

Supporting Information

A critical assessment of the synthesis and biological activity of p53/Hdm2 stapled peptide inhibitors

Running title: A critical evaluation of stapled peptides

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Instrument [BS_QT03] WSJ-507.08.12

Column: ACQUITY BEH300 C18 2.1x50mm 1.7 um at 80°C:
Gradient: from 5 to 98 % B in 4.4 min - Flow 0.8 mL/min

Eluent A: water + 0.05% TFA
Eluent B: acetonitrile + 0.04% TFA

4: UV Detector: TIC

4.328e+1
Range: 4.328e+1

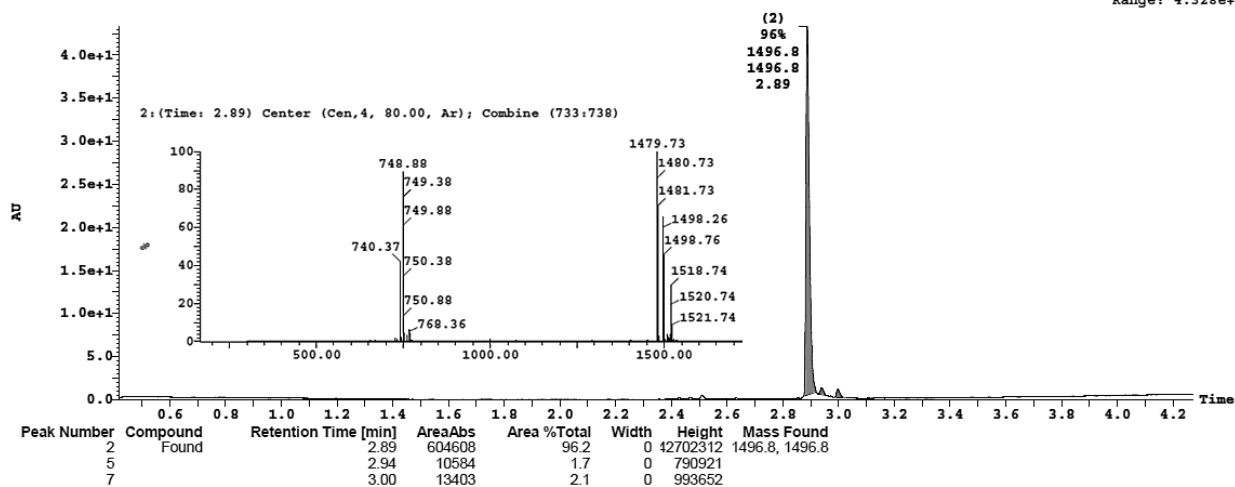


Figure S1: LC-MS of 1a

Instrument [BS_QT03] WSJ-507.08.12

Column: ACQUITY UPLC CORTECS C18 2.1x100mm 1.6 um at 80°C:
Gradient: from 5 to 98 % B in 4.4 min - Flow 0.8 mL/min

