

Supporting Information

Development of Small Molecules with a Non-Canonical Binding Mode to HIV-1 Trans Activation Response (TAR) RNA

Fardokht A. Abulwerdi,^{†,‡} Matthew D. Shortridge,[#] Joanna Sztuba-Solinska,[‡] Robert Wilson,[†]
Stuart F. J. Le Grice,[‡] Gabriele Varani,[#] and John S. Schneekloth, Jr.*[†]

[†]Chemical Biology Laboratory, National Cancer Institute, Frederick, Maryland 21702, USA

[‡]Basic Research Laboratories, National Cancer Institute, Frederick, Maryland 21702, USA

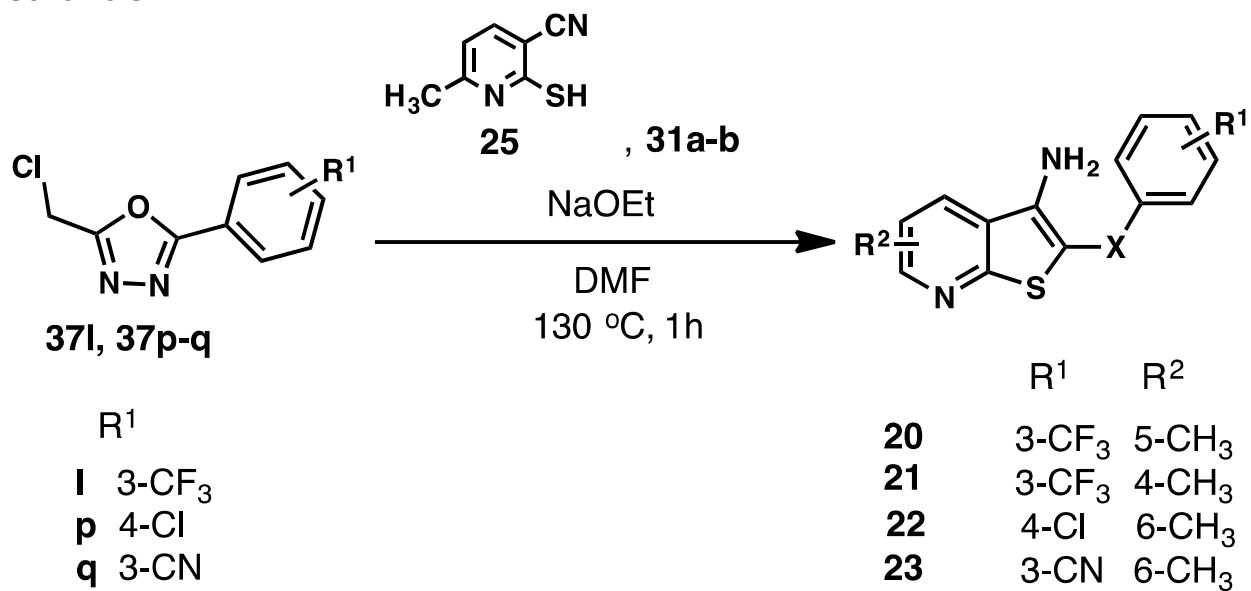
[#]Department of Chemistry, University of Washington, Seattle, Washington 98195, USA

*E-mail: schneeklothjs@mail.nih.gov

Table of Contents

Scheme S1	S2
Scheme S2	S2
Table S1.	S3
Figure S1	S3
Chemistry	S6
References	S10
1D 1H and 13C NMR spectra	S12

