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# Main Figures: 5

# Supplementary Figures: 8

# Supplementary Tables: 3

# Supplementary Videos: 1

## Reporting Checklist for Nature Neuroscience

This checklist is used to ensure good reporting standards and to improve the reproducibility of published results. For more information, please read [Reporting Life Sciences Research](#).

Please note that in the event of publication, it is mandatory that authors include all relevant methodological and statistical information in the manuscript.

### ► Statistics reporting, by figure

- Please specify the following information for each panel reporting quantitative data, and where each item is reported (section, e.g. Results, & paragraph number).
- Each figure legend should ideally contain an exact sample size (n) for each experimental group/condition, where n is an exact number and not a range, a clear definition of how n is defined (for example x cells from x slices from x animals from x litters, collected over x days), a description of the statistical test used, the results of the tests, any descriptive statistics and clearly defined error bars if applicable.
- For any experiments using custom statistics, please indicate the test used and stats obtained for each experiment.
- Each figure legend should include a statement of how many times the experiment shown was replicated in the lab; the details of sample collection should be sufficiently clear so that the replicability of the experiment is obvious to the reader.
- For experiments reported in the text but not in the figures, please use the paragraph number instead of the figure number.

**Note:** Mean and standard deviation are not appropriate on small samples, and plotting independent data points is usually more informative. When technical replicates are reported, error and significance measures reflect the experimental variability and not the variability of the biological process; it is misleading not to state this clearly.

FIGURE NUMBER	TEST USED		n			DESCRIPTIVE STATS (AVERAGE, VARIANCE)		P VALUE		DEGREES OF FREEDOM & F/t/z/R/ETC VALUE	
	WHICH TEST?	SECTION & PARAGRAPH #	EXACT VALUE	DEFINED?	SECTION & PARAGRAPH #	REPORTED?	SECTION & PARAGRAPH #	EXACT VALUE	SECTION & PARAGRAPH #	VALUE	SECTION & PARAGRAPH #
example 1a	one-way ANOVA	Fig. legend	9, 9, 10, 15	mice from at least 3 litters/group	Methods para 8	error bars are mean +/- SEM	Fig. legend	p = 0.044	Fig. legend	F(3, 36) = 2.97	Fig. legend
example results, para 6	unpaired t-test	Results para 6	15	slices from 10 mice	Results para 6	error bars are mean +/- SEM	Results para 6	p = 0.0006	Results para 6	t(28) = 2.808	Results para 6

		TEST USED		n			DESCRIPTIVE STATS (AVERAGE, VARIANCE)		P VALUE		DEGREES OF FREEDOM & F/t/z/R/ETC VALUE	
FIGURE NUMBER	WHICH TEST?	SECTION & PARAGRAPH #	EXACT VALUE	DEFINED?	SECTION & PARAGRAPH #	REPORTED?	SECTION & PARAGRAPH #	EXACT VALUE	SECTION & PARAGRAPH #	VALUE	SECTION & PARAGRAPH #	
+ -	Fig. 2D	unpaired t-test	4	yes, 4 mice per group	Line 385 and Lines 821-823	Error bars are mean +/- SEM shown in main 2d and Supplemental 1a-c show raw data plotted as a cumulative a distribution	Supplemental Figure 1 And lines 385-386	p = 0.0361, 0.0443, and 0.0071, cortex, striatum and cerebellum, respectively	Lines 378-389	(t(degrees) cortex = t6 = 2.688 striatum = t6 = 2.536 cerebellum t6 = 4.007	Cortex = line 379 Striatum = line 379 cerebellum = line 380	
+ -	supplemental Figure 1a-c	Kolmogorov-Smirnov test	4	yes, 4 mice per group	Supplemental Document Supplemental Figure 1 Legend. Last line	Individual animals are plotted within the same plot to show variability	Outlined inside supplemental figure, top of figure	P ≤ 0.0001	Reported in Supplemental 1 Figure legends, last line.	cortex = D value = 0.48 striatum = D value = 0.4536 cerebellum = D value = 0.407	NR	
+ -	Fig. 2E DAPI	unpaired t-test	4	yes, 4 mice per group	Line 385 and Lines 821-823	Error bars are mean +/- SEM	lines 385-386	p = 0.0034, 0.0541, cortex and striatum, respectively	Line 380-381	cortex = t6 = 4.669 striatum t6 = 2.390	cortex = line 381 striatum = line 382	
+ -	Fig. 2E Neun/Calb	unpaired t-test	4	yes, 4 mice per group	Line 385 and Lines 821-823	Error bars are mean +/- SEM	Lines 385-386	p = 0.037, 0.128, and 0.039, cortex, striatum and cerebellum, respectively	Lines 381-382	cortex = t6 = 2.662 sttium = t6 = 1.764 cerebellum = t6 = 3.328	cortex = line 383 striatum = 383 cerebellum = 384	
+ -	Fig. 2E s100	unpaired t-test	4	yes, 4 mice per group	Line 385 and Lines 821-823	Error bars are mean +/- SEM	Lines 385-386	p = 0.6738 and 0.1814, cortex and striatum, respectively	Lines 384-385	cortex = t6 = 0.4422 striatum = t6 = 1.512	cortex = line 385 striatum = line 386	
+ -	Fig. 3B	unpaired t-test	3	yes, 3 mice per group	Lines 399-400	Error bars are mean +/- SEM	Lines 399-400	p = 0.0014	Line 396	t4 = 7.814	line 397	
+ -	Fig. 3C	unpaired t-test	3	yes, 3 mice per group	Lines 399-400	Error bars are mean +/- SEM	Lines 399-400	P ≤ 0.0001,	Lines 396-397	t4 = 18.29	line 398	
+ -	Supplemental Figure 6c and d	unpaired t-test	NR	1 animal per dose, 3 brain sections per dose	Line 6 of Supplemental Figure legend 6 and Methods line 803-804	Error bars are mean +/- SEM between the 3 sections	Line 6 of Supplemental Figure legend 6	NR	NR	NR	NR	