Eluent A: water + 0.05% TFA
Eluent B: acetonitrile + 0.04% TFA

4: UV Detector: 214

4.123
Range: 4.123

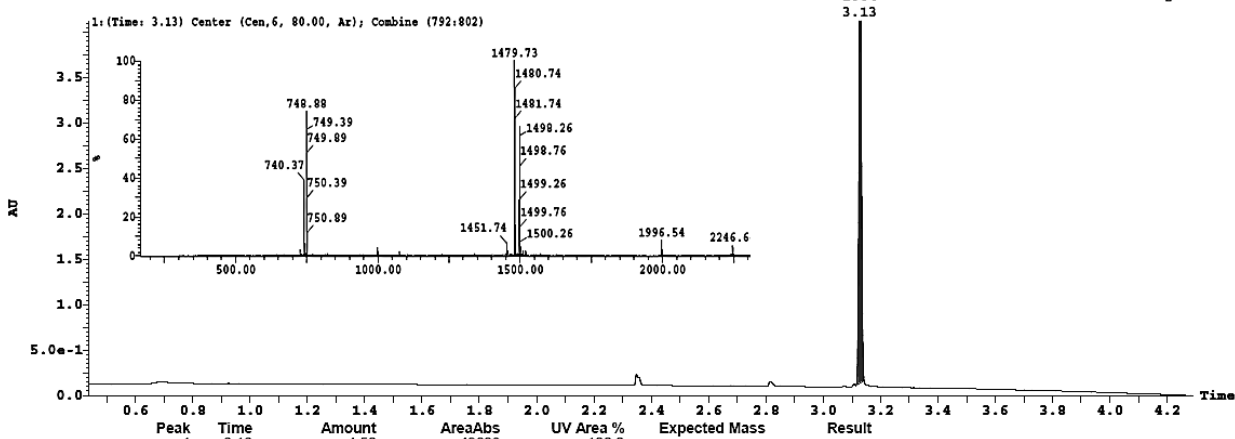


Figure S2: LC-MS of 1b

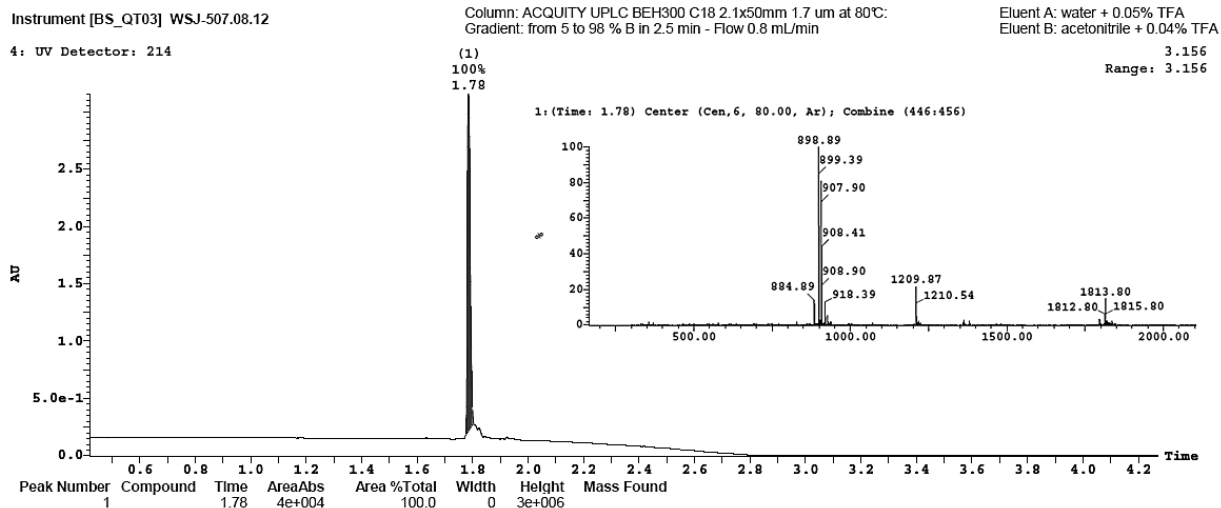


Figure S3: LC-MS of CF1a

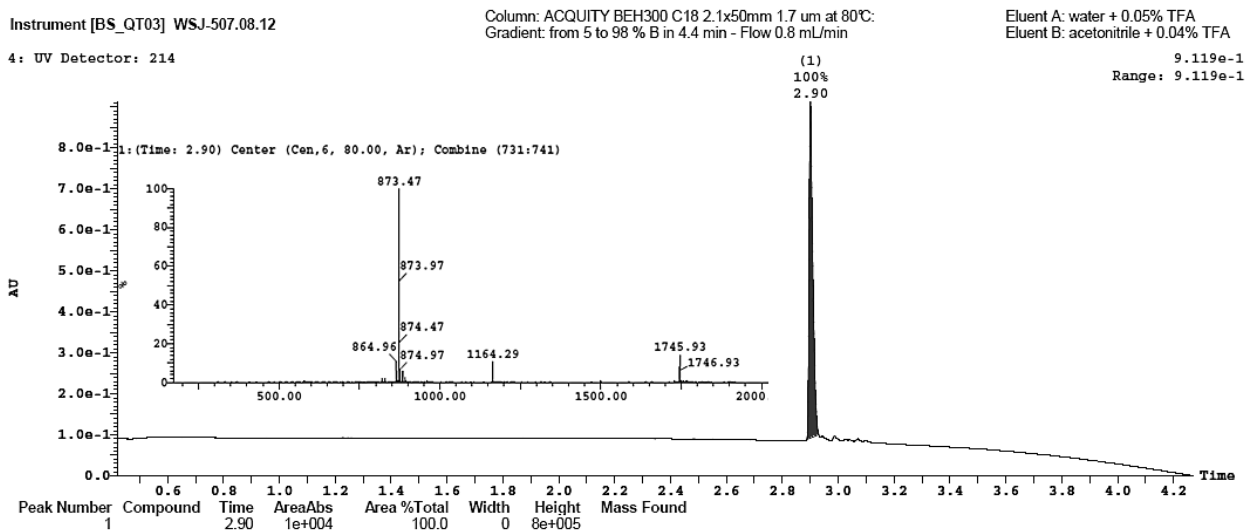


Figure S4: LC-MS of 2a

Instrument [BS_QT03] WSJ-507.08.12

Column: ACQUITY BEH300 C18 2.1x50mm 1.7 um at 80°C:
Gradient: from 5 to 98 % B in 4.4 min - Flow 0.8 mL/min

