**Figure S2. The bioactivity of biotinylated IL-37 and unbiotinylated IL-37.**

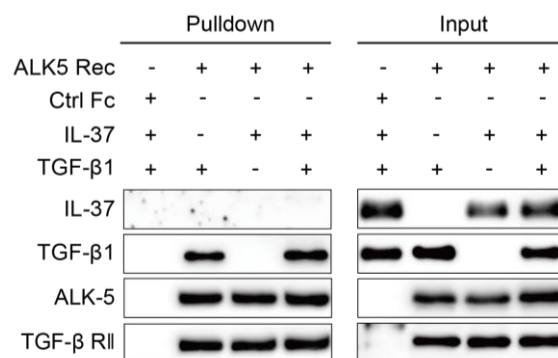
HUVECs were maintained in ECM containing 5% FBS without supplemented growth factors and stimulated with biotin-labeled or unlabeled IL-37 with indicated concentrations for 48 hours. Cell proliferation was determined by BrdU ELISA kit. Data are presented as mean  $\pm$  SEM ( $n = 5$  per group). \*\*\*  $P < 0.01$ . NS, not significant.



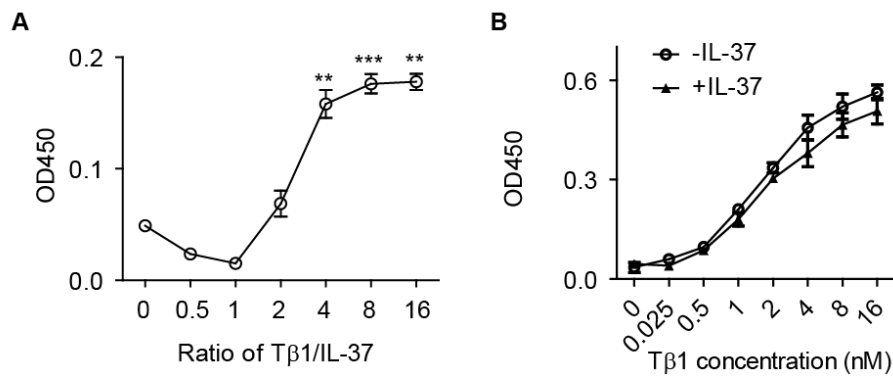
**Figure S3. IL-18 did not compete the binding of IL-37.** HUVECs were incubated with 1  $\mu$ M bio-IL-37 in the presence of 5 $\mu$ M unbiotinylated IL-37 (blue histogram) or unbiotinylated IL-18 (red histogram). Bound biot-IL-37 was determined by flow cytometry using Streptavidin-PE (SAv-PE). Gray histograms indicated cells incubated with SAv-PE alone. The mean fluorescence intensity (MFI) was quantified (n = 4). \*\*\*  $P < 0.01$ ; NS, not significant.



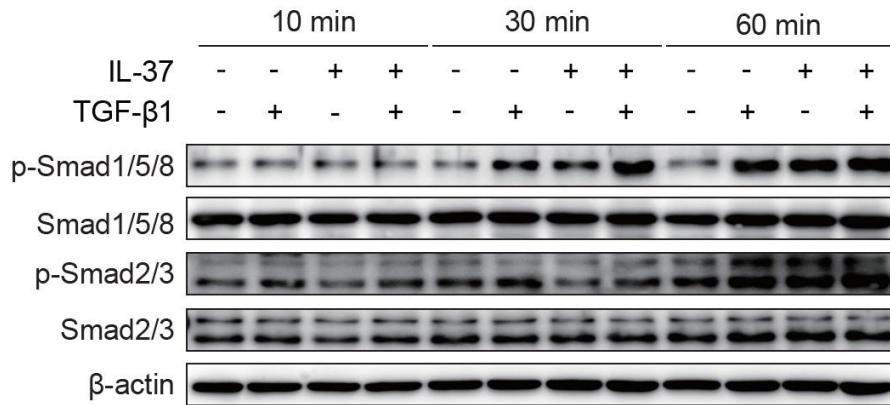
**Figure S4. IL-37 does not affect the binding of TGF- $\beta$  to HUVECs.** IL-37 does not affect the binding of TGF- $\beta$  to HUVECs. HUVECs were incubated with 1  $\mu$ M biot-TGF- $\beta$  in the presence (blue histogram) or absence (red histogram) of 1  $\mu$ M unbiotinylated IL-37. Bound biot-TGF- $\beta$  was determined by flow cytometry using SAv-PE. Gray histograms indicated cells incubated with SAv-PE alone. The mean fluorescence intensity (MFI) was quantified ( $n = 4$ ). NS, not significant.