Scheme S1



Scheme S2

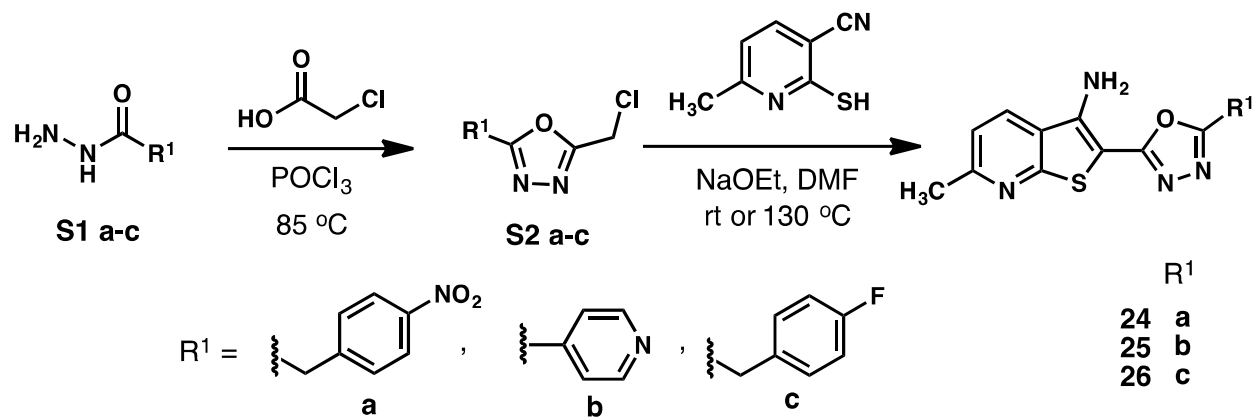


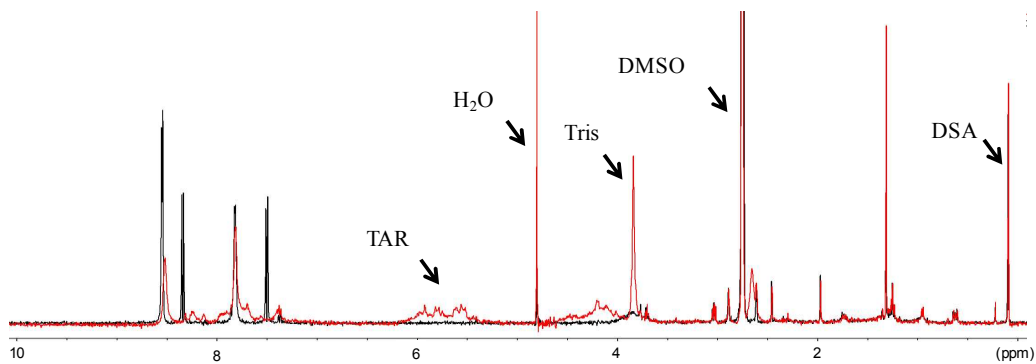
Table S1.Affinity measurement and Tat/TAR displacement assay for oxadiazole analogs **20-26**

Cpd	25-2AP TAR ^a K _{d app} ± SD (μM)	23-2AP TAR ^a K _{d app} ± SD (μM)	FRET ^a IC ₅₀ ± SD (μM)
20	>100	NT	NT
21	>100	NT	NT
22	>100	NT	NT
23	>400	>400	>400
24	>400	NT	123.6 ± 49.1
25	>400	NT	>300
26	>400	NT	>400

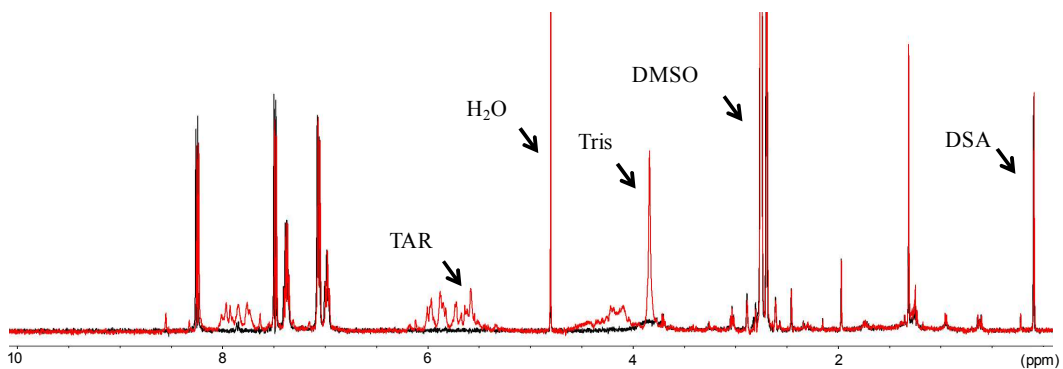
NT: not tested

^aHighest concentration evaluated was 400 μM unless otherwise indicated. Values represent the average of three replicates, ± standard deviation.**Figure S1**