Eluent A: water + 0.05% TFA
Eluent B: acetonitrile + 0.04% TFA

4: UV Detector: 214

1.943
Range: 1.943

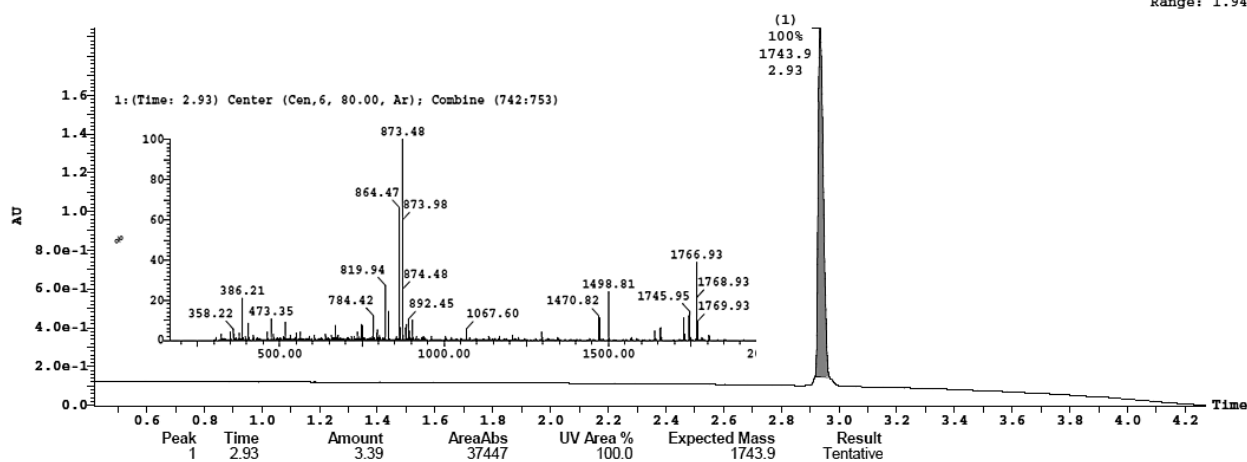


Figure S5: LC-MS of 2b

Instrument [BS_QT03] WSJ-507.08.12

Column: ACQUITY UPLC CORTECS C18 2.1x100mm 1.6 um at 80°C:
Gradient: from 5 to 98 % B in 2.5 min - Flow 0.8 mL/min

