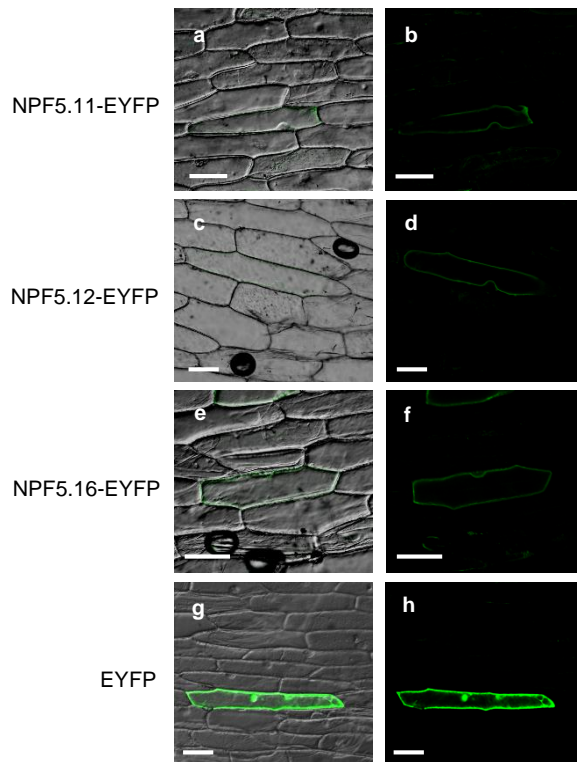


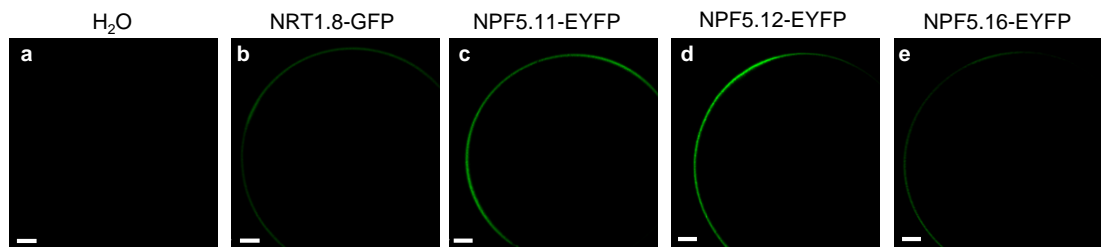
**Tonoplast-localized nitrate uptake transporters involved
in vacuolar nitrate efflux and reallocation in *Arabidopsis***

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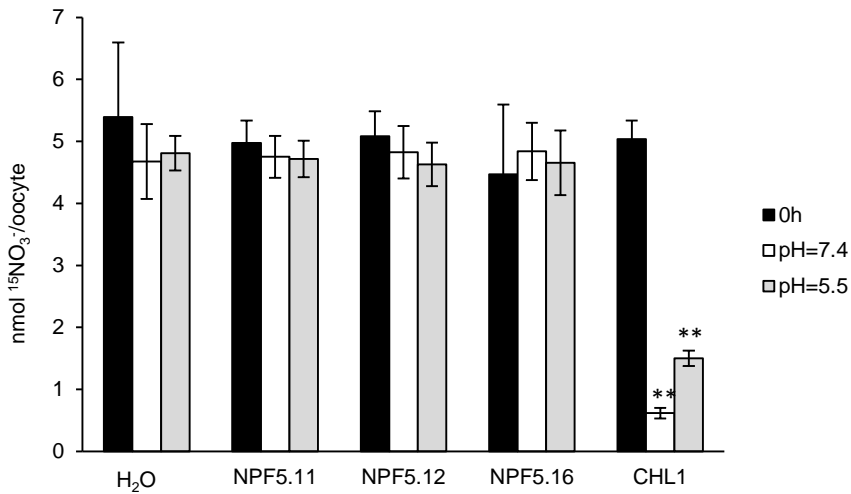
Supplementary Figure S1. Subcellular localization of NPF5.11, NPF5.12 and NPF5.16.

Onion epidermal cells transiently transformed with *NPF5.11-EYFP* (a,b), *NPF5.12-EYFP* (c,d), *NPF5.16-EYFP* (e,f) fusion or single *EYFP* (g,h) under the drive of cauliflower mosaic virus 35S promoter. Overlap images of EYFP fluorescence (green) and bright-field (a,c,e,g), EYFP fluorescence (b,d,f,h) images are shown. Bars = 100 μ m.



