

1 **Absence of deformed wing virus and *Varroa destructor* in Australia**
2 **provides unique perspectives on honeybee viral landscapes and colony**
3 **losses**

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25 **Supplementary material 1. Primers used for RT-PCR virus detection. T_a is the**
 26 **PCR annealing temperature.**

Virus	Primers	T_a	Reference
IAPV	F: AGACACCAATCACGGACCTCAC	58 °C	Maori et al 2007 ¹
	R: AGATTTGTCTGTCTCCCAGTGACAT		
KBV	F: GATGAACGTCGACTATTGA	58 °C	Stoltz et al 1995 ²
	R: TGTGGGTTGGCTATGAGTCA		
BQCV	F: TGG TCA GCT CCC ACT ACC TAA AAC	58 °C	Benjeddou et al 2001 ³
	R: GCA ACA AGA AAC GTA AAC CAC		
LSV1	F: TTA TCT CGC GCC GCC ACC TC	52 °C	Runckel et al 2011 ⁴
	R: ATC GCC GCT GCA ACG TGA CC		
LSV2	F: CGG CCG GTC TAG CGT GGT TG	52 °C	Runckel et al 2011 ⁴
	R: TGG CAA GCT GTG ACG AAT CCC T		
DWV-A/ DWV-B	F: ATA TTC ACG GAT TGT TTG AAA GA	58 °C	Meeus et al 2010 ⁵
	R: CRC TAA CAT TCA TGA TAA GAT CGT C		
SBPV	F: GAT TTG CGG AAT CGT AAT ATT GTT TG	58 °C	de Miranda 2010 ⁶
	R: ACC AGT TAG TAC ACT CCT GGT AAC TTC G		
SBV	F: AAT GGT GCG GTG GAC TAT GG	58 °C	Grabensteiner et al 2001 ⁷
	R: TGA TAC AGA GCG GCT CGA CA		
ABPV	F: TGA GAA CAC CTG TAA TGT GG	58 °C	Tentcheva et al 2004 ⁸
	R: ACC AGA GGG TTG ACT GTG TG		
CBPV	F: AGT TGT CAT GGT TAA CAG GAT ACG AG	58 °C	Ribiere et al 2000 ⁹
	R: TCT AAT CTT AGC ACG AAA GCC GAG		

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34 **Supplementary material 2. Summary of BLAST results for sequence fragments from NT, QLD and WA-2 samples that mapped to**
 35 **DWV reference genomes.**