**Figure S5. TGF- $\beta$  does not affect the binding of IL-37 to ALK-5 receptor complex.** The ALK5 receptor complex (pre-incubated ALK5-Fc and TGF- $\beta$  RII-Fc) or control Fc were conjugated to protein A/G beads, which were then incubated with IL-37 in the presence or absence of TGF- $\beta$ 1. Immobilized proteins were resolved by Western Blot. Blots are representative of three experimental replicates.

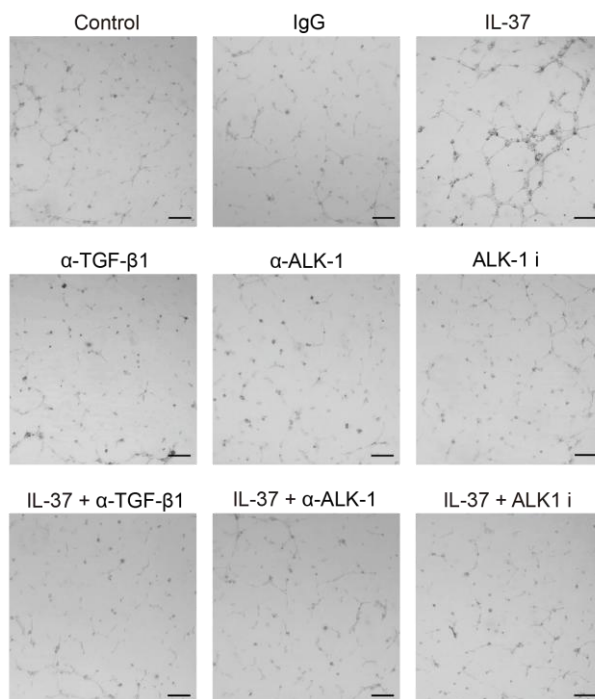


**Figure S6. The binding affinity of TGF- $\beta$  and IL-37 with the ALK1 receptor complex.** (A) The 96-well ELISA plates were coated with the ALK1 receptor complex and then incubated with 5 nmol/l of bio-IL-37 in the presence of increasing concentrations of TGF- $\beta$  (T $\beta$ 1). The binding of IL-37 was detected by Streptavidin-HRP.  $n = 4$ . (B) 96-well ELISA plates were coated with the ALK1 receptor complex and then incubated with increasing concentrations of biotinylated TGF- $\beta$  (biot-T $\beta$ 1) in the presence or absence of 20 nmol/l of IL-37. The binding of biot-TGF- $\beta$ 1 was detected by Streptavidin-HRP.  $n = 4$ . Data were presented as mean  $\pm$  SEM ( $n = 4$  per group). \*\*  $P < 0.01$ ; \*\*\*  $P < 0.001$ .