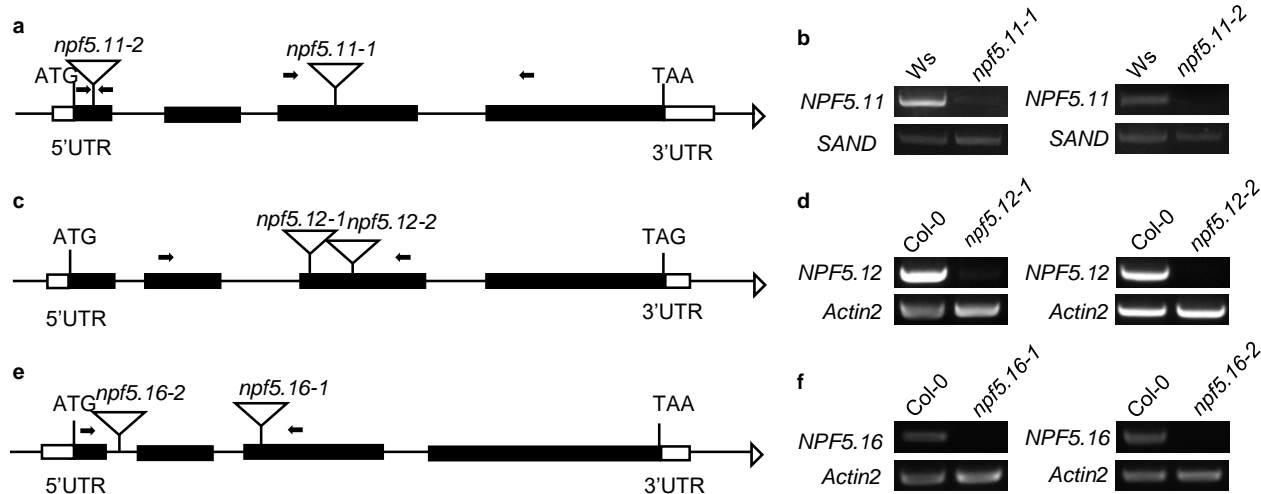
Supplementary Figure S2. Expression of *NPF5.11*, *NPF5.12* and *NPF5.16* in oocytes.

Oocytes were injected with H₂O (a) or cRNA of *NRT1.8-GFP* (b), *NPF5.11-EYFP* (c), *NPF5.12-EYFP* (d), *NPF5.16-EYFP* (e) and imaged after incubation for 2 days. Bars = 100 μ m.