## ► Representative figures

1. Are any representative images shown (including Western blots and immunohistochemistry/staining) in the paper?

If so, what figure(s)?

Figure 1c-g  
Figure 2a-c  
Figure 3a,d  
Figure 5b,e  
Supplemental Figure 3a,b  
Supplemental Figure 4a-c

2. For each representative image, is there a clear statement of how many times this experiment was successfully repeated and a discussion of any limitations in repeatability?

If so, where is this reported (section, paragraph #)?

Figure 1c-g - (n = 4 individual animals per group), Line 825-826  
Figure 2a-c (n = 4 individual animals per group), Lines 827-828  
Figure 3a (n = 3 individual animals per group), Lines 827-828 and line 400  
Figure 3d (n = 2 animals sections per section of intestine/colon)  
Lines 400-401  
Supplemental Figure 3a,b, NR  
Supplemental Figure 4a-c NR

## ► Statistics and general methods

1. Is there a justification of the sample size?

If so, how was it justified?

Where (section, paragraph #)?

Even if no sample size calculation was performed, authors should report why the sample size is adequate to measure their effect size.

Not done formally, Sample sizes were chosen based on preliminary data and suggested a large effect size. Lines 821-823

2. Are statistical tests justified as appropriate for every figure?

Where (section, paragraph #)?

Each figure legend outlines the statistics test used and n number. In addition, statistic section 820-837

- a. If there is a section summarizing the statistical methods in the methods, is the statistical test for each experiment clearly defined?

Yes, Lines 795-812

- b. Do the data meet the assumptions of the specific statistical test you chose (e.g. normality for a parametric test)?

Where is this described (section, paragraph #)?

parametric for all statistics outlined in figure legends as listed above in the statistical figure reporting. Line 832

- c. Is there any estimate of variance within each group of data?

Is the variance similar between groups that are being statistically compared?

Where is this described (section, paragraph #)?

Data were not tested for normal distribution due to small sample sizes.

- d. Are tests specified as one- or two-sided?

Yes, Fig2 and 3 statistics were all two-sided, specified in the figure legends

- e. Are there adjustments for multiple comparisons?

No multiple comparisons were made

3. To promote transparency, *Nature Neuroscience* has stopped allowing bar graphs to report statistics in the papers it publishes. If you have bar graphs in your paper, please make sure to switch them to dot-plots (with central and dispersion statistics displayed) or to box-and-whisker plots to show data distributions.
4. Are criteria for excluding data points reported?  
Was this criterion established prior to data collection?  
Where is this described (section, paragraph #)?
5. Define the method of randomization used to assign subjects (or samples) to the experimental groups and to collect and process data.  
If no randomization was used, state so.  
Where does this appear (section, paragraph #)?
6. Is a statement of the extent to which investigator knew the group allocation during the experiment and in assessing outcome included?  
If no blinding was done, state so.  
Where (section, paragraph #)?
7. For experiments in live vertebrates, is a statement of compliance with ethical guidelines/regulations included?  
Where (section, paragraph #)?
8. Is the species of the animals used reported?  
Where (section, paragraph #)?
9. Is the strain of the animals (including background strains of KO/transgenic animals used) reported?  
Where (section, paragraph #)?
10. Is the sex of the animals/subjects used reported?  
Where (section, paragraph #)?
11. Is the age of the animals/subjects reported?  
Where (section, paragraph #)?
12. For animals housed in a vivarium, is the light/dark cycle reported?  
Where (section, paragraph #)?
13. For animals housed in a vivarium, is the housing group (i.e. number of animals per cage) reported?  
Where (section, paragraph #)?
- Yes, 798-800
- Mice were randomly assigned to groups of predetermined sample size. All experiments with direct comparisons were performed in parallel to minimize variability. Lines 632-633.  
To minimize bias, when direct comparisons for quantification Lines 762-765
- Manual cell counting in the striatum and cortex was performed by a blinded observer (Fig. 2e). Automated counting was performed of Purkinje cells in the cerebellum (Fig. 2e). Lines 765-767.
- Yes, methods, lines 628-632.
- Yes, methods, lines 629-631
- Yes, methods, lines 629-631
- Yes, methods, line 631-632
- Yes, line 640
- No
- No