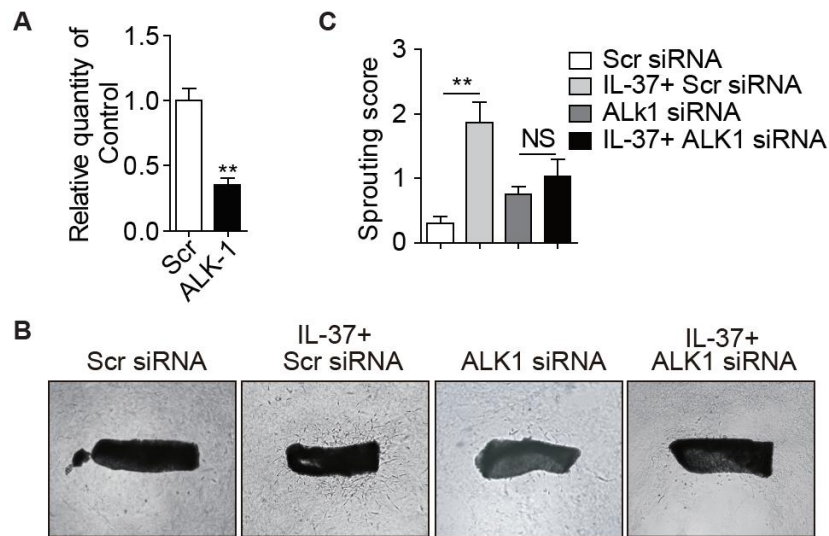


**Figure S7. IL-37 and TGF- $\beta$ 1 synergistically stimulated phosphorylation of Smad1/5/8 rather than Smad2/3.** HUVECs were pre-starved under serum-free conditions without supplemented growth factors overnight and then treated with IL-37 in the presence or absence of TGF- $\beta$ 1 for indicated time (10 min, 30 min and 60min). Phosphorylated Smad1/5/8 and Smad2/3 were determined by Western blot. Blots are representative of three experimental replicates.