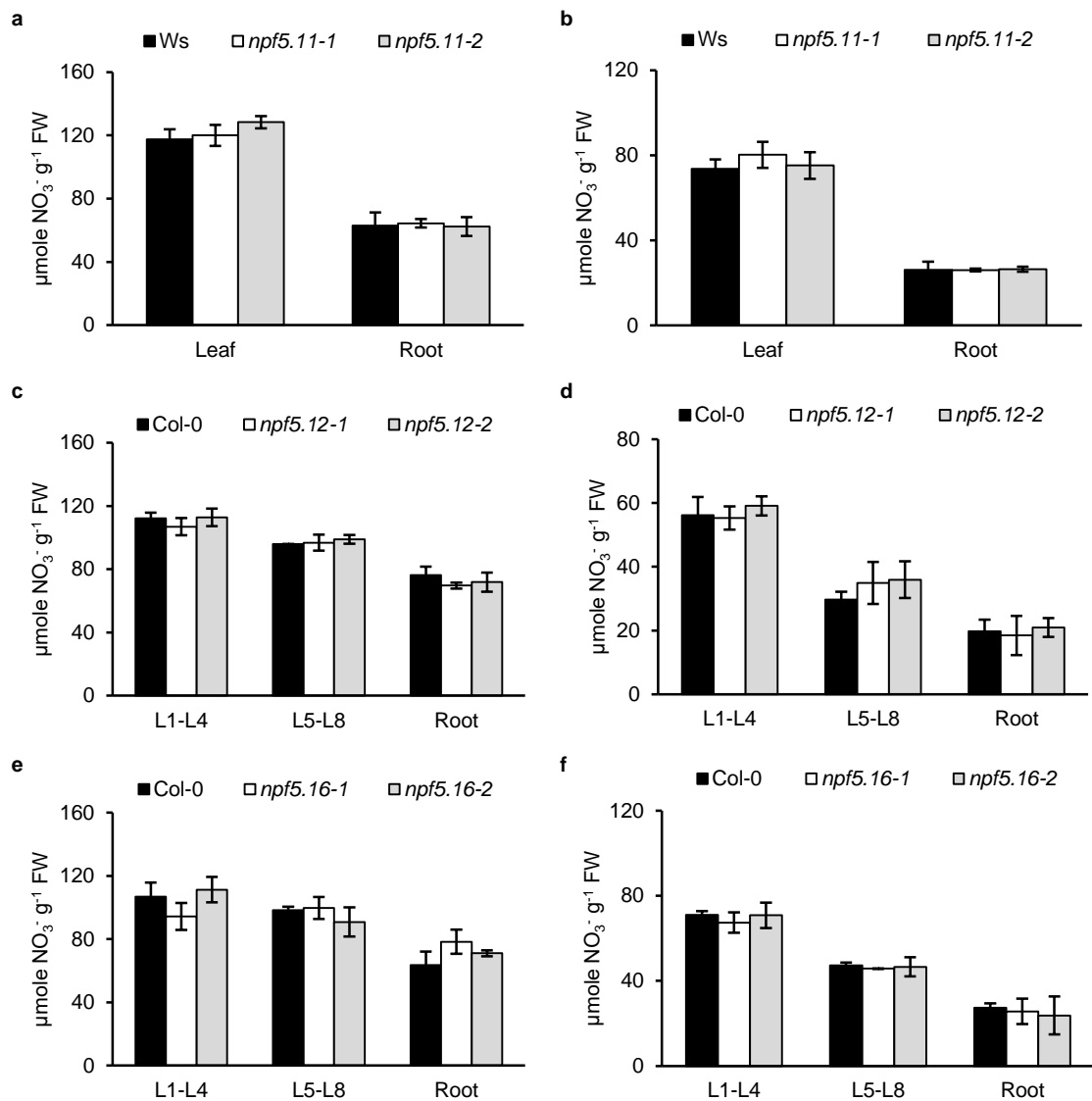
Supplementary Figure S3. NPF5.11, NPF5.12 and NPF5.16 could not export nitrate in oocytes.

50 nL of K¹⁵NO₃ (100 mM) was injected into oocytes previously injected with water, NPF5.11-, NPF5.12-, NPF5.16- or CHL1-cRNA. At 0 h (immediately after injection) or after 3 h of incubation in ND96 buffer at pH 5.5 or 7.4, oocytes were washed three times with ND96 buffer and ¹⁵N was determined. Values are means ± SD (n = 6-10). Asterisks indicate difference at *P* < 0.01 (**) compared with 0 h by Student's *t*-test.



