Supplementary Materials

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SUPPLEMENTAL MATERIALS

Supplemental Figures







Fig. S2. Relative abundance of *B. vulgatus* **ATCC 8482 and** *B. dorei* **DSM 17855 in gnotobiotic mice across experimental stages and treatment groups.** COPRO-Seq analysis of the effects of the micronutrient-deficient versus sufficient diets on the relative abundances of (**A**) *B. vulgatus* and (**B**) *B. dorei.* Each data point indicates the relative abundance of the indicated organism in each mouse at the indicated experimental day and are coded by color to indicate experimental group. Grey-shaded boxes indicate time points for which mice were exposed to the specified micronutrient-deficient diet. Mean

values \pm SD are indicated by bars. See table S4 and table S5 for the results of statistical tests of observed differences within and between treatment groups.



Fig. S3. Characterization of the *B. vulgatus* ATCC 8482 INSeq library. (A) Distribution of Tn mutants in the genome of *B. vulgatus* ATCC 8482. Key; Track 1 (innermost circle), plot of GC skew for the genome using a sliding window size of 10kb (yellow, GC skew > 0; purple, GC skew < 0); Track 2 (middle circle), genes with transposon insertions are depicted in light grey; Track 3 (outermost circle), all genes in the genome are shown with those represented in polysaccharide utilization loci (PULs) colored green, components of capsular polysaccharide synthesis (CPS) loci colored red, and all others colored light grey. (B) Estimating the saturation of *B. vulgatus* ATCC 8482 transposon mutant libraries by *in silico* simulation.



Fig. S4. Maximum likelihood phylogenetic tree of BVU0240/AcrR orthologs identified in human gut-associated *Bacteroides* **and other members of the family Bacteroidaceae.** Multiple amino acid sequence alignments were generated using ClustalX and exported in PHYLIP format. PhyML was used to generate the maximum likelihood tree, with bootstrap support (out of 100) indicated for given nodes.



В		K _d (FPA)		
	Compound	AcrR _{BV}	AcrR _{BD}	
	DMSO (1%)	18.5 ± 6.2 nM	25.5 ± 10.9 nM	
	Retinol (125 µM)	49.5 ± 11.6 nM	56.5 ± 25.1 nM	

	K _d (FPA)	
Compound	AcrR _{BV}	AcrR _{BD}
DMSO (1%)	9.4 ± 2.4 nM	19.3 ± 6.8 nM
CA (100 µM)	14.7 ± 3.5 nM	48.2 ± 14.5 nM
βMCA (100 μM)	12.3 ± 3.9 nM	53.5 ± 33.4 nM
DCA (100 µM)	20.2 ± 10.0 nM	74.0 ± 35.5 nM
GCA (100 µM)	23.9 ± 6.6 nM	51.0 ± 13.7 nM
ΤβΜCA (100 μΜ)	65.2 ± 29.6 nM	28.2 ± 7.1 nM
TCA (100 µM)	12.5 ± 5.0 nM	45.2 ± 14.4 nM

Fig. S5. DNA-binding characteristics of $AcrR_{BV}$ and $AcrR_{BD}$ in the presence and absence of possible

effectors. (A) Electrophoretic Mobility Shift Assays (EMSA) of $AcrR_{BV}$ and $AcrR_{BD}$ in the presence and absence of retinol and various bile acid species. (B,C) Binding characteristics (K_d) derived from curve fitting of Fluorescence Polarization (FP) Assay data for $AcrR_{BV}$ and $AcrR_{BD}$ in the presence and absence of (B) retinol or (C) bile acid species. FP data are reported as mean values ± SEM for experiments performed in triplicate. Abbreviations: ROL, retinol; CA, β MCA, β -muricholic acid; DCA, deoxycholic acid; GCA, glycocholic acid; TCA, taurocholic acid; T β MCA, tauro- β -muricholic acid.