

# Evaluating Genotoxicity of E-Cigarettes with an Automated 3-D Printed Array

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## SUPPORTING INFORMATION

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## Chemicals & Reagents

**Safety note:** Benzo[a]pyrene (B[a]P), 4-[methyl(nitroso)amino]-1-(3-pyridinyl)-1-butanone (NNK) and N'-Nitroso-2-(3-pyridyl)pyrrolidine (NNN) and their metabolites are potential carcinogens. Handling of these chemicals was done taking protective measures including wearing gloves, safety glasses and working in a hood.

B[a]P (MW 252.31), NNK (MW 207.23), NNN (MW 177.20), poly(diallyldimethylammonium chloride) (PDDA, avg. MW=100,000-200,000), poly(acrylic acid) (PAA, avg. MW= 1800), calf thymus DNA (Type I), and other chemicals were from Sigma Aldrich. Pooled male human liver microsomes were from BD Gentest.  $[\text{Ru}(\text{bpy})_2(\text{PVP})_{10}]^{2+}$   $\{\text{Ru}^{\text{II}}\text{PVP}; (\text{bpy}=2,2\text{-bipyridyl}; \text{PVP}=\text{poly}(4\text{-vinylpyridine)})\}$  was synthesized and characterized as described previously.<sup>[1]</sup> Pyrolytic graphite (PG) sheets are from Panasonic PGS-P13689-ND 70  $\mu\text{m}$  thick.

## 3D printing specifications.

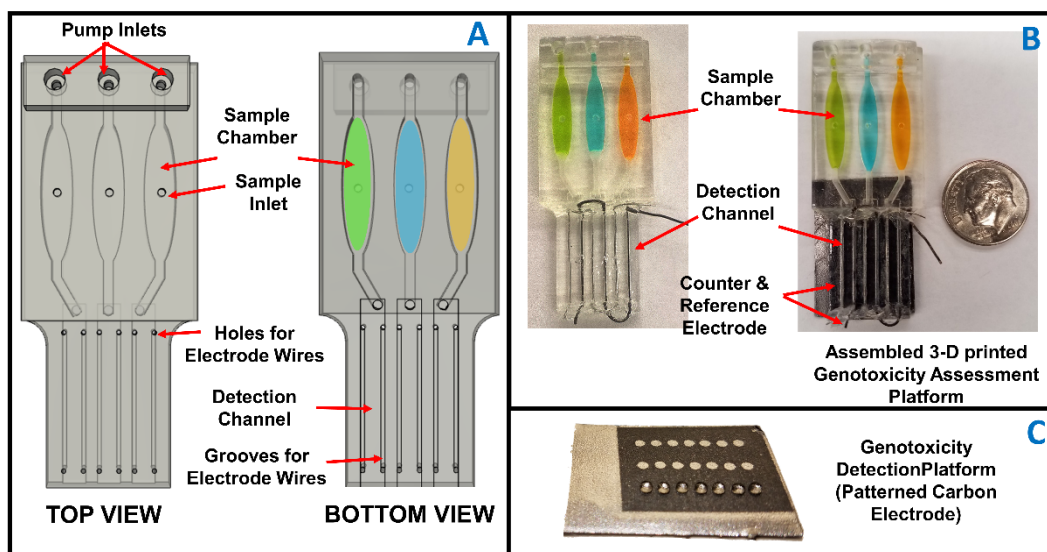
Resin: Formlabs Clear Photopolymer resin FLGPCL02