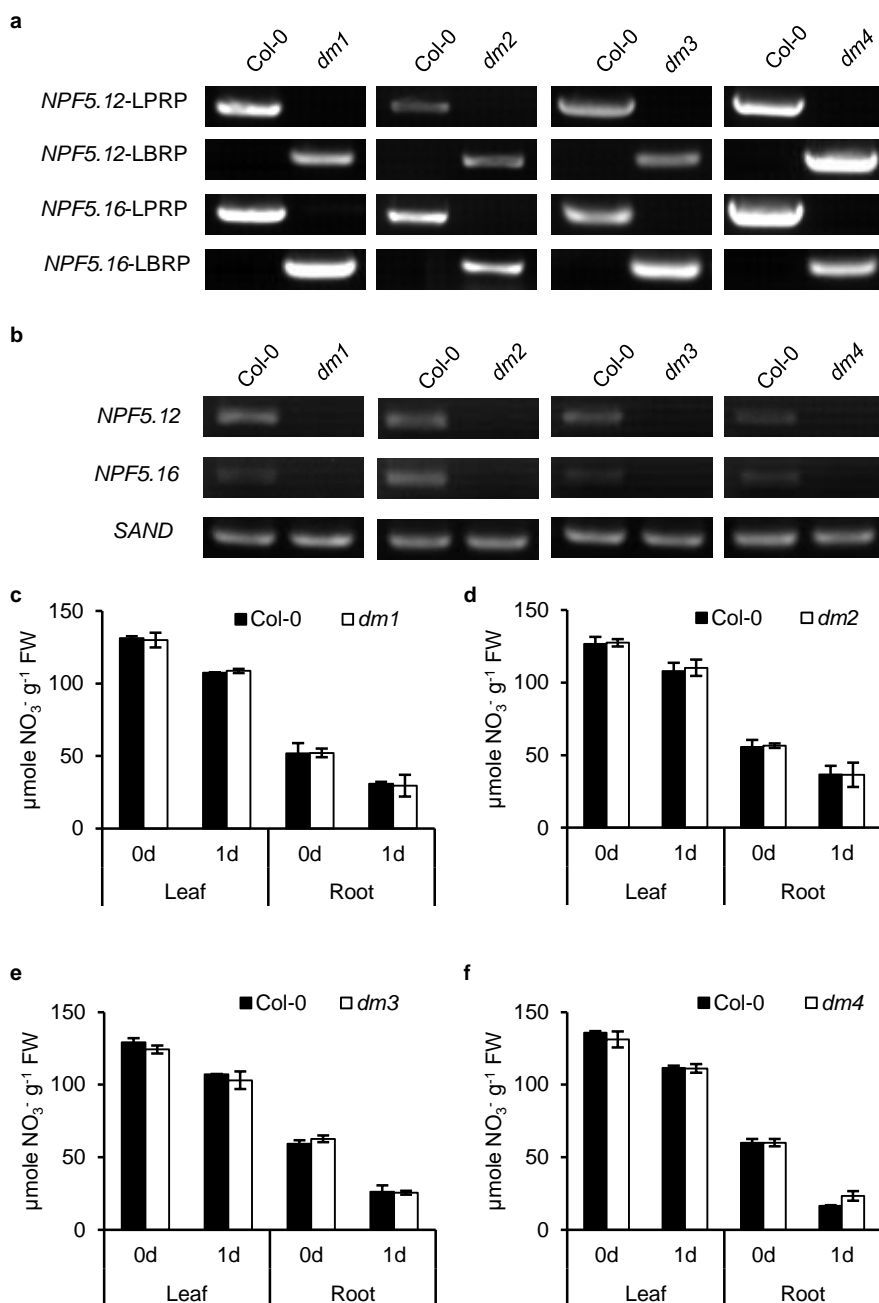
Supplementary Figure S4. The identification of *npf5.11*, *npf5.12* and *npf5.16* knock-out mutants.

(a,c,e) Schematic map of the T-DNA insertion sites in *npf5.11* (a), *npf5.12* (c) or *npf5.16* (e). Black boxes, coding regions; white boxes, untranslated regions (UTRs); Arrows, primers used for RT-PCR. (b,d,f) Expression of *NPF5.11* (b), *NPF5.12* (d) and *NPF5.16* (f) in the corresponding mutant lines, respectively. RNA isolated from roots (d) or leaves (b,f) of 28-d-old plants grown in hydroponics was used for RT-PCR analysis. *ACTIN* or *SAND* (loading control) was amplified for 28 cycles, *NPF5.11* (b) was amplified for 32 cycles, *NPF5.12* (d) and *NPF5.16* (f) were amplified for 35 cycles.