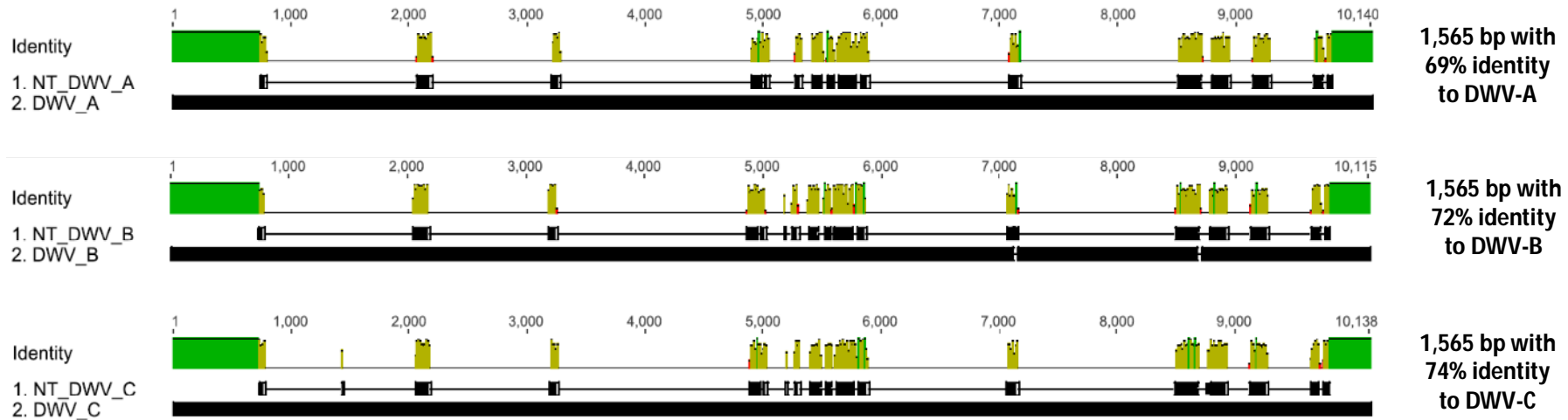
Sequence fragment and length	BLAST aligned sequence	Location on aligned sequence	Highest nucleotide similarity to DWV isolates
NT DWV-like 1 (54bp)	GTAGGGACCCCTCTATCCCTAGGTAAGTATGTCGCGAAAGTATGAAAAGTAAT	734-787	91% similar to DWV isolate leuven-dwv1 (KX783225)
NT DWV-like 2 (127bp)	ATATGCGATGTACCTAATACAATACCTTTCAAGGTGCATGCATATTGGCGTGGAGAAATGCAAGTTCGAATACAAGTTAACTCGAATAAGTCCAGGTTGGTCAATGCAAGCCACTTGGTACTATT	2045-2171	84% similar to DWV isolate leuven-dwv1 (KX783225)
NT DWV-like 3 (66bp)	TGTTTATGTATGTGCAAGTACCATTGATACCGATGGAAGCTGTAGCTGATACTATCGATATTAATG	3189-3254	92% similar to DWV isolate leuven-dwv1 (KX783225)
NT DWV-like 4 (148bp)	TGAAATGATGAATACTGTATCACCATCGTAAAACGATTGTTAGGCAAATATAGATTGGCGACGCAACCCAGGACCCAGCCGATTACGTAAGCTACCCCA GAGGGACCTAGTCTGAGGCTGAGGAAACAAGTCGCTGG	4867-5017	75% similar to DWV isolate leuven-dwv1 (KX783225)
NT DWV-like 5 (51bp)	GTGAAGATGAACCTGAAATTGTAAGCGATGGGTTAAGAATGTTTGATT	5240-5291	89% similar to DWV isolate leuven-dwv1 (KX783225)
NT DWV-like 6 (421bp)	ATTTTGTGCATGACCTTACAGCTGAGATGAATCAGTCGAGAAATTTGACTGTGTTTACTCGGGTGTATGATCAAATATCGAAATGAAGACCGACCTGAATGTTTTAC GATTTGTATGACTGGTCTCCTGGTATTGGTAAGTCATTCTTAACTGATGCAAGTAGTACTCCGTAACACGGGCATCAAGTGTGTCGTTAATCCCTTATCAGATTATT GGGATCAGTGTGACTTTCAACCAGTCTTGTGCGTAGACGACATGTGGAGTGTGAGACTGGAGCAACTTTGGACAAACAACTAAATTTGCTCTTCAGGTTCACTCTCC AATTGTCCTAAAGCTGATTAGATGGTAAGAAGATGAGGTATAATCCTGAAATCTTCATTTATAATACCAATAAACCTTTCCGGTATTGAT	5375-5863	72% similar to DWV isolate leuven-dwv1 (KX783225)
NT DWV-like 7 (95bp)	TTTGATAATCCTACGTGGCGATTAATATATAATGCATCAAGAAAGGGTTGCCAGAATATTATACACTACCAAGTATGAAATTTTCATTAGATTC	7049-7143	78% similar to DWV isolate leuven-dwv1 (KX783225)
NT DWV-like 8 (203bp)	CACTTGAAGAGAAAGTTTATAACCGTTATTAACCTATAAACGGTTGAAAATTCGTAGTCTGAGGATGCTATTTTGGCGCACCTGGAGTAGAAGGATATGATTCGA TTTCTGGAAATACGAGTGCCGTTTTCCGTTGTCTACAATGAAGCCGGCTGGAACATCAGGAAAGCGCTGGTTGTTTGATATTGAACCTACAAG	8480-8681	77% similar to DWV isolate leuven-dwv1 (KX783225)
NT DWV-like 9 (153bp)	GGGCATAAAGCCACATACGATATTTACTGACTGCCTCAAAGATACATGTTGCCCATAGAGAAGTGTATGATACCGGGTAAAACGAGGATATTTAGTATTGCGCCTGTT CAATATACAATACCTTTTCGACAATATTTCTCGATTATGCGC	8761-8913	77% similar to DWV isolate leuven-dwv1 (KX783225)
NT DWV-like 10	ATTATACAGAGGAAAAAGATAAAGACTGAGATGAGTCGAGTCATGTGGACTATGGCTCAGGAGATCTTGCAACCAAGCCATCTTTGTAGAGATTGGTGTATAGAGTAC	9111-9253	75% similar to DWV isolate leuven-dwv1 (KX783225)

