

S3 Table. List of primers used in this study.

Mapping primers			
Primer name	Forward sequence (5'-3')	Reverse sequence (5'-3')	
Indel			
6-26	CTCAACGTTGACACCTCGTG	TCCTCCATCGAGCAGTATCA	
Q-7	CATCCCAAACCTACCAACCAA	GAGATGTGACGAGGGAAGAG	
Q-17	GACACGTTTAGCTGCGTAGTT	CCTTCAATGGTCGTATCTTCT	
Q-26	TACTAGATGGGATGAAAATCA	GAATAATGCCTCACCTGTGC	
Y-6	TAGGTGGATCTATTGTATTTA	CAATTCTCTTATACTTCAACA	
Y-13	TCCGATGCTTCAATCAACAGTC	GGTTTGGGAGGATTGGGAAT	
Y-14	AAATAAAAAAAGAAGGGGCT	TTCAATGAGAAGTTGGTGGT	
Y-80	GCTGCTACTACCTCTGCCTCTTCC	TCAATCGATCCATCCATCTCTCC	
dCAPS			Restriction enzyme
Q-20	GTCAGTAGCAGATC	AACGTTGGTGCATAGCGGAC	BglII
Primers for confirming the fragment inversion			
Primer name	Primer sequence (5' to 3')		
1F	ACAAGAATCCCGTAGACAT		
1R	CATTAGCACGTTGGACATA		
2F	CATACAATGACCCACTCCTAAC		
2R	CTTCATCATGCTCCTCTTCC		
3F	TCTTAGCAGCGAGATGCAGTTCC		
3R	TGTGCCGCACGCAATGGT		
4F	AGATTGTATTCAGAAAGGCAGGTC		
4R	TGGGTGTAGATGTACGAGGAGC		
Quantitative PCR Primers			
Primer name	Primer sequence (5' to 3')		
5F	AGTAGTGGGAACTGGAGATTTGG		
5R	CATCACCTGAGGGAGTGGAAAG		
6F	TGCTAACACAGAACCTGGACTT		
6R	GGACCCTATGAGTAATGCGAAAA		
7F	TGGGCATGTCGTTGGTTCA		
7R	ACAGGCTTGGGTGGTTGG		
Primer name	Forward sequence (5'-3')	Reverse sequence (5'-3')	
qRT-ORF2	AGACAGAGACAGGGAGATGTTGC	AGCAGCAGAGGATGAATGGTTCCG	
qRT-ORF3	CGCCGAGAACGACAAGAT	CGACACGCGGTCCAGC	
OsMPK20-4	GATTCCAGGTAGAACAGGAC	GCTCTCTTTCTGAGCTATCC	
OsAOS2	CAATACGTGTACTGGTCGAATGG	AAGGTGTCGTACCGGAGGAA	
OsVPE3	CGGTAACACAGGCACCAGGC	GTGACTTCGTCTCCAGTGAATCC	
Ubq	GCTCCGTGGCGGTATCAT	CGGCAGTTGACAGCCCTAG	
3' RACE gene-specific primers			
Primer name	Primer sequence (5' to 3')		
RACE-1	TAGTGGGAACTGGAGATTTGGTTGATGAG		