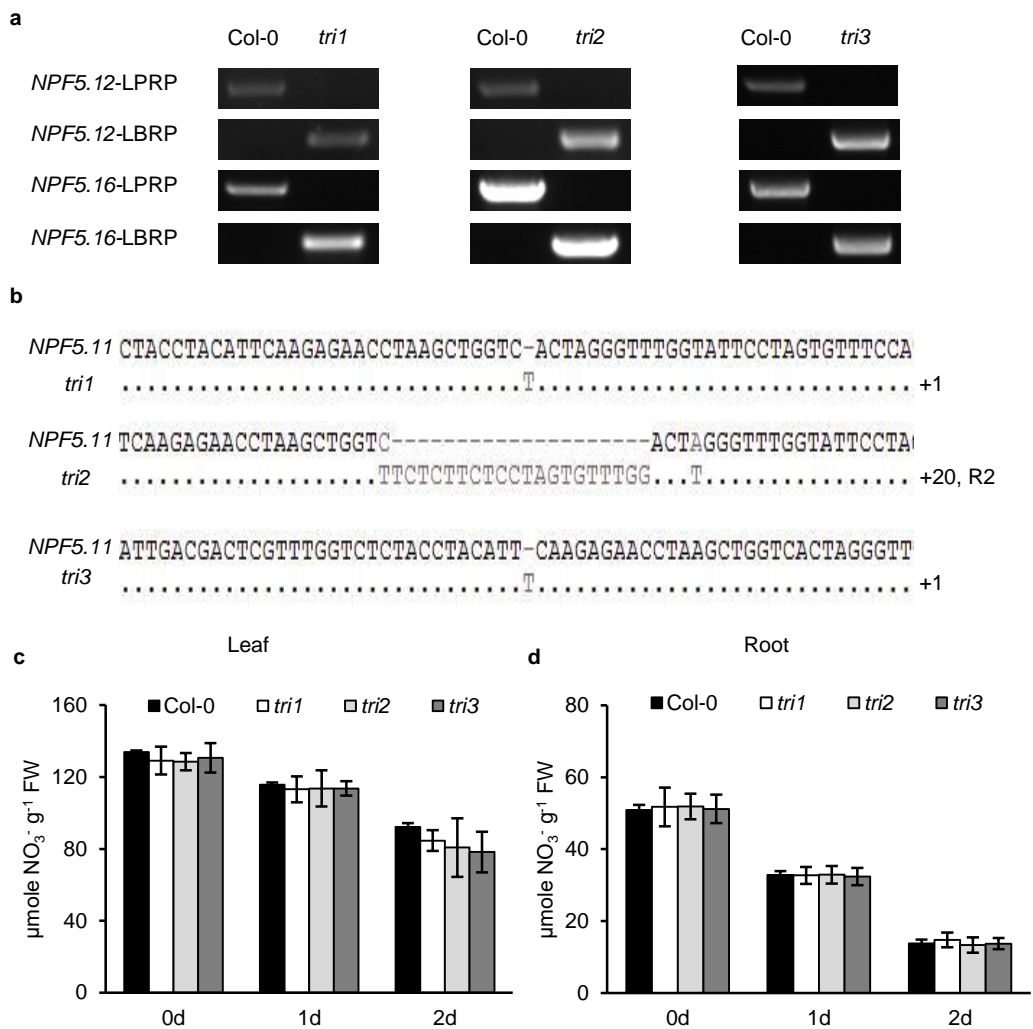
Supplementary Figure S5. Mutation of *NPF5.11*, *NPF5.12* or *NPF5.16* do not affect nitrate accumulation under both control and nitrogen-starved conditions.

(a,b) Plants were cultivated in hydroponics for 24 d (a) and then were treated with nitrogen-starved nutrient solution for 1 d (b), the leaves and roots of Ws and *npf5.11* were harvested to analyze nitrate concentration. Values are means \pm SD, n = 3. (c,d) Plants were cultivated in hydroponics for 28 d (c) and then were treated with nitrogen-starved nutrient solution for 1 d (d), old leaves (L1-L4), young leaves (L5-L8) and roots of Col-0 and *npf5.12* were harvested to analyze nitrate concentration. Values are means \pm SD, n = 3. (e,f) Plants were cultivated in hydroponics for 28 d (e) and then were treated with nitrogen-starved nutrient solution for 1 d (f), old leaves (L1-L4), young leaves (L5-L8) and roots of Col-0 and *npf5.16* were harvested to analyze nitrate concentration. Values are means \pm SD, n = 3.



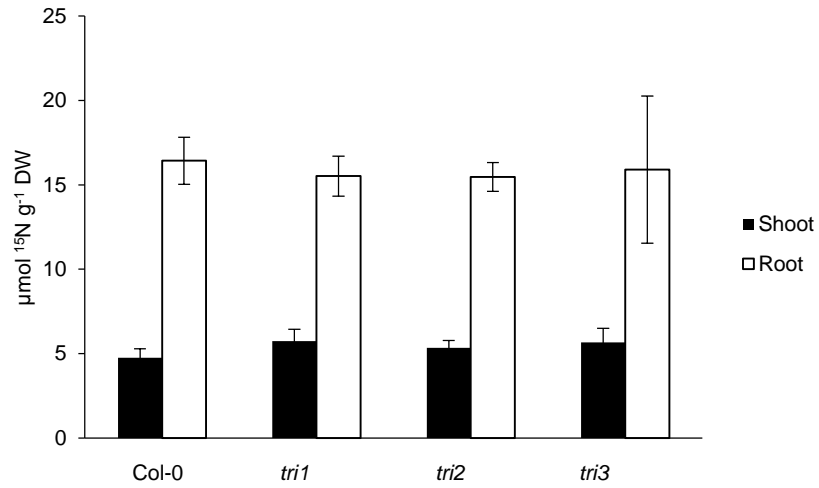
Supplementary Figure S6. Nitrate contents is not affected in *npf5.12 npf5.16* double mutant lines.