Printer: Form1+

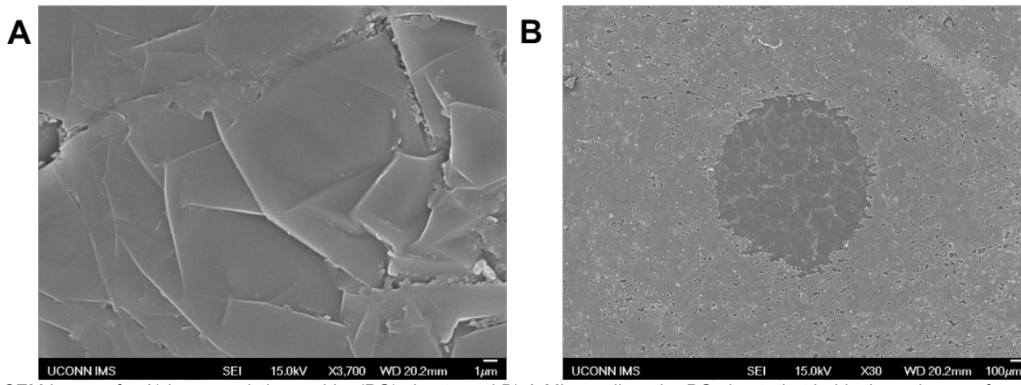
Resolution: 0.05 mm

Steps

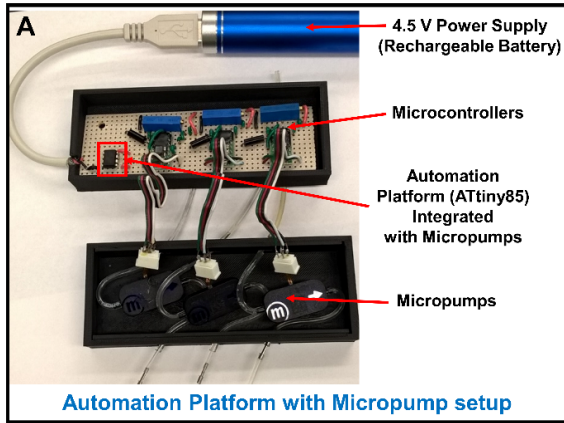
1. Convert 123 design file to printer preform slicer program file format (.preform).
2. Apply orientation and supports to the object.
3. Support specifications: Density 1.5 point size 1.0 mm, no internal supports.
4. Upload the design file to printer and start the print.
5. Post printing
  - a. Remove the printed object from the platform and cut the supports to free the object.
  - b. Flush the channels and the sample chambers with isopropanol and water three times.
  - c. Sanding performed on the devices where the supports were initially present to smooth the surface.
  - d. Dry and spray coat with Krylon colormaster acrylic crystal clear coat and allow to dry for several hours.



**Figure S1.** 3-D Printed genotoxicity array: (A) CAD design showing top and bottom view of 3-D printed arrays with pump inlets, sample chambers, detection channels and grooves for counter and reference electrodes; (B) Printed, assembled devices showing sample chamber containing dye solutions and electrodes wires inserted; (C) microwell patterned PG detection sheet showing droplet surrounded by hydrophobic boundary.



**Figure S2.** SEM images for A) bare pyrolytic graphite (PG) sheets and B) A Microwell on the PG sheet that holds tiny volumes of reagent required to complete layer-by-layer film assembly of enzymes, DNA and RuPVP.



**Figure S3.** Assembled automated array: (A) Automated platform for micropumps showing ATtiny 85 integrated with 4.5 V battery and microcontrollers connected to micropumps that feed the array for the genotoxicity screening assay.

**A**

Pump\_ON\_OFF\_135\_sec\_600\_sec | Arduino 1.6.9

```

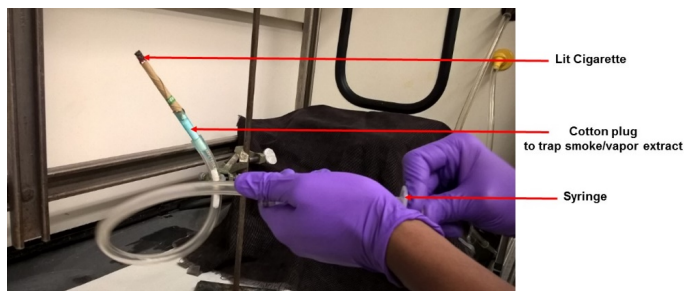
1 // the setup function runs once when you press reset or power the board
2 void setup() {
3   // initialize digital pin 3 as an output.
4   pinMode(0, OUTPUT);
5 }
6
7 // the loop function runs over and over again forever
8 void loop() {
9   digitalWrite(0, HIGH); // turn the micropumps on (HIGH is the voltage level)
10  delay(135000); // wait for a 135 second
11  digitalWrite(0, LOW); // turn the micropumps off by making the voltage LOW
12  delay(600000); // wait for a 600 second
13 }
14