(143bp)	CTTGTGGCATACTTCGGGCTCACCCATCACGGAC		
NT DWV-like 11 (72bp)	CGTGTAGCCACTATAGAAAATGCTAAACAATCTCTAGAGTTGGCATTGCGTTGGGGTCCGCAACTTCAAT	9626-9697	83% similar to DWV isolate leuven-dwv1 (KX783225)
NT DWV-like 12 (44bp)	AAGCTTGGTATTTATGAGGATCTCATTACATGGGAGGAGATGGA	9731-9774	89% similar to DWV isolate leuven-dwv1 (KX783225)
QLD DWV-like 1 (105bp)	TAGACCTCTACGGCATCGAGTAGAGATTAAGAGTAGTGAATAGTATAATCACTGTCTACCACCACATATGGTAATGATATGGTTGTGTAACCGGTTAT	488-592	80% similar to VDV-1 genome (AY251269)
QLD DWV-like 2 (99bp)	ATGTCAGGTTATATTGAATGCTCGAGTATGATTTCTGCGTTGGAGTCGGGACCCCTCGGTCTCCAGGTGTCGCATGTGACGAAAGTGCAGAAAATA	687-785	80% similar to VDV-1 genome (AY251269)
QLD DWV-like 3 (135bp)	ATATGCGATGTACCTAATACAATACCATTCAAGGTACATGCATATTGGCGTGGAGAGATGCAAGTTCGAATACAAGTTAATCGAATAAATCCAAGTAGGTC AATTGC AAGCCACTTGGTATTATATGGATTAT	2045-2179	84% similar to VDV-1 genome (AY251269)
QLD DWV-like 4 (232bp)	ATGGATTATGACACTTAAAGCATCAAGTTATGTTGATTGTTGCAAGAAACAATAGCTTTACGTTGAAGTGCCTTATGTGCATATAGACCATGGTGGTGT ATGTATGTACAAGTACCGTTGATACCTATGGAAGCTGTTGCAGATACCATAGATATCAATGTGTATTTAAGAGGAGGCTCTTCATTTGAAGTATGTATTCTGTACAACC TAGTGTAGGATT	3029-3315	69% similar to VDV-1 genome (AY251269)
QLD DWV-like 5 (326bp)	TTTCCGCAACACCAGAAGGACCTAGTGCTGAGGCTGAGGAAACAAGTGCCTGGGTATCCGTTATTTATAACGGTGTGGTGAATATGTTGAATGTGGTGGCACAAAAGC CAAAACAATTTAAAGACTGGGTAAAATGTTTCCCAAGATTTGGAAACAATGTAGAAGTAGTAATCAGGTATTATATCTTTAAGAACACCTTTGAGGTTTTGAA GAATTTGTAGGACTAATCCTGCCCTCGCCTATTGAAGGCTGTGAGCGATGAGCCCGAGATACTCAAAGCGTGGTTAAGAATGTTGTACTTAAATGATCCCAAAT	4965-5307	77% similar to VDV-1 genome (AY251269)
QLD DWV-like 6 (111bp)	TACACCTGTCAACCTGGAATTAAGTGTGTTGTAATCCCTCTCTGACTATTGGGATCAGTGTGATTCCAACCGGTTTTGTGCGTTGATGATATGGGAGTGTGGAAA C	5599-5709	82% similar to VDV-1 genome (AY251269)
QLD DWV-like 7 (98bp)	AACTATTTAGTATTTATCCATAAATCATGGTTATACGCAAATCCACGTGGAGACTGATTTTAAATGGTTCAAAGAAAGGTATGCCAGAGTATTT	7013-7110	80% similar to VDV-1 genome (AY251269)
QLD DWV-like 8 (90bp)	TGGAATACGAGTGCAGGTTTTCCATTGTCAACAATGAAACCTGCTGGAACATCTGGAAGAGATGGTGTGATATTGAACTAAAAGAT	8594-8683	80% similar to VDV-1 genome (AY251269)
QLD DWV-like 9 (88bp)	CAACGACTCAAGAAATGAGAAAGAGAGGAATCAAGCCTCACACCATCTCACAGATTGCCTCAAAGATACATGTTGCCAGTAGAAAA	8736-8823	82% similar to VDV-1 genome (AY251269)
QLD DWV-like 10	AGGGAAGTACCACCCCTCGTTGTTTTAAATCTGCTAATAGGAATGAACCTATTATAAGAGTCCAAAAGCAGTGTGATTAGACCATCACTTTGGCTTATAC	9854-10050	85% similar to VDV-1 genome (AY251269)