(a,b) Analysis of T-DNA insertion (a) and *NPF5.12*, *NPF5.16* expression in seedlings of *npf5.12 npf5.16* double mutant lines (b). *dm1*, *npf5.12-1 npf5.16-1*; *dm2*, *npf5.12-1 npf5.16-2*; *dm3*, *npf5.12-2 npf5.16-1*; *dm4*, *npf5.12-2 npf5.16-2*. *SAND* (loading control) was amplified for 28 cycles, *NPF5.12* and *NPF5.16* was amplified for 32 cycles in (b). (c-f) Nitrate concentration in *dm1* (c), *dm2* (d), *dm3* (e), *dm4* (f). 23-day-old plants were subjected to nitrogen-starved nutrient solution for indicated time. The leaves and roots of wild type and double mutant lines were harvested to analyze nitrate concentration. Values are means \pm SD, n = 3.



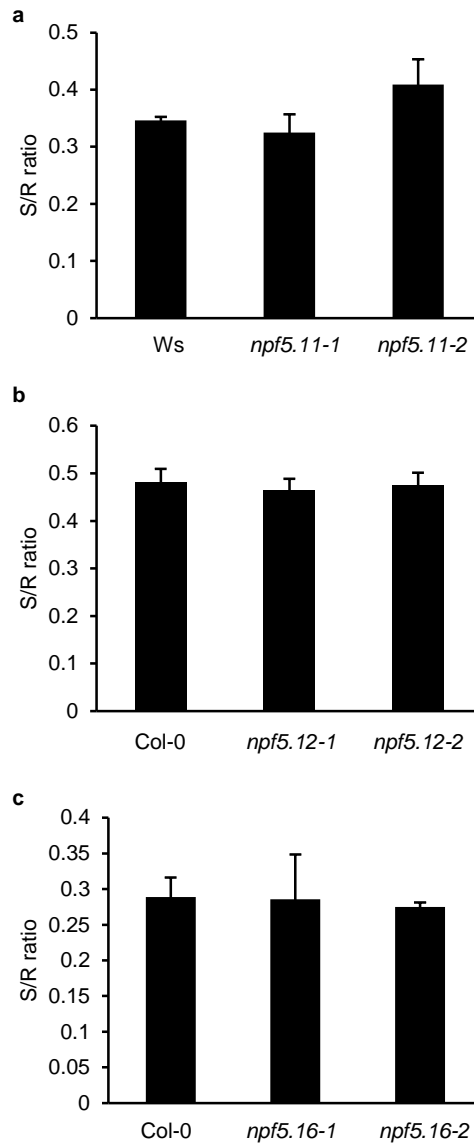
Supplementary Figure S7. Nitrate contents is not affected in *npf5.11 npf5.12 npf5.16* triple mutant lines.

(a) PCR identification of T-DNA insertion in *npf5.11 npf5.12 npf5.16* triple mutant lines. (b) Sequence of edited *NPF5.11* in triple mutant lines. The nucleotide changes were on the right of each sequence (+, insertion; R, replace). (c,d) Nitrate concentration in leaves (c) or roots (d) of triple mutant lines. 28-day-old plants were subjected to nitrogen-starved nutrient solution for indicated time. Values are means \pm SD, n = 3.