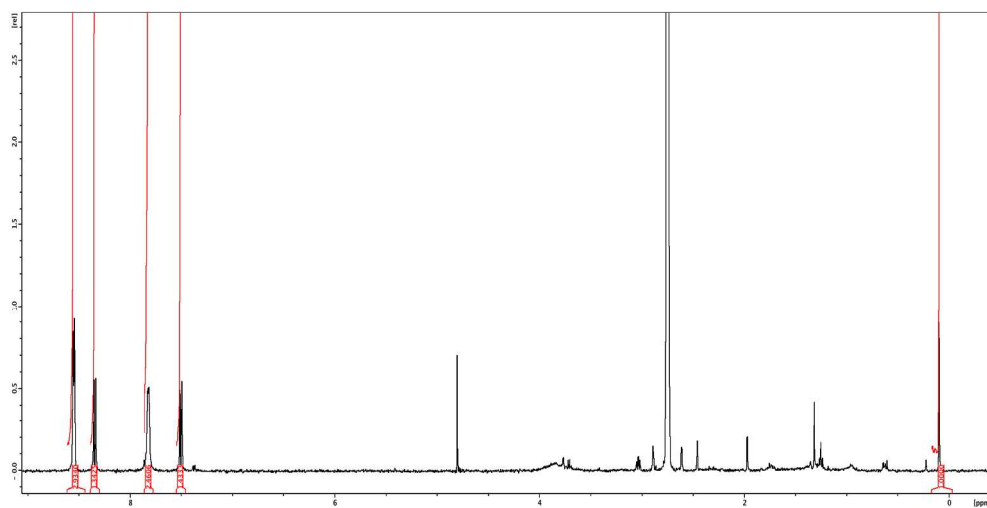
A



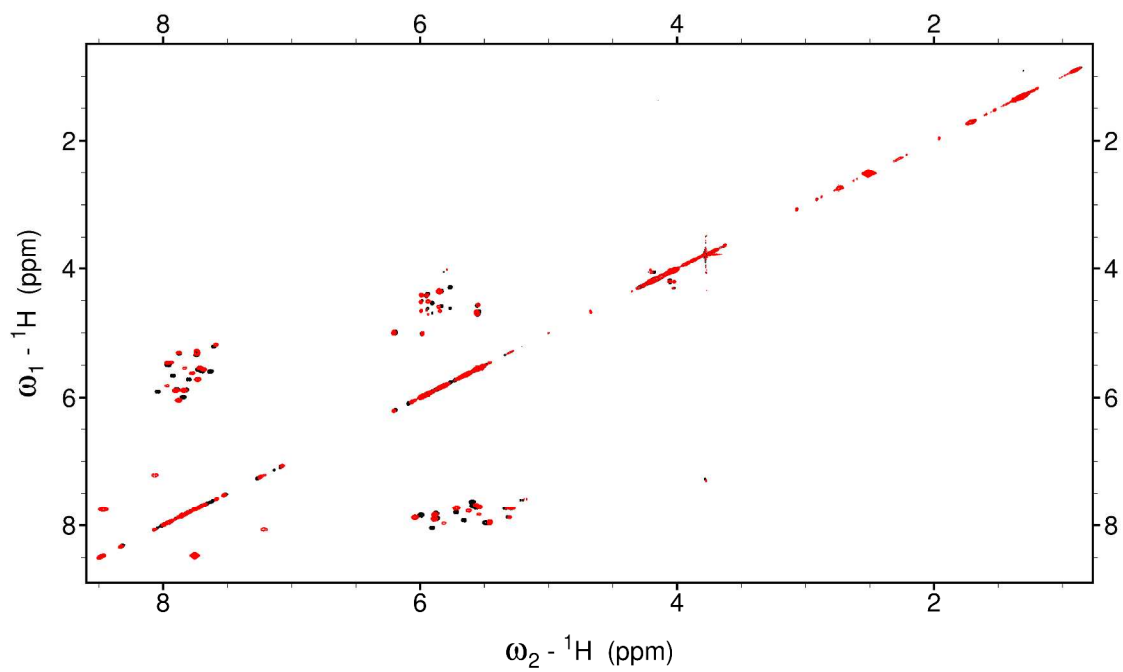
B



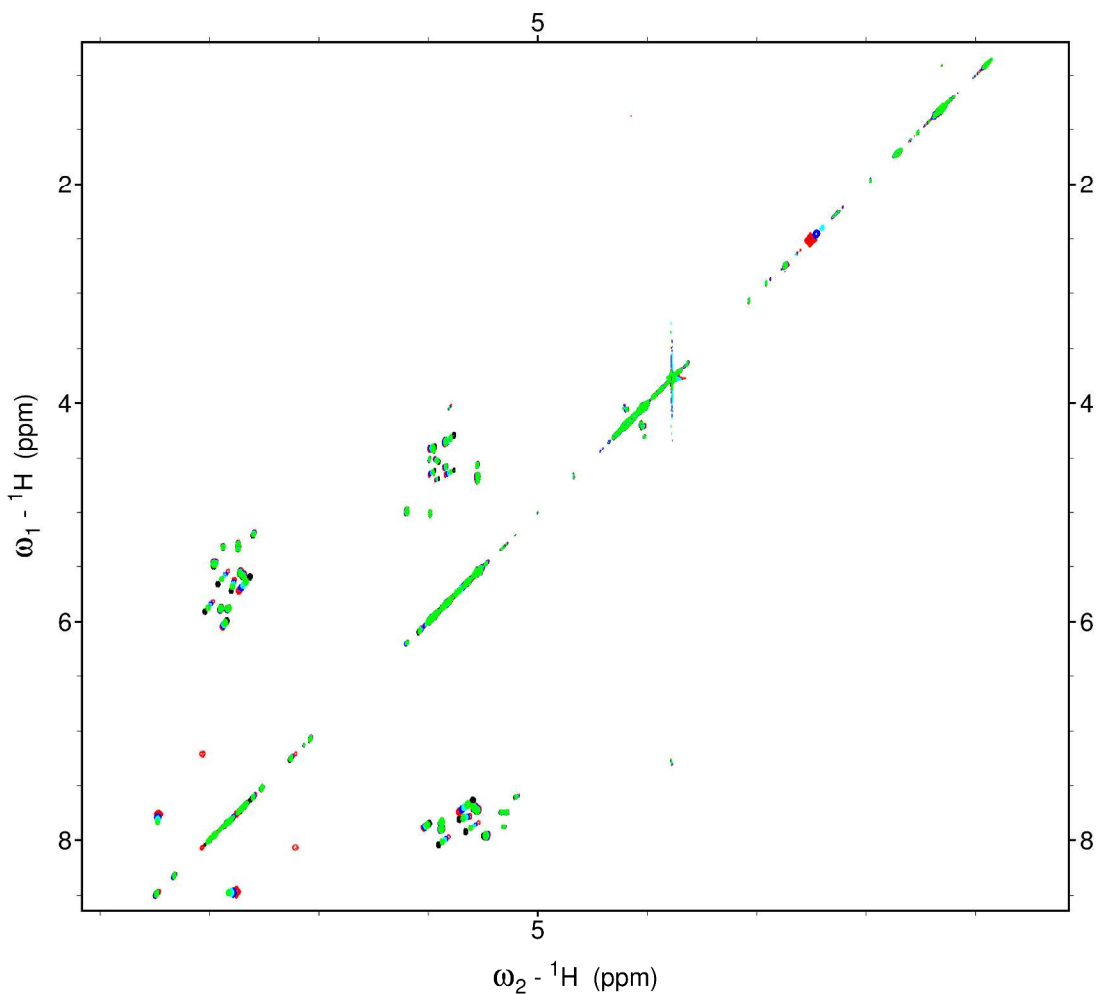
C



D



E



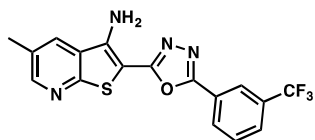
A) Ligand-detected full ${}^1\text{H}$ NMR spectra of **2** (150 μM) and B) **14** (150 μM) in the presence (red spectrum) and absence of TAR hairpin (10 μM) (black spectrum). C) Free ligand **2** in NMR screening buffer showing DSA reference integrating to 100 μM while the ligand signal integrates to \sim 150 μM . D) Full TOCSY spectrum showing free RNA (111 μM , black) and RNA bound to 500 μM **2** (red). E) Full TOCSY spectrum showing free RNA (111 μM , black) titrated from 100-500 μM **2**, with 500 μM **2** (red).