(198bp)	TAGAGAAAGGATGAGTGCCTCTAAAGGCTCAATCCGTAGTAGAGTAGGTTTAATTAGGATTAAGTGGTACTCTAGGTTAGGTATT		
WA-2 DWV-like 1 (127bp)	ATAGGTAATGATCCCAATACGGTACCATTAGAGTACATGCATATTGGCGTGGAGATATTGAAGTTAAAAATTCAGATAAATCTAATAAATTTCAAGTTGGGCAATTGC AAGCTACTTGGTATTATT	2072-2198	84% similar to DWV isolate Chilensis A1 (JQ413340)
WA-2 DWV-like 2 (91bp)	TGGCGTGGATCTTTGGAGTATCGATTTGATATCGTTGCTTCTCAATTCATACTGGTAGGTTGATTGTAGGCTATATCCCCGGATATGACA	2948-3037	80% similar to DWV isolate Chilensis A1 (JQ413340)
WA-2 DWV-like 3 (98bp)	AATTGTCAAAGTAGTAATCAAGTATTTATTTTCTTTAAGAATACATTTGAAGTATTAAGAAGACTTGGGCTATTTATTTTGTCAAGTAATCCTGC	5150-5247	81% similar to DWV isolate Chilensis A1 (JQ413340)

36 **Supplementary material 3. Alignment of mapped reads from the NT sample to DWV-A (NC_004830), DWV-B (NC_006494) and**
37 **DWV-C (ERS657949) using Geneious (Version 9.1.5, created by Biomatters, Auckland, New Zealand)**

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46 **Supplementary material 4. Australian apiary survey results within the context of comparable molecular-based surveys. In relation**
 47 **to Figure 4, populations with profiles 1-3 are on the lower end of the stress spectrum, populations with profile 4 range across the**
 48 **middle of the stress spectrum and populations with profile 5 are on the high end of the stress spectrum.**

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Survey reference	Country: Region	Increased colony losses recorded	Colony stressors						Stress profile	Commercial-scale beekeepers
			<i>Varroa destructor</i>	DWV complex	Other viruses detected	Viruses tested for but <u>not</u> detected	Other pathogens	Abiotic stressors		
This survey	Australia	No	No	No	BQCV SBV LSV1 LSV2 IAPV ALPV	DWV SBPV KBV CBPV ABPV	Most	Most	1	Yes
Malfroy et al. ¹⁰	Australia: Norfolk Island	No	No	No	LSV	DWV ABPV IAPV KBV BQCV SBV CBPV SBPV	Few	Few	2	No

Kajobe et al. ¹¹	Uganda	No	No	No	BQCV	DWV SBV CBPV ABPV IAPV	Few	Few	2	No
Shutler et al. ¹²	Canada : Newfoundland	No	No	DWV-A	BQCV	IAPV KBV SBV	Few	Few	3	No
Sanpa and Chantawannakul ¹³	Thailand : Northern provinces	No	Yes	DWV-A	ABPV KBV SBV	BQCV CBPV	Most	Some	4	Yes
Ai et al. ¹⁴	China	No	Yes	DWV-A	ABPV IAPV BQCV SBV CBPV	KBV	Most	Most	4	Yes
Kojima et al. ¹⁵	Japan	No	Yes	DWV-A	BQCV SBV IAPV	CBPV KBV ABPV	Most	Most	4	Yes
Strauss et al. ¹⁶	South Africa	No	Yes	DWV-B	BQCV IAPV	DWV-A SBV CBPV KBV VdMLV	Most	Some	4	Yes
Muli et al. ¹⁷	Kenya	No	Yes	DWV-A	BQCV ABPV	SBV KBV IAPV	Some	Some	4	No