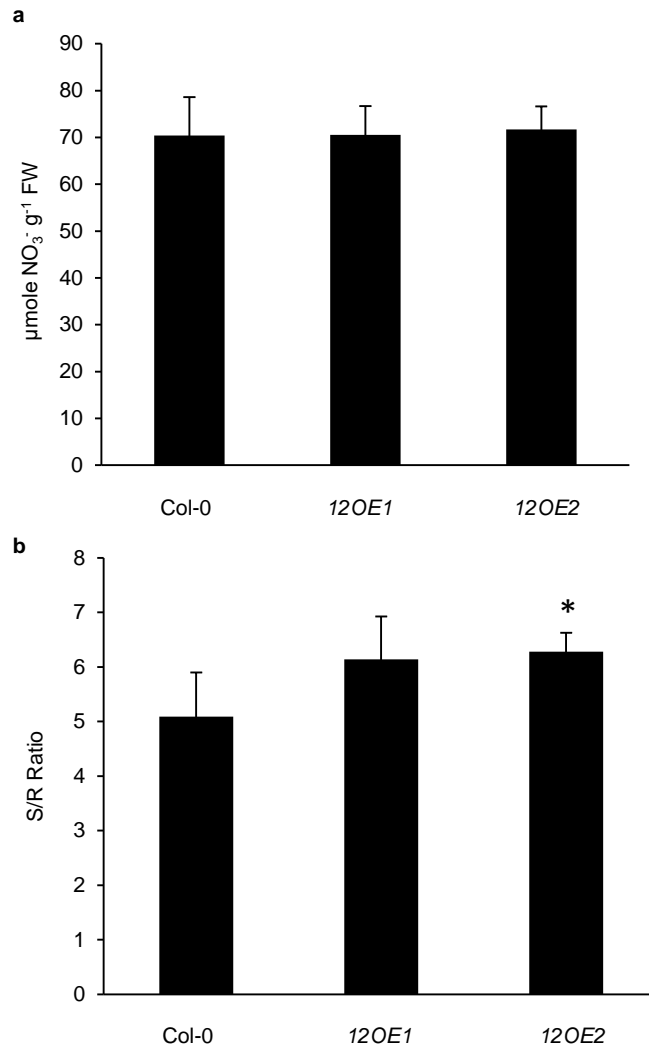
Supplementary Figure S8. $^{15}\text{NO}_3^-$ concentration in shoots and roots of triple mutant plants.

Plants were grown in hydroponics for 28 days and treated with 2.25 mM K^{15}NO_3 for 30 min. ^{15}N concentration in shoots and roots were analyzed. Values are means \pm SD, $n = 3$.



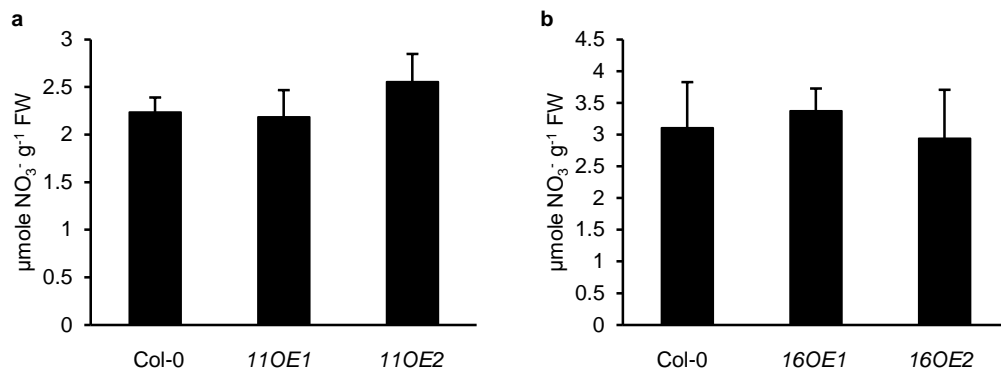
Supplementary Figure S9. Root-to-shoot nitrate transport was not affected in *npf5.11*, *npf5.12*, *npf5.16* single mutants.

28-day-old plants were treated with 2.25 mM $K^{15}NO_3$ for 30 min, ^{15}N contents in shoots and roots were detected and the ^{15}N concentration ratio in shoot and root (S/R ratio) of *npf5.11* (a), *npf5.12* (b), *npf5.16* (c) were calculated. Values are means \pm SD, n = 3.



Supplementary Figure S10. Nitrate accumulation in shoots and the S/R ratio of *NPF5.12* overexpression lines.

24 days old plants grown hydroponically were subjected to nitrogen-starvation for 30 h, then nitrate contents in shoots were determined by HPLC (**a**) and the S/R ratios were calculated (**b**). *12OE1* and *12OE2* were two independent *NPF5.12* overexpression lines. Values are means \pm SD, n = 5-7. Asterisks indicate difference between wild type and overexpression lines at $P < 0.05$ (*) by Student's t-test.



Supplementary Figure S11. Overexpression of *NPF5.11* or *NPF5.16* do not affect nitrate contents in roots under nitrogen-starved condition.