Chemistry

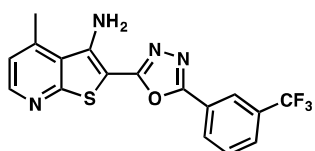
Chemicals were purchased from commercial sources and used without further purification unless otherwise noted. Anhydrous solvents were prepared by passage over activated alumina. Thin-layer chromatography was conducted with E. Merck silica gel 60 F254 precoated plates (0.25 mm) and visualized by exposure to UV light (254 nm). Flash column chromatography was performed using normal phase or reverse phase on a CombiFlash® Rf 200i (Teledyne Isco Inc). ¹H NMR spectra were recorded at 400 or 500 MHz, and are reported relative to deuterated solvent signals. Data for ¹H NMR spectra are reported as follows: chemical shift (δ ppm), multiplicity, coupling constant (Hz), and integration. ¹³C NMR spectra were recorded at 100 or 125 MHz. Data for ¹³C NMR spectra are reported in terms of chemical shift. All NMR spectra were standardized to the NMR solvent signal as reported previously.¹ Analytical LC/MS was performed using a Shimadzu LCMS-2020 Single Quadrupole utilizing a Kinetex 2.6 μm C18 100 Å (2.1 x 50 mm) column obtained from Phenomenex Inc. Runs employed a gradient of 0→90% ACN/0.1% aqueous formic acid over 4 minutes at a flow rate of 0.2 mL/min. Purities of final compounds were assessed by analytical reverse-phase HPLC using Agilent Proshell 120 EC-C18 (4.6 mm x 50 mm; particle size 2.7 μm). Runs employed a gradient of 10% ACN/water (1 min), 10-90% ACN/water (6 min), and 90% ACN/water (2 min) flow = 1 mL/min. High-resolution LC/MS analyses acquired on an Agilent 6520 Accurate-Mass Q-TOF LC/MS System, (Agilent Technologies, Inc., Santa Clara, CA) equipped with a dual electro-spray source, operated in the positive-ion mode. Separation was performed on Zorbax 300SB-C18 Poroshell column (2.1 mm x 150 mm; particle size 5 μm). All final compounds were purified to >95% purity unless otherwise stated.

Representative procedure for oxadiazolyl thienopyridine preparation

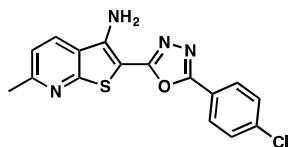
To a stirring solution of desired mercaptonicotinonitrile (0.28 mmol) in anhydrous DMF (2 mL) was added commercially available chloromethyl substituted-1,3,4-oxadiazole (0.24 mmol) and sodium ethoxide (0.63 mmol). The mixture was heated to 130 °C for 1 h. The mixture was then cooled to room temperature and solid precipitated at the bottom of the flask. The solid was collected by filtration and rinsed with cold MeOH and dried *in vacuo* to give the desired product.



5-Methyl-2-(5-(3-(trifluoromethyl)phenyl)-1,3,4-oxadiazol-2-yl)thieno[2,3-*b*]pyridin-3-amine (20). Synthesized using the representative procedure and with **31b** and **37m**. Precipitated solid was filtered, rinsed with cold MeOH, and dried *in vacuo* to give the title compound (23 mg, 56% yield) as a yellow solid. Purity 85%. ^1H NMR (500 MHz, DMSO- d_6) δ 8.56 – 8.55 (m, 1H), 8.44 (s, 1H), 8.42 (d, J = 8.0 Hz, 1H), 8.37 (s, 1H), 8.02 (d, J = 7.8 Hz, 1H), 7.90 – 7.86 (m, 1H), 7.22 (s, 2H), 2.46 (s, 3H). ^{13}C NMR (125 MHz, DMSO- d_6) δ 162.28, 160.64, 156.87, 150.93, 143.22, 130.92, 130.81, 130.56, 129.28, 125.56, 124.43, 122.85, 87.38, 17.96. ESI HRMS: 377.06828 (M+H) $^+$.