						CBPV				
Yang et al. ¹⁸	China	No	Yes	DWV-A	BQCV IAPV SBV CBPV	ABPV KBV	Most	Most	4	Yes
Mondet et al. ¹⁹	New Zealand	No	Yes	DWV-A	BQCV KBV SBV CBPV		Some	Most	4	Yes
Adjlane et al. ²⁰	Algeria	No	Yes	DWV-A	BQCV SBV ABPV IAPV CBPV	DWV-B	Most	Some	4	No
Tsevegmid et al. ²¹	Mongolia: northern regions	No	Yes	DWV-A	SBV BQCV CBPV	ABPV KBV IAPV LSV2	Some	Some	4	No
Giacobino et al. ²²	Argentina	No	Yes	DWV-A	BQCV ABPV CBPV	KBV IAPV	Most	Some	4	Yes
Anido et al. ²³	Uruguay	No	Yes	DWV-A	BQCV SBV ABPV	IAPV KBV	Most	Some	4	Yes
Teixeira et al. ²⁴	Brazil: southeast	No	Yes	DWV-A	BQCV ABPV	KBV CBPV	Most	Some	4	Yes

	region					SBV				
Soroker et al. ²⁵	Israel	No	Yes	DWV-A DWV-B	BQCV SBV ABPV IAPV	KBV CBPV	Most	Most	4	Yes
Rodriguez et al. ²⁶	Chile: Biobio region	No	Yes	DWV-A	BQCV SBV ABPV		Most	Some	4	Yes
Desai et al. ²⁷	Canada	Yes	Yes	DWV-A	BQCV SBV ABPV KBV IAPV CBPV		Most	Most	5	Yes
Forgach et al. ²⁸	Hungary	Yes	Yes	DWV-A	BQCV SBV ABPV	KBV CBPV	Most	Most	5	Yes
Tentcheva et al. ⁸	France	Yes	Yes	DWV-A	BQCV SBV ABPV KBV CBPV		Most	Most	5	Yes
Berenyi et al. ²⁹	Austria	Yes	Yes	DWV-A	BQCV SBV ABPV CBPV	KBV	Most	Most	5	Yes

Genersch et al. ³⁰	Germany	Yes	Yes	DWV-A	SBV ABPV KBV	IAPV	Most	Most	5	Yes
Berthoud et al. ³¹	Switzerland	Yes	Yes	DWV-A	ABPV	KBV CBPV	Most	Most	5	Yes
Nguyen et al. ³²	Belgium	Yes	Yes	DWV-A	BQCV SBV ABPV CBPV		Most	Most	5	Yes
Baker and Schroeder ³³	England: Devon	Yes	Yes	DWV-A	SBV BQCV ABPV	KBV CBPV	Most	Most	5	Yes
Antunez et al. ³⁴	Spain	Yes	Yes	DWV-A	BQCV SBV IAPV KBV		Most	Most	5	Yes
Cornman et al. ³⁵	USA	Yes	Yes	DWV-A	BQCV SBV LSV1 LSV2 ABPV IAPV KBV		Most	Most	5	Yes
Cox-Foster et al. ³⁶	USA	Yes	Yes	DWV-A	CBPV SBV BQCV ABPV		Most	Most	5	Yes

					KBV IAPV					
Dainat et al. ³⁷	Switzerland	Yes	Yes	DWV-A	BQCV SBV SBPV ABPV	IAPV KBV CBPV	Most	Most	5	Yes
Budge et al. ³⁸	England and Wales	Yes	Yes	DWV-A	BQCV CBPV SBV ABPV KBV		Most	Most	5	Yes
Welch et al. ³⁹	USA: Massachusetts	Yes	Yes	DWV-A	BQCV SBV	ABPV KBV IAPV CBPV	Most	Most	5	Yes
Bacandritsos et al. ₄₀	Greece	Yes	Yes	DWV-A	CBPV BQCV SBV ABPV		Most	Most	5	Yes
Runckel et al. ⁴	USA	Yes	Yes	DWV-A	BQCV CBPV SBV ABPV KBV IAPV		Most	Most	5	Yes
Traynor et al. ⁴¹	USA	Yes	Yes	DWV-A	BQCV SBV CBPV	SBPV	Most	Most	5	Yes

					LSV2 ABPV IAPV KBV					
Tozkar et al. ⁴²	Turkey	Yes	Yes	DWV-A DWV-B	BQCV LSV CBPV ABPV KBV IAPV	SBV SBPV	Most	Some	5	Yes
Ravoet et al. ⁴³	Belgium	Yes	Yes	DWV-A	BQCV SBV CBPV LSV ABPV VdMLV ALPV	IAPV SBPV	Most	Most	5	Yes
Amiri et al. ⁴⁴	Denmark	Yes	Yes	DWV-A	SBV BQCV CBPV ABPV KBV		Most	Most	5	Yes
Gajger et al. ⁴⁵	Croatia	Yes	Yes	DWV-A	BQCV SBV ABPV CBPV	KBV IAPV	Most	Most	5	Yes
Granberg et al. ⁴⁶	Spain: Navarre	Yes	Yes	No	IAPV ALPV	BQCV SBV ABPV	Most	Most	5	Yes