14. For behavioral experiments, is the time of day reported (e.g. light or dark cycle)?  
Where (section, paragraph #)?
- No
15. Is the previous history of the animals/subjects (e.g. prior drug administration, surgery, behavioral testing) reported?  
Where (section, paragraph #)?
- No
- a. If multiple behavioral tests were conducted in the same group of animals, is this reported?  
Where (section, paragraph #)?
- No
16. If any animals/subjects were excluded from analysis, is this reported?  
Where (section, paragraph #)?
- 1 animal in each group did not show any virus transduction in figure 2 and were excluded. Line 823-825
- a. How were the criteria for exclusion defined?  
Where is this described (section, paragraph #)?
- complete lack of expression. Line 823-825
- b. Specify reasons for any discrepancy between the number of animals at the beginning and end of the study.  
Where is this described (section, paragraph #)?
- complete lack of expression was observed, possible reason was due to failed retro orbital injection during virus injection. Line 823-825

## ► Reagents

1. Have antibodies been validated for use in the system under study (assay and species)?
- All antibodies used within the study are reported in methods section lines 696-751
- a. Is antibody catalog number given?  
Where does this appear (section, paragraph #)?
- Yes. 696-751
- b. Where were the validation data reported (citation, supplementary information, Antibodypedia)?  
Where does this appear (section, paragraph #)?
- NA
2. Cell line identity
- No
- a. Are any cell lines used in this paper listed in the database of commonly misidentified cell lines maintained by [ICLAC](#) and [NCBI Biosample](#)?  
Where (section, paragraph #)?
- b. If yes, include in the Methods section a scientific justification of their use--indicate here in which section and paragraph the justification can be found.

- c. For each cell line, include in the Methods section a statement that specifies:
- the source of the cell lines
  - have the cell lines been authenticated? If so, by which method?
  - have the cell lines been tested for mycoplasma contamination?

Where (section, paragraph #)?

Virus production , line 682

## ► Data availability

Provide a Data availability statement in the Methods section under "Data availability", which should include, where applicable:

- Accession codes for deposited data
- Other unique identifiers (such as DOIs and hyperlinks for any other datasets)
- At a minimum, a statement confirming that all relevant data are available from the authors
- Formal citations of datasets that are assigned DOIs
- A statement regarding data available in the manuscript as source data
- A statement regarding data available with restrictions

See our [data availability and data citations policy page](#) for more information.

Data deposition in a public repository is mandatory for:

- a. Protein, DNA and RNA sequences
- b. Macromolecular structures
- c. Crystallographic data for small molecules
- d. Microarray data

Deposition is strongly recommended for many other datasets for which structured public repositories exist; more details on our data policy are available [here](#). We encourage the provision of other source data in supplementary information or in unstructured repositories such as [Figshare](#) and [Dryad](#).

We encourage publication of Data Descriptors (see [Scientific Data](#)) to maximize data reuse.

Where is the Data Availability statement provided (section, paragraph #)?

At the stage of publication, novel capsid sequences, will be deposited to Genbank and all viral plasmids will be deposited to addgene.com

## ► Computer code/software

Any custom algorithm/software that is central to the methods must be supplied by the authors in a usable and readable form for readers at the time of publication. However, referees may ask for this information at any time during the review process.

1. Identify all custom software or scripts that were required to conduct the study and where in the procedures each was used.

custom Matlab and Python scripts were used to assist in data analysis

2. If computer code was used to generate results that are central to the paper's conclusions, include a statement in the Methods section under "**Code availability**" to indicate whether and how the code can be accessed. Include version information as necessary and any restrictions on availability.

Code will be deposited to github near publication.

## ▶ Human subjects

1. Which IRB approved the protocol?

Where is this stated (section, paragraph #)?

NA

2. Is demographic information on all subjects provided?

Where (section, paragraph #)?

3. Is the number of human subjects, their age and sex clearly defined?

Where (section, paragraph #)?

4. Are the inclusion and exclusion criteria (if any) clearly specified?

Where (section, paragraph #)?

5. How well were the groups matched?

Where is this information described (section, paragraph #)?

6. Is a statement included confirming that informed consent was obtained from all subjects?

Where (section, paragraph #)?

7. For publication of patient photos, is a statement included confirming that consent to publish was obtained?

Where (section, paragraph #)?

## ▶ Additional comments

Additional Comments