Eluent A: water + 0.05% TFA
Eluent B: acetonitrile + 0.04% TFA

4: UV Detector: 214

3.032
Range: 3.032

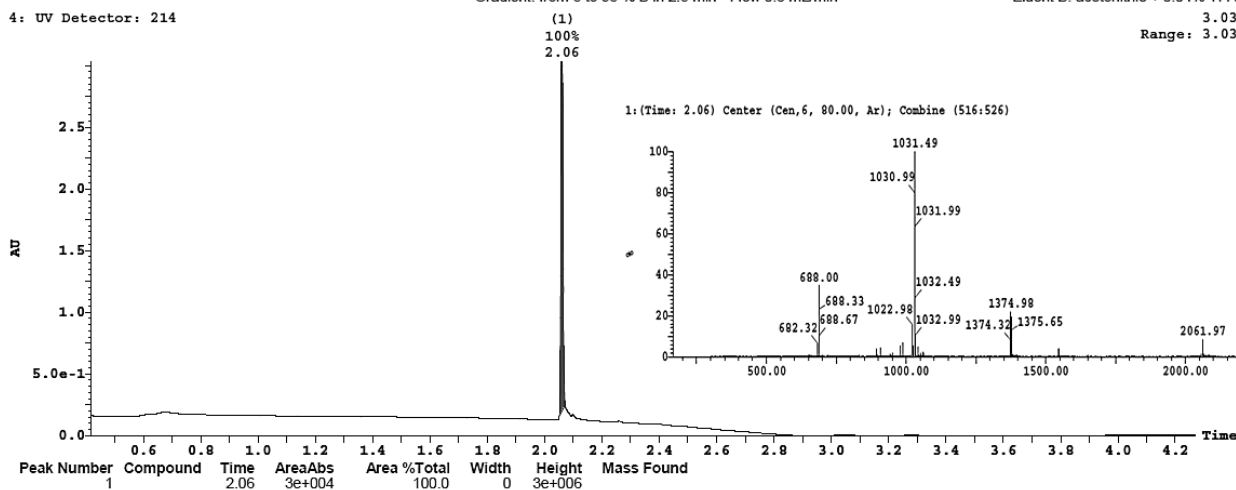


Figure S6: LC-MS of CF2a

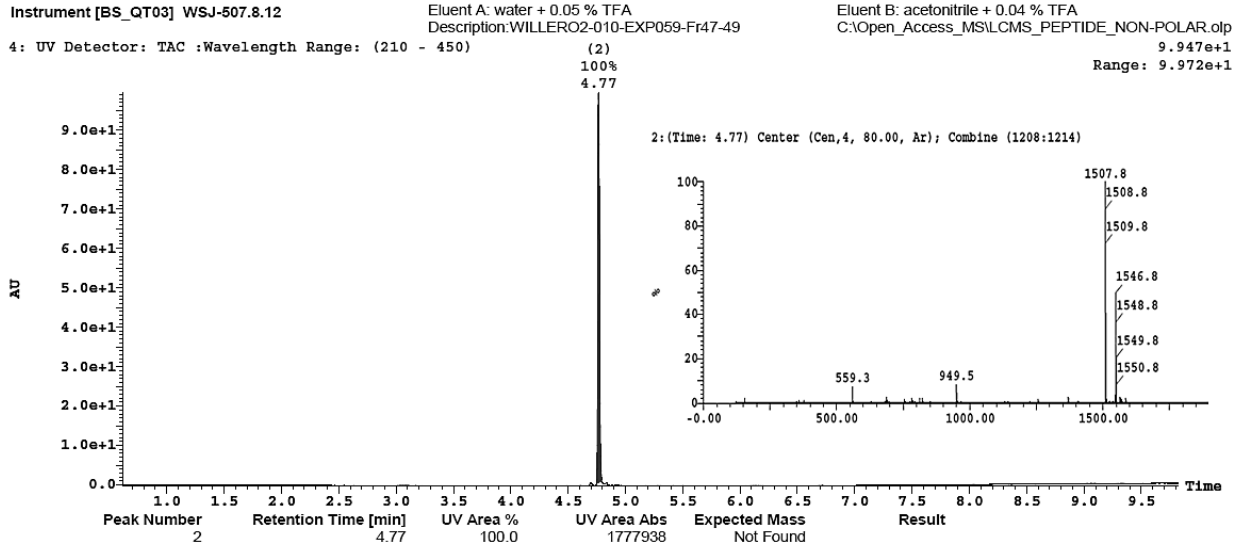


Figure S7: LC-MS of 3

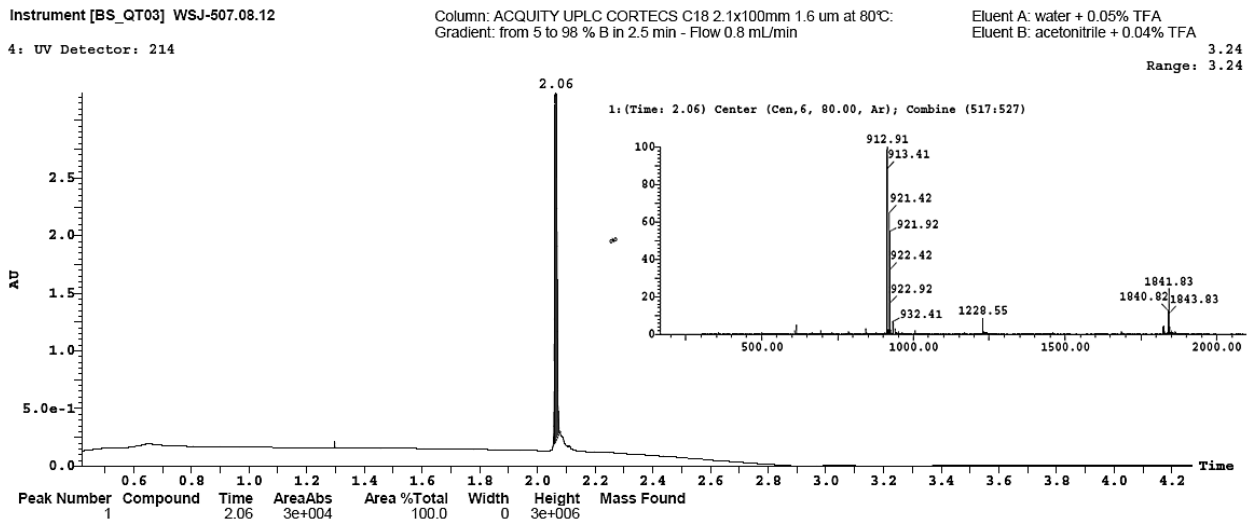


Figure S8: LC-MS of CF3

Instrument [BS_QT03] WSJ-507.08.12
4: UV Detector: 214

Column: ACQUITY UPLC CORTECS C18 2.1x100mm 1.6 um at 80°C:
Gradient: from 5 to 98 % B in 4.4 min - Flow 0.8 mL/min

Eluent A: water + 0.05% TFA
Eluent B: acetonitrile + 0.04% TFA

2.523
Range: 2.523

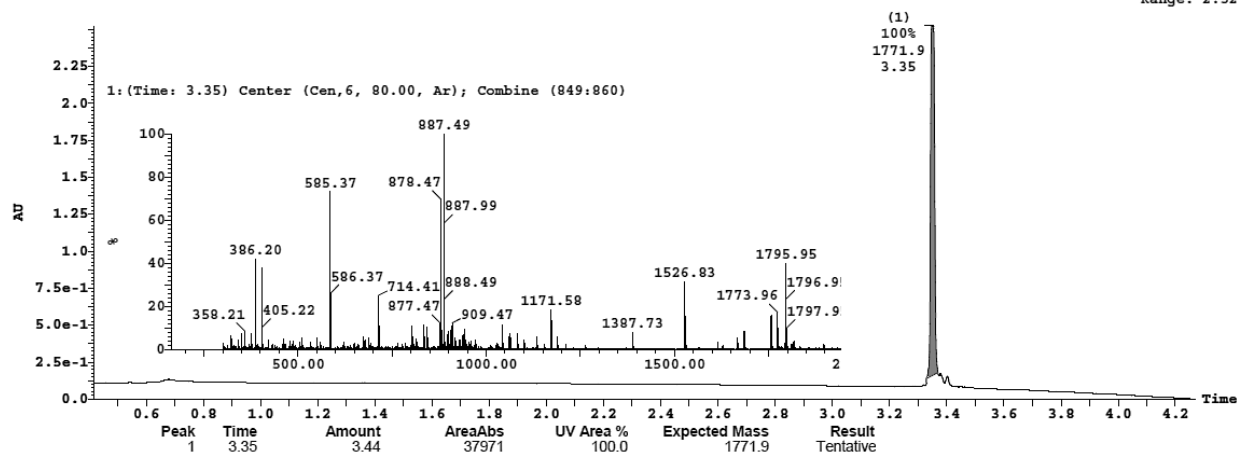


Figure S9: LC-MS of 4

Instrument [BS_QT03] WSJ-507.08.12
4: UV Detector: 214

Column: ACQUITY BEH300 C18 2.1x50mm 1.7 um at 80°C:
Gradient: from 5 to 98 % B in 4.4 min - Flow 0.8 mL/min