RACE-2	GGGCATGTCGTTGGTTCATGTTGGTA	
RACE-3	TCAGATACTTCCCAACCACCCAAGCC	
Binary vector construction primers		
Primer name	Primer sequence (5' to 3')	Restriction enzyme
pORF1-1305-EcoRI-F	ATT GAATTC CATCCATTTTAGGGTCCAC	EcoRI
pORF1-1305-KpnI-R	ATT GGTACC TTCTTACACGGAACCCAAC	KpnI
ORF1-1305-BamHI-F	AGGTACCCGG GGATCC ATGTCTGGCCAAGAGAGAGA	BamHI
ORF1-1305-XbaI-R	GCAGGTGCGACT TAGACT ATTGCAACCTTTGCATCT	XbaI
ORF1-1305-GFP-SpeI-F	ATT ACTAGT ATGTCTGGCCAAGAGAGAG	SpeI
ORF1-1305-GFP-BamHI-R	AAT GGATCC TTGCAACCTTTGCATC	BamHI
gORF3-1305-EcoRI-F	CCATGATTAC GAATTC TTCTTACCCATCATAAACCAT	EcoRI
gORF3-1305-EcoRI-R	AAAATGGATG GAATTC TACCCGAACCATTCATCCT	EcoRI
ORF3-1305-GFP-SpeI-F	ATT ACTAGT ATGGATTTGCGCGACGAACC	SpeI
ORF3-1305-GFP-BamHI-R	AAT GGATCC GGGGTGGTGGCCATGGGCGG	BamHI
OsCNGC13-D-1305-BamHI-F	AGGTACCCGG GGATCC ATGTCTGGCCAAGAGAGAGA	BamHI
OsCNGC13-D-1305-XbaI-R	GCAGGTGCGACT TAGATT ATGTCCAACGTGCTAATG	XbaI
20bp-pOs-Cas9-ORF1F ¹	AGATGATCCGTGGCAGACTCAATCATATCCACAGGT TTTAGAGCTATGC	BsaI
20bp-pOs-Cas9-ORF1R	GCATAGCTCTAAAACCTGTGGGATATGATTGAGCTG CCACGGATCATCT	BsaI
20bp-pOs-Cas9-ORF3F	AGATGATCCGTGGCACCCGCTGGTTCTCGTCCGGGG TTTTAGAGCTATGC	BsaI
20bp-pOs-Cas9-ORF3R	GCATAGCTCTAAAACCCCGGACGAGAACCAGCGGGT GCCACGGATCATCT	BsaI
ORF1RNAi-B-K	AAT GGATCC GGTACC TTGAACAACCTCAGACTTAAC	BamHI, KpnI
ORF1RNAi-M-S	AAT ACGCGT GAGCTC AGCCGGACCTTCTTAAGGAA	MluI, SacI
pOsCNGC13-1305-GUS-EcoRI-F	CCATGATTAC GAATTC CATCCATTTTAGGGTCCAC	EcoRI
pOsCNGC13-1305-GUS-NcoI-R	CTCAGATCT ACCATGG TTCTTACACGGAACCCAAC	NcoI
In situ hybridization primers		
OsCNGC13-pGEM-T-F	ATGCTAGAGCTACAAAGATT	
OsCNGC13-pGEM-T-R	CTGGTCATAAGGCCAGTTAC	
Subcellular localization in rice protoplasts primer		
OsCNGC13-pA7-GFP-XhoI-F	ACGAACGATA CTCGAG ATGTCTGGCCAAGAGAGAGA	XhoI
OsCNGC13-pA7-GFP-SpeI-R	GCTCACCAT CTAG TTGCAACCTTTGCATCTCAG	SpeI
Yeast two-hybrid vector construction primers		
OsCNGC13-pDHB1-NcoI-F	GGCCAGGCCT CCATGG ATGTCTGGCCAAGAGAGAGA	NcoI
OsCNGC13-pDHB1-NcoI-R	CCAAGATATA CCATGG TTGCAACCTTTGCATCTCAG	NcoI
OsCNGC13-D-pDHB1-NcoI-F	GGCCAGGCCT CCATGG ATGTCTGGCCAAGAGAGAGA	NcoI

OsCNGC13-D-pDHB1-NcoI-R	CCAAGATAT CCATGGT GTCCAACGTGCTAATGACA	NcoI
OsCNGC13-pPR3-N-BamHI-F	AGATTACGCT GGATCC ATGTCTGGCCAAGAGAGAGA	BamHI
OsCNGC13-pPR3-N-BamHI-R	ACCACTGCTT GGATCC TTGCAACCTTTGCATCTCAG	BamHI
OsCNGC13-D-pPR3-N-BamHI-F	AGATTACGCT GGATCC ATGTCTGGCCAAGAGAGAGA	BamHI
OsCNGC13-D-pPR3-N-BamHI-R	ACCACTGCTT GGATCC GTCCAACGTGCTAATGACA	BamHI
BiFC assay primers		
OsCNGC13-p2Y-N- PacI-F	CATTTACGAACGATAG TTAATTA AATGTCTGGCCAAGA GAGAGA	PacI
OsCNGC13-p2Y-N- SpeI-R	CACTGCCACCTCCTCC ACTAGT TTGCAACCTTTGCAT CTCAG	SpeI
OsCNGC13-D-p2Y-C- PacI-F	CATTTACGAACGATAG TTAATTA AATGTCTGGCCAAGA GAGAGA	PacI
OsCNGC13-D-p2Y-C- SpeI-R	CACTGCCACCTCCTCC ACTAGT TTGTCCAACGTGCTAA TGACA	SpeI
Primers used in patch-clamping recordings from HEK293 cells		
OsCNGC13-pCl-neo-EcoRI-F	CTAGCCTCGAG GAATTC ATGTCTGGCCAAGAGAGAGA	EcoRI
OsCNGC13-pCl-neo-XbaI-R	CCGGGTCGACT CTAGACT ATTGCAACCTTTGCATCT	XbaI
OsCNGC13-D-pCl-neo-EcoRI-F	CTAGCCTCGAG GAATTC ATGTCTGGCCAAGAGAGAGA	EcoRI
OsCNGC13-D-pCl-neo-XbaI-R	CCGGGTCGACT CTAGATT ATGTCCAACGTGCTAATG	XbaI

¹ The 20-bp gene-specific sequences are underlined in 20bp-pOs-Cas9-ORF1F, 20bp-pOs-Cas9-ORF1R, 20bp-pOs-Cas9-ORF3F and 20bp-pOs-Cas9-ORF3R, respectively.