```

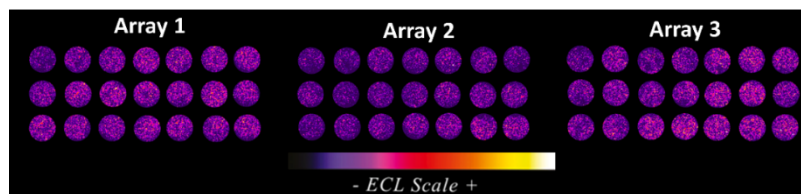
**B**

A schematic diagram showing an Arduino Uno board connected to a breadboard. The ATtiny85 chip is placed on the breadboard, and its pins are connected to the Arduino board's pins. The connections are color-coded to match the code in part A.

**Figure S4.** A) Arduino program for pumps automation B) Schematic upload of Arduino program to ATtiny85 chip.



**Figure S5.** Artificial inhalation setup to extract smoke/vapor from cigarettes on to a cotton plug.



**Figure S6.** Recolorized ECL images from PDDA/PAA/(Ru/DNA)<sub>2</sub>/Ru/Enzyme/DNA films in microwells captured by CCD camera in 10mM phosphate buffer, pH 7.4 upon application of 1.25 V against Ag/AgCl reference electrode for 180 s.

In order to assess the reproducibility of ECL generated between spot to spot and array to array, ECL captured from PDDA/PAA/(Ru/DNA)<sub>2</sub>/Ru/Enzyme/DNA films in 3 different arrays were analyzed upon treatment with 10 mM phosphate buffer and 45s electrolysis followed by 180 s ECL capture. Spot to spot variability of ~ 6 % (n=21 spots) and array to array variability ~7% (n=3) was observed, Figure 3. ECL obtained from 3 different arrays were analyzed by one way analysis of variance, ANOVA and they did not differ statistically at 95 % confidence interval. (p > 0.05)

**Table S1.** Genotoxic reactivity of cigarette sample assessed in terms of known carcinogen concentration.

Sample	% ECL	NNK		NNN		B[a]P	
		[Conc.]	STDEV	[Conc.]	STDEV	[Conc.]	STDEV
1 Tob. Cig	23.32	45.74	2.58	26.40	1.49	45.54	2.56
3 Tob. Cig	33.58	85.52	8.50	49.74	4.94	77.84	7.74
5 Tob. Cig	40.46	117.76	3.36	68.77	1.96	102.37	2.92
1 nf- Tob. Cig	24.35	49.25	3.43	28.45	1.98	48.52	3.38
3 nf- Tob. Cig	39.58	113.41	10.10	66.20	5.90	99.12	8.83
5 nf- Tob. Cig	72.57	320.99	23.45	189.80	13.86	241.63	17.65
20 puff e-cig	33.17	83.74	3.26	48.70	1.89	76.45	2.97
60 puffs e- cig	58.11	219.19	21.79	128.99	12.82	174.28	17.33
100 puffs e-cig	84.24	414.64	35.43	245.96	21.01	300.87	25.71
20 puffs nn e-cig	27.48	60.61	4.04	35.11	2.34	57.96	3.87
60 puffs nn e-cig	39.47	112.84	3.17	65.86	1.85	98.70	2.78
100 puffs nn e-cig	42.71	129.25	6.47	75.56	3.78	110.86	5.55
1 e-cig cartridge	98.53	542.60	44.94	322.95	26.75	378.81	31.37
3 e-cig cartridges	166.15	1330.50	169.94	800.78	102.28	816.67	104.31
5 e-cig cartridges	218.43	2127.83	209.67	1288.16	126.93	1220.95	120.31

Abbrev.: Tob.=tobacco; nf=non-filtered; nn= non-nicotine

## Reference

[1] L. Dennany, R. J. Forster, J. F. Rusling, *J. Am. Chem. Soc.*, **2003**, 125, 5213-5218.