Eluent A: water + 0.05% TFA
Eluent B: acetonitrile + 0.04% TFA

9.065e-1
Range: 9.065e-1

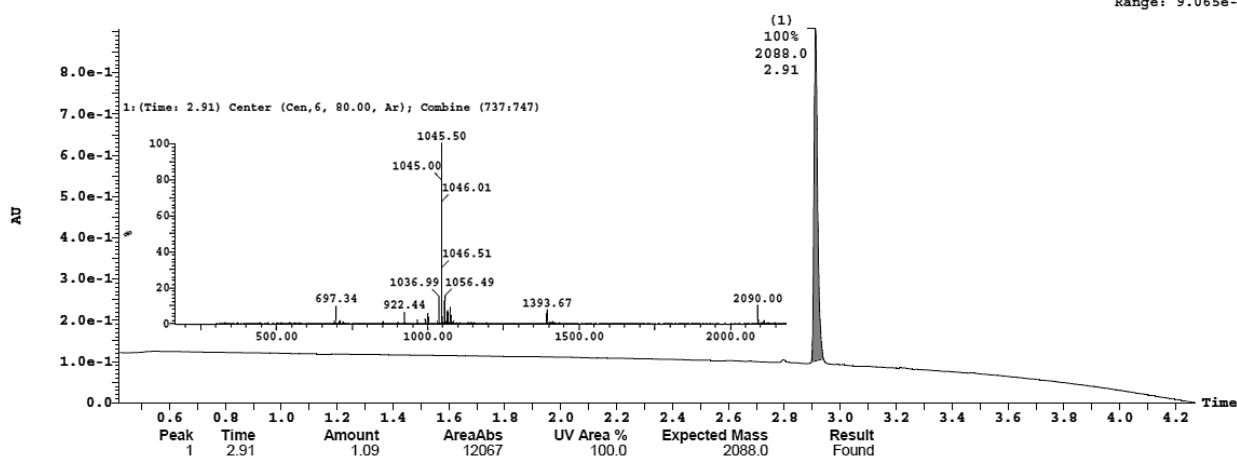


Figure S10: LC-MS of CF4

Table S1: Calculated and observed masses for the investigated peptides.

Compound	Formula	Calcd. mass	Found mass
1a	C ₇₅ H ₁₀₆ CIN ₁₃ O ₁₇	[M+H] ⁺ = 1498.2	1498.3
1b	C ₇₅ H ₁₀₆ CIN ₁₃ O ₁₇	[M+H] ⁺ = 1498.2	1498.3
CF1a	C ₉₄ H ₁₁₄ CIN ₁₃ O ₂₂	[M+H] ⁺ = 1814.4	1813.8
2a	C ₈₇ H ₁₂₅ N ₁₇ O ₂₁	[M+H] ⁺ = 1746.0	1745.9
2b	C ₈₇ H ₁₂₅ N ₁₇ O ₂₁	[M+H] ⁺ = 1746.0	1745.9
CF2a	C ₁₀₆ H ₁₃₃ N ₁₇ O ₂₆	[M+H] ⁺ = 2062.3	2062.0
3	C ₇₇ H ₁₁₀ CIN ₁₃ O ₁₇	[M-H ₂ O] ⁺ = 1507.2*	1507.8
CF3	C ₉₆ H ₁₁₈ CIN ₁₃ O ₂₂	[M+H] ⁺ = 1842.5	1841.8
4	C ₈₉ H ₁₂₉ N ₁₇ O ₂₁	[M+H] ⁺ = 1774.1	1774.0
CF4	C ₁₀₈ H ₁₃₇ N ₁₇ O ₂₆	[M+H] ⁺ = 2090.3	2090.0

*Loss of water is attributed to the ionization process during MS analysis.

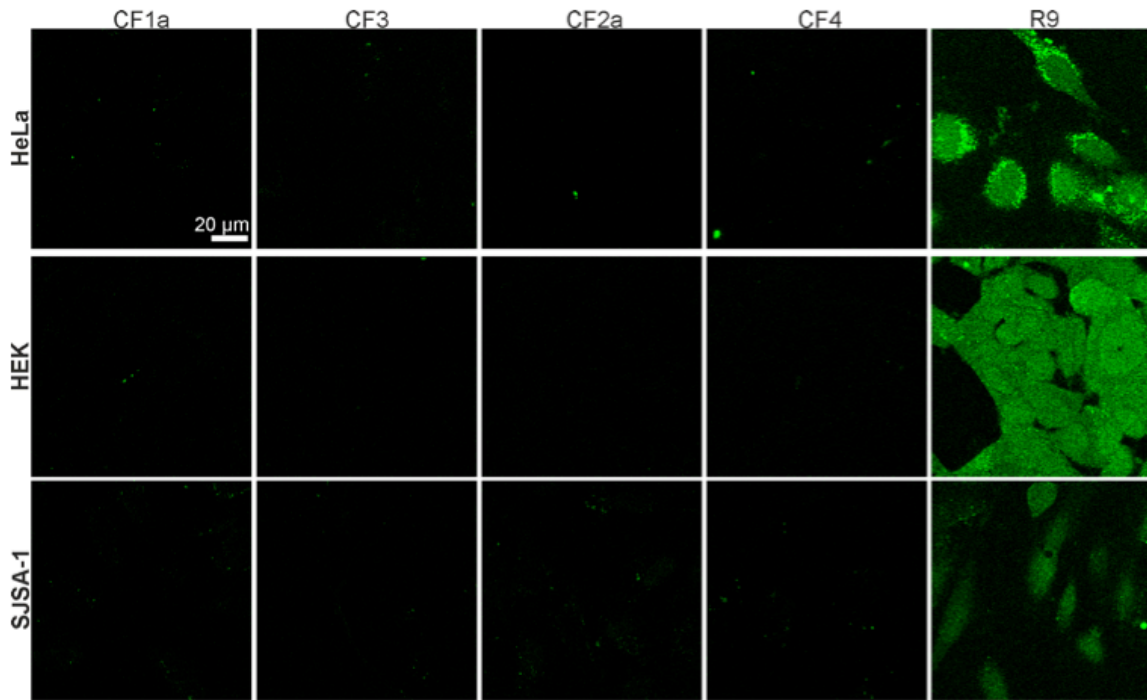


Figure S11: Intracellular distribution of stapled peptides after a 30 min incubation. Cells were incubated with 5 μM of the peptides in RPMI + 10 % FCS, washed and confocal images were recorded. Fluorescein-labeled R9 was used as a reference. Scale bar denotes 20 μm . The figure shows one representative experiment of two independent repetitions.