					LSV	KBV DWV				
Nielsen et al. ⁴⁷	Denmark	Yes	Yes	DWV-A	BQCV CBPV SBV ABPV KBV		Most	Most	5	Yes

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52 **Supplementary material references**

- 53 1 Maori, E. *et al.* Isolation and characterization of Israeli acute paralysis virus, a dicistrovirus affecting honeybees in Israel: evidence for diversity due
54 to intra- and inter-species recombination. *Journal of General Virology* **88**, 3428-3438, doi:10.1099/vir.0.83284-0 (2007).
- 55 2 Stoltz, D., Shen, X. R., Boggis, C. & Sisson, G. Molecular diagnosis of Kashmir bee virus infection. *Journal of Apicultural Research* **34**, 153-160 (1995).
- 56 3 Benjeddou, M., Leat, N., Allsopp, M. & Davison, S. Detection of acute bee paralysis virus and black queen cell virus from honeybees by reverse
57 transcriptase PCR. *Applied and Environmental Microbiology* **67**, 2384-2387, doi:10.1128/aem.67.5.2384-2387.2001 (2001).
- 58 4 Runckel, C. *et al.* Temporal analysis of the honey bee microbiome reveals four novel viruses and seasonal prevalence of known viruses, *Nosema* and
59 *Crithidia*. *PLOS One* **6**, e20656 (2011).
- 60 5 Meeus, I., Smaghe, G., Siede, R., Jans, K. & de Graaf, D. C. Multiplex RT-PCR with broad-range primers and an exogenous internal amplification
61 control for the detection of honeybee viruses in bumblebees. *Journal of Invertebrate Pathology* **105**, 200-203 (2010).
- 62 6 de Miranda, J. R. *et al.* Genetic characterization of slow bee paralysis virus of the honeybee (*Apis mellifera* L.). *Journal of General Virology* **91**, 2524-
63 2530, doi:10.1099/vir.0.022434-0 (2010).
- 64 7 Grabensteiner, E. *et al.* Sacbrood virus of the honeybee (*Apis mellifera*): Rapid identification and phylogenetic analysis using reverse transcription-
65 PCR. *Clinical and Diagnostic Laboratory Immunology* **8**, 93-104 (2001).
- 66 8 Tentcheva, D. *et al.* Prevalence and seasonal variations of six bee viruses in *Apis mellifera* L. and *Varroa destructor* mite populations in France.
67 *Applied and Environmental Microbiology* **70**, 7185-7191, doi:10.1128/aem.70.12.7185-7191.2004 (2004).
- 68 9 Ribiere, M., Faucon, J. P. & Pepin, M. Detection of chronic honey bee (*Apis mellifera* L.) paralysis virus infection: application to a field survey.
69 *Apidologie* **31**, 567-577, doi:10.1051/apido:2000147 (2000).