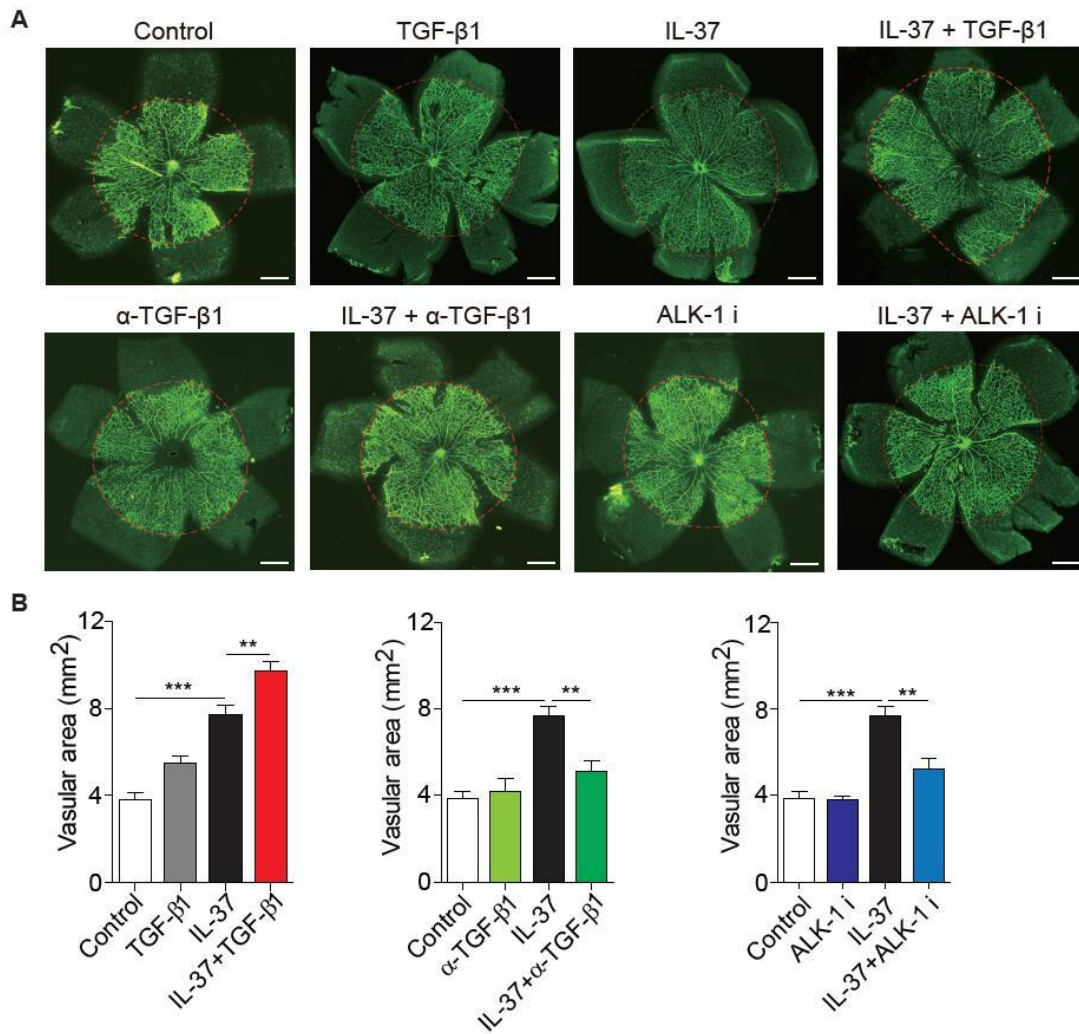




**Figure S8. IL-37 promotes tube formation of HUVECs through TGF-ALK1 signaling.** HUVECs were stimulated with IL-37 (1 ng/ml) in present or absent TGF- $\beta$ 1 antibody (10  $\mu$ g/ml), ALK-1 antibody (10  $\mu$ g/ml) and ALK-1 inhibitor (0.5  $\mu$ M) for 12 hours. Representative images of tube structure were shown. Scale bars, 100  $\mu$ m.



**Figure S9. Knockdown of ALK1 reduced IL-37 (1 ng/mL) stimulated vessel growth from aortic rings. (A)** Mouse aortic rings were transfected with 120 nmol/l scrambled siRNA or 120 nmol/l siALK1 composed of 40 nmol/l of the three antisense sequences using Lipofectamin RNAiMAX (Invitrogen) according to manufacturer's instructions. After overnight transfection, mRNA was isolated and *Alk1* mRNA level was quantified by quantitative PCR. Data were presented as mean  $\pm$  SEM ( $n = 5$  per group). \*\* $P < 0.01$ . **(B)** Knockdown of ALK1 by siRNA inhibited IL-37-stimulated vessel growth from aortic rings. **(C)** Aggregate analysis of the sprouting.  $n = 10$  per group. Data are presented as mean  $\pm$  SEM. \*\*  $P < 0.01$ . NS, not significant.



**Figure S10. IL-37 promoted developmental angiogenesis through TGF- $\beta$  signaling.** (A) Neonatal mice were administrated with IL-37 (1 ng/g bodyweight) with or without TGF- $\beta$ 1 (1 ng/g bodyweight) from postnatal day 1 to day 4. For blockade of TGF- $\beta$ 1 or ALK1, TGF- $\beta$ 1 neutralizing antibodies was administrated intraocularly at 0.5  $\mu$ g per eye and ALK1 inhibitor LDN193189 was administrated intraperitoneally at 2 mg/kg bodyweight. (n = 10 per group). (B) Vascular area of the retina whole mounts was assessed. Scale bars, 500  $\mu$ m. Data are presented as mean  $\pm$  SEM. \*  $P < 0.05$ , \*\*  $P < 0.01$  and \*\*\*  $P < 0.001$ .

**Additional Tables****Table S1. siRNA sequences targeting ALK1**

Primer name	Sense (5'-3')	antiense (3'-5')
ALK1_mouse_1	TGGTAGAGTGTGTGGGAAA	TTTCCCACACACTCTACCA
ALK1_mouse_2	CAGGAGAAGCAGCGGGATT	AATCCCGCTGCTTCTCCTG
ALK1_mouse_3	CCAGAGAAGCCCAAAGTGA	TCACTTTGGGCTTCTCTGG

**Table S2. Primer sequences of real-time quantitative PCR**

Primer name	Sequence (5' to 3')
h $\beta$ -actin_For	TTCCATATCGTCCCAGTTGGT
h $\beta$ -actin_Rev	CCAGGGCGTTATGGTAGGCA
m $\beta$ -actin For	GCTCGTTGCCAATAGTGATGACC
m $\beta$ -actin Rev	TGAGAGGGAAATCGTGCGTGAC
mALK1_For	GGCCTTTTGATGCTGTCTG
mALK1_Rev	ATGACCCCTGGCAGAATG
hld1_For	TTCCTCTGGTTGACTGTTGTTCTTC
hld1_Rev	CTCTCTAAACTCCCTACGCCTTGTT
hld3_For	GGAGCTTTTGCCACTGACTCG
hld3_Rev	CTCCAGGAAGGGATTTGGTGAAGT