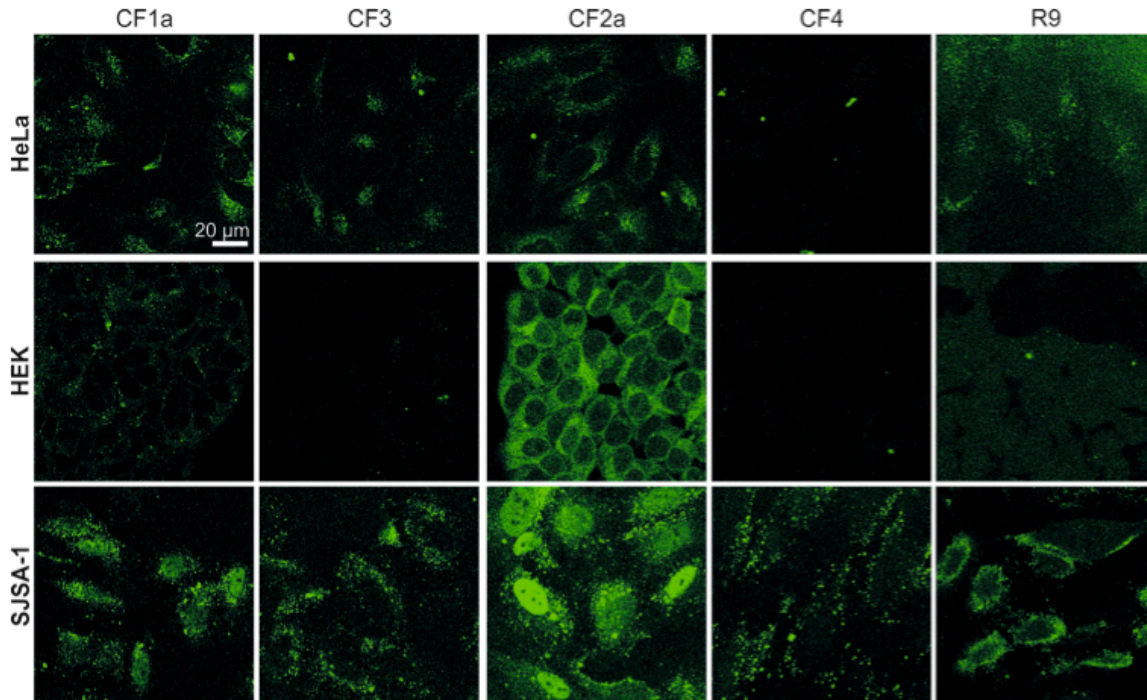


Figure S12: Intracellular distribution of stapled peptides after a 24 h incubation. Cells were incubated with 5 μM of the CF-labeled peptides in RPMI + 10 % FCS, washed and confocal images were recorded. R9 was used as a control CPP. The bars indicate the size. In

comparison to 20 μM , different contrast settings were used to improve visualization. The figure shows one representative experiments of three independent repetitions.