- 70 10 Malfroy, S. F., Roberts, J. M. K., Perrone, S., Maynard, G. & Nadine, C. A pest and disease survey of the isolated Norfolk Island honey bee (*Apis*
71 *mellifera*) population. *Journal of Apicultural Research*,
72 doi:10.1080/00218839.2016.1189676 (2016).
- 73 11 Kajobe, R. *et al.* First molecular detection of a viral pathogen in Ugandan honey bees. *Journal of Invertebrate Pathology* **104**, 153-156,
74 doi:10.1016/j.jip.2010.02.007 (2010).
- 75 12 Shutler, D. *et al.* Honey Bee *Apis mellifera* Parasites in the Absence of *Nosema ceranae* Fungi and *Varroa destructor* Mites. *Plos One* **9**,
76 doi:10.1371/journal.pone.0098599 (2014).
- 77 13 Sanpa, S. & Chantawannakul, P. Survey of six bee viruses using RT-PCR in Northern Thailand. *Journal of Invertebrate Pathology* **100**, 116-119,
78 doi:10.1016/j.jip.2008.11.010 (2009).
- 79 14 Ai, H. X., Yan, X. & Han, R. C. Occurrence and prevalence of seven bee viruses in *Apis mellifera* and *Apis cerana* apiaries in China. *Journal of*
80 *Invertebrate Pathology* **109**, 160-164, doi:10.1016/j.jip.2011.10.006 (2012).
- 81 15 Kojima, Y. *et al.* Infestation of Japanese Native Honey Bees by Tracheal Mite and Virus from Non-native European Honey Bees in Japan. *Microbial*
82 *Ecology* **62**, 895-906, doi:10.1007/s00248-011-9947-z (2011).
- 83 16 Strauss, U. *et al.* Seasonal prevalence of pathogens and parasites in the savannah honeybee (*Apis mellifera* *scutellata*). *Journal of Invertebrate*
84 *Pathology* **114**, 45-52, doi:10.1016/j.jip.2013.05.003 (2013).
- 85 17 Muli, E. *et al.* Evaluation of the Distribution and Impacts of Parasites, Pathogens, and Pesticides on Honey Bee (*Apis mellifera*) Populations in East
86 Africa. *Plos One* **9**, doi:10.1371/journal.pone.0094459 (2014).
- 87 18 Yang, B., Peng, G., Li, T. & Kadowaki, T. Molecular and phylogenetic characterization of honey bee viruses, *Nosema* microsporidia, protozoan
88 parasites, and parasitic mites in China. *Ecology and Evolution* **3**, 298-311, doi:10.1002/ece3.464 (2013).
- 89 19 Mondet, F., de Miranda, J. R., Kretzschmar, A., Le Conte, Y. & Mercer, A. R. On the Front Line: Quantitative Virus Dynamics in Honeybee (*Apis*
90 *mellifera* L.) Colonies along a New Expansion Front of the Parasite *Varroa destructor*. *Plos Pathogens* **10**, doi:10.1371/journal.ppat.1004323 (2014).
- 91 20 Adjlane, N., Dainat, B., Gauthier, L. & Dietemann, V. Atypical viral and parasitic pattern in Algerian honey bee subspecies *Apis mellifera* *intermissa*
92 and *A-m. sahariensis*. *Apidologie* **47**, 631-641, doi:10.1007/s13592-015-0410-x (2016).
- 93 21 Tsevegmid, K., Neumann, P. & Yanez, O. The Honey Bee Pathosphere of Mongolia: European Viruses in Central Asia. *Plos One* **11**,
94 doi:10.1371/journal.pone.0151164 (2016).
- 95 22 Giacobino, A. *et al.* *Varroa destructor* and viruses association in honey bee colonies under different climatic conditions. *Environmental Microbiology*
96 *Reports* **8**, 407-412, doi:10.1111/1758-2229.12410 (2016).
- 97 23 Anido, M. *et al.* Prevalence and distribution of honey bee pests and pathogens in Uruguay. *Journal of Apicultural Research* **54**, 532-540,
98 doi:10.1080/00218839.2016.1175731 (2015).
- 99 24 Teixeira, E. W., Chen, Y. P., Message, D., Pettis, J. & Evans, J. D. Virus infections in Brazilian honey bees. *Journal of Invertebrate Pathology* **99**, 117-
100 119, doi:10.1016/j.jip.2008.03.014 (2008).
- 101 25 Soroker, V. *et al.* Evaluation of colony losses in Israel in relation to the incidence of pathogens and pests. *Apidologie* **42**, 192-199,
102 doi:10.1051/apido/2010047 (2011).

103 26 Rodriguez, M. *et al.* Prevalence and phylogenetic analysis of honey bee viruses in the Biobio Region of Chile and their association with other honey
104 bee pathogens. *Chilean Journal of Agricultural Research* **74**, 170-177, doi:10.4067/s0718-58392014000200007 (2014).

105 27 Desai, S. D., Kumar, S. & Currie, R. W. Occurrence, detection, and quantification of economically important viruses in healthy and unhealthy honey
106 bee (Hymenoptera: Apidae) colonies in Canada. *Canadian Entomologist* **148**, 22-35, doi:10.4039/tce.2015.23 (2016).

107 28 Forgach, P., Bakonyi, T., Tapasztai, Z., Nowotny, N. & Rusvai, M. Prevalence of pathogenic bee viruses in Hungarian apiaries: Situation before joining
108 the European Union. *Journal of Invertebrate Pathology* **98**, 235-238, doi:10.1016/j.jip.2007.11.002 (2008).

109 29 Berenyi, O., Bakonyi, T., Derakhshifar, I., Koglbberger, H. & Nowotny, N. Occurrence of six honeybee viruses in diseased Austrian apiaries. *Applied
110 and Environmental Microbiology* **72**, 2414-2420, doi:10.1128/aem.72.4.2414-2420.2006 (2006).