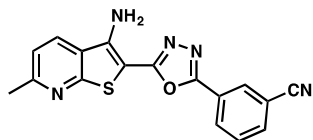


4-Methyl-2-(5-(3-(trifluoromethyl)phenyl)-1,3,4-oxadiazol-2-yl)thieno[2,3-*b*]pyridin-3-amine (21). Synthesized using the representative procedure and with **31a** and commercially available **37m**. Precipitated solid was filtered, rinsed with cold MeOH, and dried *in vacuo* to give the title compound (19 mg, 39% yield) as a yellow solid. Purity 96%. ^1H NMR (500 MHz, DMSO- d_6) δ 8.50 (d, J = 4.7 Hz, 1H), 8.38 (d, J = 7.9 Hz, 1H), 8.33 (s, 1H), 8.03 (d, J = 7.8 Hz, 1H), 7.89 (t, J = 7.8 Hz, 1H), 7.27 (d, J = 4.7 Hz, 1H), 6.82 (s, 2H), 2.86 (s, 3H). ^{13}C NMR (125 MHz, DMSO- d_6) δ 162.39, 160.42, 160.23, 149.53, 145.43, 145.01, 130.89, 130.52, 130.22, 129.97, 128.24 (q, J = 3.3 Hz), 124.76, 124.58, 124.34, 122.80 (q, J = 3.7 Hz), 122.26, 88.43, 19.87. ESI HRMS: 377.06831 (M+H) $^+$.



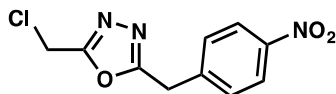
2-(5-(4-Chlorophenyl)-1,3,4-oxadiazol-2-yl)-6-methylthieno[2,3-*b*]pyridin-3-amine (22). Synthesized using the representative procedure and with **25** and commercially available 2-(chloromethyl)-5-(4-chlorophenyl)-1,3,4-oxadiazole, **37p**. Precipitated solid was filtered, rinsed with cold MeOH, and dried *in vacuo* to give the title compound (47 mg, 76% yield) as a yellow solid. Purity 97%. ^1H NMR (500 MHz, DMSO- d_6) δ 8.47 (d, J = 8.3 Hz, 1H), 8.11 (d, J = 8.6 Hz, 2H), 7.69 (d, J = 8.6 Hz, 2H), 7.40 (d, J = 8.3 Hz, 1H), 7.20 (s, 2H), 2.61 (s, 3H). ^{13}C NMR (125

MHz, DMSO-*d*₆) δ 162.08, 160.75, 159.26, 159.22, 143.52, 136.33, 131.16, 129.56, 128.31, 123.38, 122.19, 119.89, 86.09, 24.21. ESI HRMS: 343.04218 (M+H)⁺.



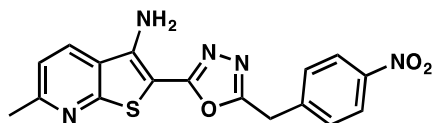
3-(5-(3-Amino-6-methylthieno[2,3-*b*]pyridin-2-yl)-1,3,4-oxadiazol-2-yl)benzonitrile (23).

Synthesized using the representative procedure and with **25** and commercially available 3-(5-(chloromethyl)-1,3,4-oxadiazol-2-yl)benzonitrile, **37q**. Precipitated solid was filtered, rinsed with cold MeOH, and dried *in vacuo* to give the title compound (20 mg, 50% yield) as a yellow solid. Purity 94%. ¹H NMR (500 MHz, DMSO-*d*₆) δ 8.57 (s, 1H), 8.47 (d, *J* = 8.3 Hz, 1H), 8.42 – 8.39 (m, 1H), 8.09 (d, *J* = 7.8 Hz, 1H), 7.82 (t, *J* = 7.9 Hz, 1H), 7.39 (d, *J* = 8.3 Hz, 1H), 7.21 (s, 2H), 2.61 (s, 3H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 162.34, 160.20, 159.35, 159.30, 143.61, 134.94, 131.19, 130.89, 130.66, 129.96, 124.58, 123.38, 119.85, 117.95, 112.60, 86.04, 24.21. ESI HRMS: 334.07596 (M+H)⁺.