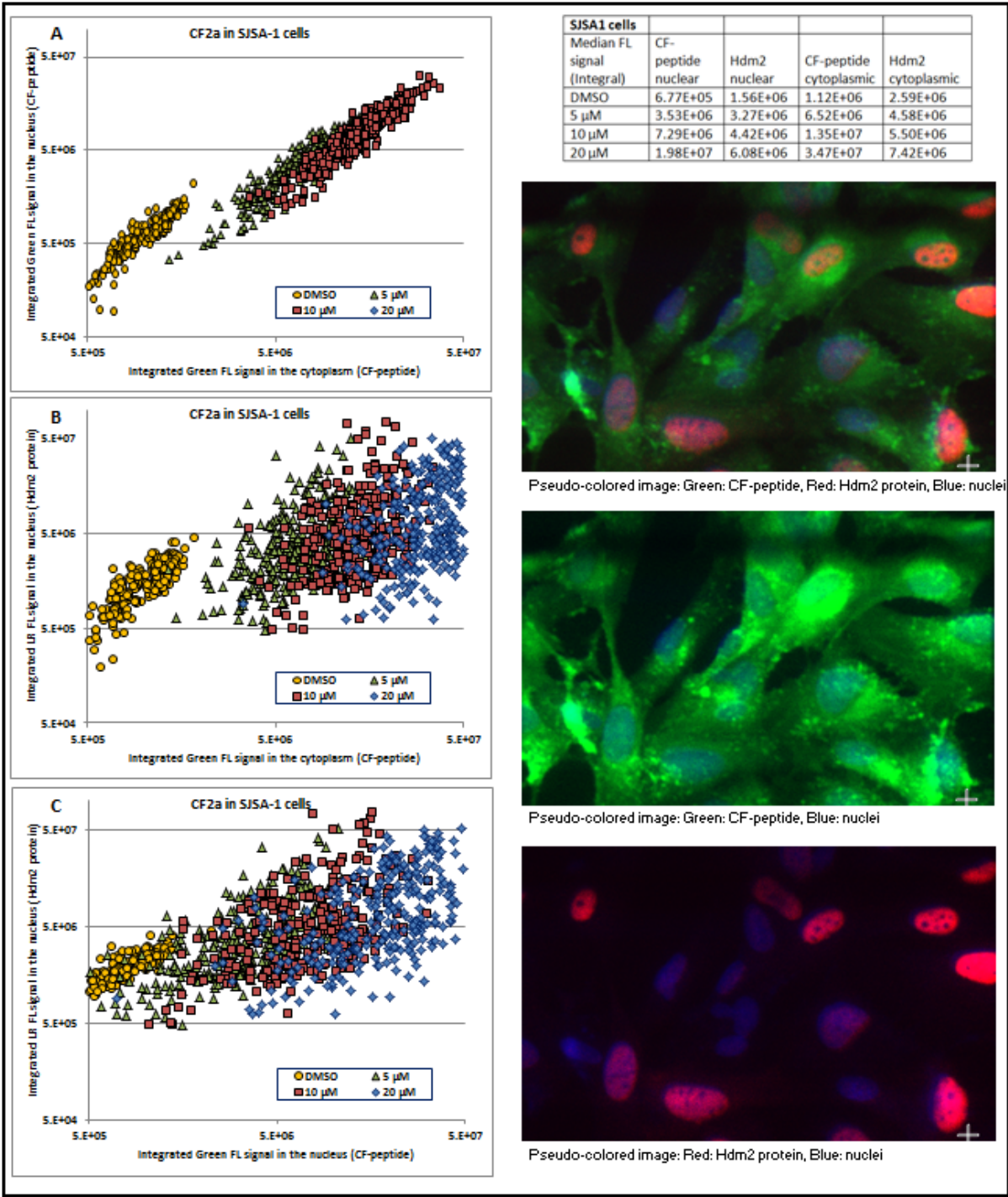


Figure S13: Colocalization of CF2a and Hdm2 in SJS1-1 cells. After treatment of cells with the carboxyfluorescein-labeled peptide at the indicated concentrations or DMSO control, cells were fixed, stained for detection of nuclei using DAPI and for Hdm2 using immunofluorescence and imaged by multi-channel fluorescence microscopy (right panels). (A) Correlation of integrated fluorescein fluorescence in the nucleus and cytoplasm. (B, C)

Correlation of integrated Hdm2 fluorescence in the nucleus versus carboxyfluorescein fluorescence in the cytoplasm (B) and in the nucleus (C).