28 days old hydroponically grown plants were subjected to nitrogen-starvation for 2 d, then roots of 11OE1 and 11OE2 (a), 16OE1 and 16OE2 (b) were sampled and nitrate contents were measured by HPLC. 11OE1 and 11OE2 were two independent *NPF5.11* overexpression lines. 16OE1 and 16OE2 were two independent *NPF5.16* overexpression lines. Values are means \pm SD, n = 3.

Supplemental Table S1. List of primer sequences

Destination	Forward primer	Reverse primer
Genotyping		
<i>npf5.11-1</i>	CTGAGGGTTTTCTGGAAATCC	CTGCCCTGTCTAGGAATCTGC
<i>npf5.11-2</i>	GTATGAATGTACGGCCAGTGC	TCCCTGTTTGTTTTGTTTTCG
<i>npf5.12-1</i>	CAAACGTTAACCTCTGGCTTG	TTTTGTTTGTAACAACAAAACACAGTG
<i>npf5.12-2</i>	CAAACGTTAACCTCTGGCTTG	TTTTCATCTCGACCAAAGCAG
<i>npf5.16-1</i>	TCGTGTTTCATTATTCACGCAG	GAAAACTCTACCGATCCTCGC
<i>npf5.16-2</i>	CGCCTTGTTTTGTGAAGAAAG	TCGTGTTTCATTATTCACGCAG
LB4	CGTGTGCCAGGTGCCACGGAATAGT	
GABI-LB	ATATTGACCATCATACTCATTGC	
LB3	TAGCATCTGAATTTCATAACCAATCTCGATACAC	
LBb1.3	ATTTTGCCGATTTTCGGAAC	
CRISPR-Cas9 system		
Target sequence-1	GAGAACCTAAGCTGGTCACT	
Target sequence-2	GCTTAGGTTCTCTTGAATGT	
<i>Cas9</i>	AGCAGCCGACAAGAAGTACAGC	GCTTTCAGCAGGGTCAGGTCCT
Transient expression & Plant transformation		
<i>NPF5.11</i>	ctcgagATGGCTATCACCTACTCCTC	actagtAAAGGTGTTTGATCTGCTGT
<i>NPF5.12</i>	ggatccATGTCGACATCCATCGGCGATA	actagtCTTTGGGCTGTTGTAGAGAT
<i>NPF5.16</i>	gtcgacATGGCGATAGCCGAAGA	ggatccGACTTGATCTACACGGC
<i>NPF5.11pro</i>	gtcgacTACAGCTCTTGTGTCAGGCTTA	ggatccTAATGTGTATGATTGATTGG
<i>NPF5.12pro</i>	gtcgacCTAACGCGACAGCAAGCACT	ggatccTGCTTCTTGTTATTTGTTT
<i>NPF5.16pro</i>	gtcgacCATTGCGCGGTGGGCAAT	cccgggTTTCTTGTGCGAGGAAAC
RT-PCR & qRT-PCR		
<i>npf5.11-1</i>	CGAATCAGCTGCAAGAGACTATCT	AAGAGTCCTGGTGAGATTGACCTG
<i>npf5.11-2</i>	ATGGCTATCACCTACTCCTCCGC	GATGATGAGCCTCGCTGATTTCC
<i>npf5.12-1/npf5.12-2</i>	AATCTGATAACCTACTTCACCGAGGC	CGGTAAGTTTTGATTCCAAGTAAGAACAAG
<i>npf5.16-1/npf5.16-2</i>	GATAGCCGAAGAAGAAGCTGCA	CCAAAAGCTTGAACACATGGCT
<i>Actin2</i>	CCCTGTTCTTCTTACCGAG	CCACATCTGCTGGAATG
<i>SAND</i>	GATGAGGATGATGCTTCTACG	CCTGAGCGTTGTATCTTGGT
<i>qNPF5.11</i>	GCGTTGGTATGGCTTTGAAC	GCCAGTAGCCAGTAGAAGTAATC
<i>qNPF5.12</i>	CATGGTATCAGTCATCGAGGAAG	GACAAGCAAGTAGCCAGTAGAA
<i>qNPF5.16</i>	CCACGGAGCTAAGGAGTATTG	GGGCTCTGTTGAGTTAGTATT
<i>qSAND</i>	ATATGACACCCTTGCTTGGAGGGA	TGAGAATAAGACACCAGACGCGCA