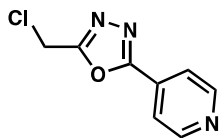
Representative procedure for chloromethyl substituted 1,3,4-oxadiazole preparation

2-(Chloromethyl)-5-(4-nitrobenzyl)-1,3,4-oxadiazole (S2a). Synthesized using reported procedure with modification.² A round bottom flask was charged with commercially available 2-(4-nitrophenyl)acetohydrazide, **S1a**, (103 mg, 0.5 mmol), chloroacetic acid (49 mg, 0.52 mmol) and phosphoryl chloride (2 mL). The reaction mixture was heated at 85 °C overnight. The flask was removed from the oil bath and allowed to cool to room temperature. Excess phosphoryl chloride was removed *in vacuo* and crushed ice (5 mL) was added slowly to the reaction mixture followed by slow addition of solid sodium bicarbonate while keeping the flask on ice. The mixture was diluted with H₂O (10 mL) and extracted with EtOAc (10 mL x 2) and finally washed with brine (10 mL). The organic layer was dried (Na₂SO₄), filtered, and concentrated *in vacuo*. Crude was purified by silica gel column chromatography (hexanes/EtOAc) to give **S2a** (72 mg, 57% yield) as a clear oil which solidified to a white solid upon standing. ¹H NMR (400 MHz, CDCl₃) δ 8.23 (d, *J* = 8.7 Hz, 2H), 7.51 (d, *J* = 8.6 Hz, 2H), 4.66 (s, 2H), 4.34 (s, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 165.57, 163.25, 147.76, 140.46, 130.04, 124.36, 32.88, 31.72. ESI MS: *m/z* 253.55, 255.55 (M+H)⁺.

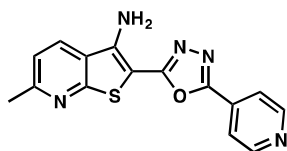


6-Methyl-2-(5-(4-nitrobenzyl)-1,3,4-oxadiazol-2-yl)thieno[2,3-*b*]pyridin-3-amine (24).

Synthesized using the representative procedure and with **25** and **S2a**. Reaction was performed at room temperature and was completed in 30 min. The mixture was diluted with H₂O (10 mL) and extracted with EtOAc (10 mL x 2) and finally washed with brine (10 mL). The organic layer was dried (Na₂SO₄), filtered, and concentrated *in vacuo*. Crude was purified by silica gel column chromatography (hexanes/EtOAc) to give **24** (9 mg, 41% yield) as an orange solid. Purity 97%. ¹H NMR (500 MHz, DMSO-*d*₆) δ 8.42 (d, *J* = 8.3 Hz, 1H), 8.25 (d, *J* = 8.8 Hz, 2H), 7.68 (d, *J* = 8.8 Hz, 2H), 7.38 (d, *J* = 8.3 Hz, 1H), 7.10 (s, 2H), 4.54 (s, 2H), 2.59 (s, 3H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 162.15, 161.85, 159.08, 159.00, 146.77, 143.12, 142.52, 131.07, 130.37, 123.82, 123.33, 119.87, 85.89, 30.27, 24.17. ESI HRMS: 368.08196 (M+H)⁺.



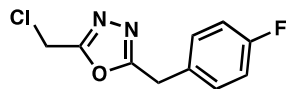
2-(Chloromethyl)-5-(pyridin-4-yl)-1,3,4-oxadiazole (S2b). Synthesized using a similar procedure used to prepare **S2a** except using isoniazid, **S1b**. Crude (112 mg, 79% yield), a dark green solid, was used in the next step without further purification. ¹H NMR (500 MHz, DMSO-*d*₆) δ 8.90 – 8.87 (m, 2H), 8.03 – 8.01 (m, 2H), 5.19 (s, 2H). ¹³C NMR (125 MHz, DMSO-*d*₆) δ 163.95, 163.34, 150.32, 130.83, 120.68, 33.12. ESI MS: *m/z* 195.45, 197.45 (M+H)⁺.



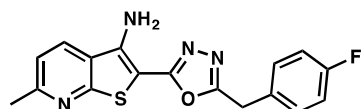
6-Methyl-2-(5-(pyridin-4-yl)-1,3,4-oxadiazol-2-yl)thieno[2,3-*b*]pyridin-3-amine (25).