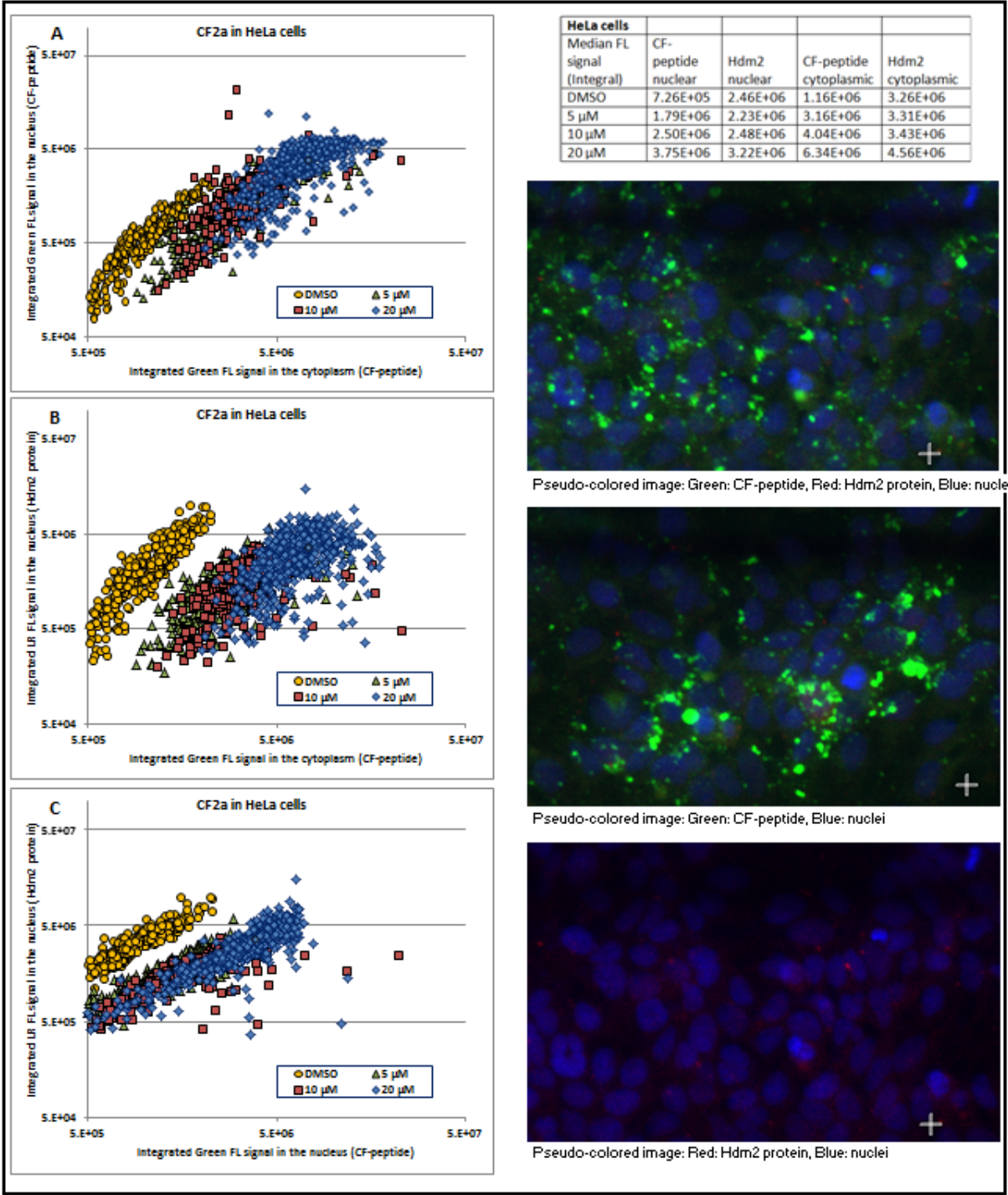


Figure S14: Colocalization of CF2a and Hdm2 in HeLa cells. After treatment of cells with the carboxyfluorescein-labeled peptide at the indicated concentrations or DMSO control, cells were fixed, stained for detection of nuclei using DAPI and for Hdm2 using immunofluorescence and imaged by multi-channel fluorescence microscopy (right panels). (A) Correlation of integrated fluorescein fluorescence in the nucleus and cytoplasm. (B, C)

Correlation of integrated Hdm2 fluorescence in the nucleus versus carboxyfluorescein fluorescence in the cytoplasm (B) and in the nucleus (C).

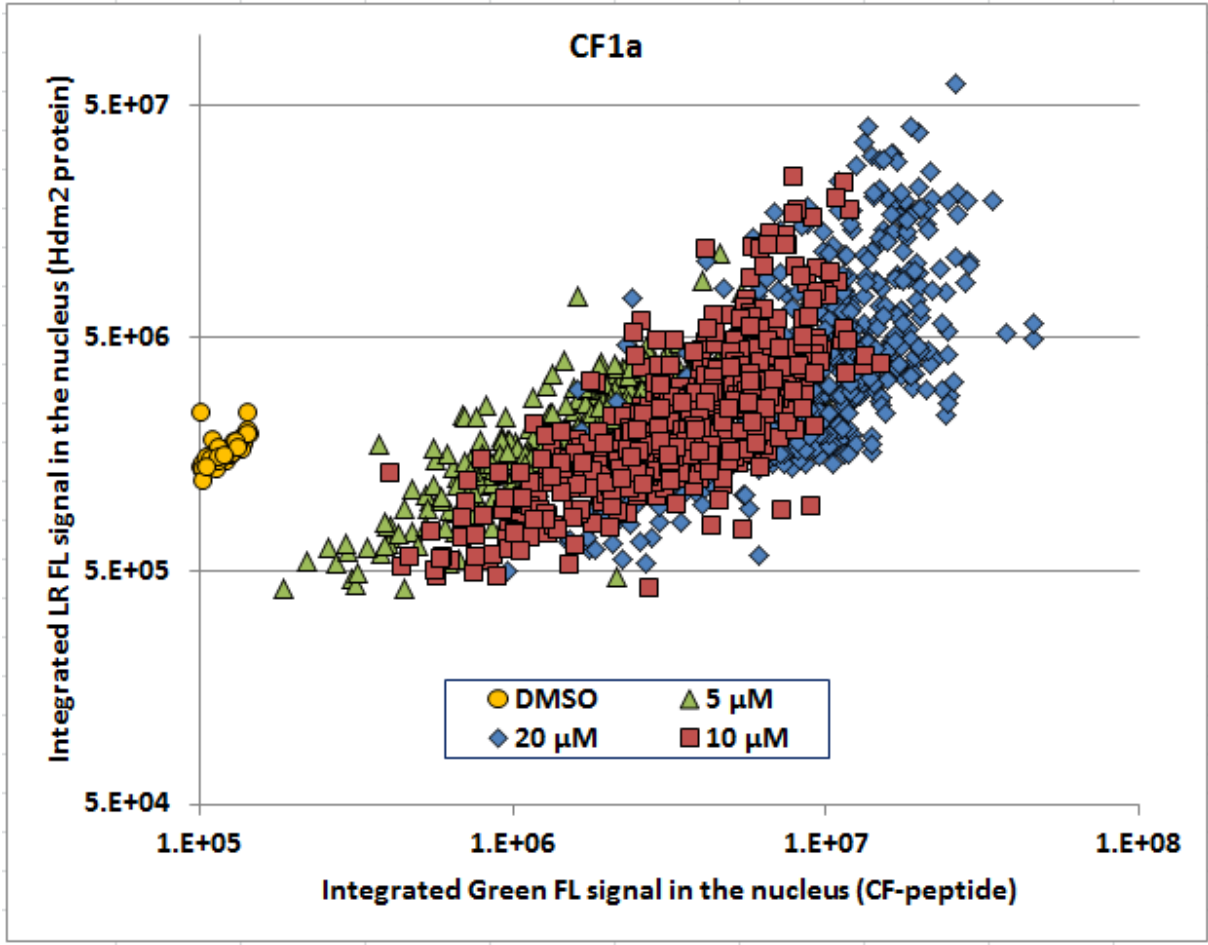


Figure S15: Co-localization of the labeled peptide CF1a with the PD marker Hdm2. After treatment of SJSA1 cells for 24 h with the carboxyfluorescein-labeled peptide at the indicated concentrations, the cells were fixed and stained with a specific antibody against the PD marker Hdm2 linked with a red fluorescent label.