Synthesized using the representative procedure and with **25** and **S2b**. Reaction was heated for 1 hr and was let to cool down to room temperature. The mixture was diluted with H₂O (15 mL) and extracted with EtOAc (10 mL x 3) and MeOH:CH₂Cl₂ 3:7 (10 mL x 2). The organic layer was dried (Na₂SO₄), filtered, and concentrated *in vacuo*. Crude was purified by silica gel column chromatography (hexanes/EtOAc) to give **25** (3.2 mg, 10% yield) as a light yellow solid. Purity 99%. ¹H NMR (500 MHz, DMSO-*d*₆) δ 8.84 (d, *J* = 6.0 Hz, 2H), 8.49 (d, *J* = 8.3 Hz, 1H), 8.04 (d,

$J = 6.1$ Hz, 2H), 7.40 (d, $J = 8.3$ Hz, 1H), 7.27 (s, 2H), 2.62 (s, 3H). ^{13}C NMR (125 MHz, $\text{DMSO-}d_6$) δ 162.73, 160.00, 159.50, 159.35, 150.86, 144.05, 131.28, 130.37, 123.32, 120.09, 119.94, 85.82, 24.23. ESI HRMS: 310.07548 (M+H) $^+$.



2-(Chloromethyl)-5-(4-fluorobenzyl)-1,3,4-oxadiazole (S2c). Synthesized using a similar procedure used to prepare **S2a** except using 4-fluorophenylacetic acid hydrazide, **S1c**. Crude (132 mg, 96% yield), a brown oil, was used in the next step without further purification. ^1H NMR (500 MHz, CDCl_3) δ 7.29 (dd, $J = 8.5, 5.3$ Hz, 2H), 7.05 (t, $J = 8.6$ Hz, 2H), 4.65 (s, 2H), 4.20 (s, 2H). ^{13}C NMR (125 MHz, CDCl_3) δ 166.86, 162.99, 162.45 (d, $J = 246.7$ Hz), 130.66 (d, $J = 8.2$ Hz), 129.05 (d, $J = 3.4$ Hz), 116.10 (d, $J = 21.7$ Hz), 32.96, 31.21. ESI MS: m/z 226.50 (M+H) $^+$.



2-(5-(4-Fluorobenzyl)-1,3,4-oxadiazol-2-yl)-6-methylthieno[2,3-*b*]pyridin-3-amine (26). Synthesized using the representative procedure and with **25** and **S2c**. Reaction was heated to 130 °C and was completed in 40 min. The mixture was diluted with H_2O (10 mL) and extracted with EtOAc (10 mL x 2) and finally washed with brine (10 mL). The organic layer was dried (Na_2SO_4), filtered, and concentrated *in vacuo*. Crude was purified by silica gel column chromatography (hexanes/EtOAc) to give **26** (2 mg, 4% yield) as a yellow solid. Purity 97%. ^1H NMR (500 MHz, $\text{DMSO-}d_6$) δ 8.41 (d, $J = 8.3$ Hz, 1H), 7.45 – 7.40 (m, 2H), 7.37 (d, $J = 8.3$ Hz, 1H), 7.20 (t, $J = 8.9$ Hz, 2H), 7.08 (s, 2H), 4.34 (s, 2H), 2.59 (s, 3H). ^{13}C NMR (125 MHz, $\text{DMSO-}d_6$) δ 162.68, 162.00, 161.36 (d, $J = 243.1$ Hz), 159.01, 158.97, 143.00, 131.04, 130.89 (d, $J = 8.2$ Hz), 130.82 (d, $J = 3.2$ Hz), 123.34, 119.85, 115.51 (d, $J = 21.4$ Hz), 85.99, 29.70, 24.17. ESI HRMS: 341.08686 (M+H) $^+$.

References

1. Fulmer, G. R.; Miller, A. J. M.; Sherden, N. H.; Gottlieb, H. E.; Nudelman, A.; Stoltz, B. M.; Bercaw, J. E.; Goldberg, K. I. NMR chemical shifts of trace impurities: common laboratory solvents, organics, and gases in deuterated solvents relevant to the organometallic chemist. *Organometallics* **2010**, 29 (9), 2176-2179.

2. Zhang, M. Z.; Mulholland, N.; Beattie, D.; Irwin, D.; Gu, Y. C.; Chen, Q.; Yang, G. F.; Clough, J. Synthesis and antifungal activity of 3-(1,3,4-oxadiazol-5-yl)-indoles and 3-(1,3,4-oxadiazol-5-yl)methyl-indoles. *Eur. J. Med. Chem.* **2013**, *63*, 22-32.

1D ¹H and ¹³C NMR spectra