111 30 Genersch, E. *et al.* The German bee monitoring project: a long term study to understand periodically high winter losses of honey bee colonies.
112 *Apidologie* **41**, 332-352, doi:10.1051/apido/2010014 (2010).

113 31 Berthoud, H., Imdorf, A., Haueter, M., Radloff, S. & Neumann, P. Virus infections and winter losses of honey bee colonies (*Apis mellifera*). *Journal of
114 Apicultural Research* **49**, 60-65, doi:10.3896/ibra.1.49.1.08 (2010).

115 32 Nguyen, B. K. *et al.* Effects of honey bee virus prevalence, Varroa destructor load and queen condition on honey bee colony survival over the winter
116 in Belgium. *Journal of Apicultural Research* **50**, 195-202, doi:10.3896/ibra.1.50.3.03 (2011).

117 33 Baker, A. & Schroeder, D. Occurrence and genetic analysis of picorna-like viruses infecting worker bees of *Apis mellifera* L. populations in Devon,
118 South West England. *Journal of Invertebrate Pathology* **98**, 239-242, doi:10.1016/j.jip.2008.02.010 (2008).

119 34 Antunez, K. *et al.* Low prevalence of honeybee viruses in Spain during 2006 and 2007. *Research in Veterinary Science* **93**, 1441-1445,
120 doi:10.1016/j.rvsc.2012.03.006 (2012).

121 35 Cornman, R. S. *et al.* Pathogen webs in collapsing honey bee colonies. *Plos One* **7**, doi:10.1371/journal.pone.0043562 (2012).

122 36 Cox-Foster, D. L. *et al.* A metagenomic survey of microbes in honey bee colony collapse disorder. *Science* **318**, 283-287,
123 doi:10.1126/science.1146498 (2007).

124 37 Dainat, B., Evans, J. D., Chen, Y. P., Gauthier, L. & Neumann, P. Predictive Markers of Honey Bee Colony Collapse. *Plos One* **7**,
125 doi:10.1371/journal.pone.0032151 (2012).

126 38 Budge, G. E. *et al.* Pathogens as Predictors of Honey Bee Colony Strength in England and Wales. *Plos One* **10**, doi:10.1371/journal.pone.0133228
127 (2015).

128 39 Welch, A., Drummond, F., Tewari, S., Averill, A. & Burand, J. P. Presence and Prevalence of Viruses in Local and Migratory Honeybees (*Apis
129 mellifera*) in Massachusetts. *Applied and Environmental Microbiology* **75**, 7862-7865, doi:10.1128/aem.01319-09 (2009).

130 40 Bacandritsos, N. *et al.* Sudden deaths and colony population decline in Greek honey bee colonies. *Journal of Invertebrate Pathology* **105**, 335-340,
131 doi:10.1016/j.jip.2010.08.004 (2010).

132 41 Traynor, K. S. *et al.* Multiyear survey targeting disease incidence in US honey bees. *Apidologie* **47**, 325-347, doi:10.1007/s13592-016-0431-0 (2016).

133 42 Tozkar, C., Kence, M., Kence, A., Huang, Q. & Evans, J. Metatranscriptomic analyses of honey bee colonies. *Frontiers in genetics* **6**, 100 (2015).

134 43 Ravoet, J. *et al.* Comprehensive bee pathogen screening in Belgium reveals *Crithidia mellificae* as a new contributory factor to winter mortality. *Plos
135 One* **8**, doi:10.1371/journal.pone.0072443 (2013).

136 44 Amiri, E., Meixner, M., Nielsen, S. L. & Kryger, P. Four Categories of Viral Infection Describe the Health Status of Honey Bee Colonies. *Plos One* **10**,
137 doi:10.1371/journal.pone.0140272 (2015).
138 45 Gajger, I. T., Kolodziejek, J., Bakonyi, T. & Nowotny, N. Prevalence and distribution patterns of seven different honeybee viruses in diseased
139 colonies: a case study from Croatia. *Apidologie* **45**, 701-706, doi:10.1007/s13592-014-0287-0 (2014).
140 46 Granberg, F. *et al.* Metagenomic Detection of Viral Pathogens in Spanish Honeybees: Co- Infection by Aphid Lethal Paralysis, Israel Acute Paralysis
141 and Lake Sinai Viruses. *Plos One* **8**, doi:10.1371/journal.pone.0057459 (2013).
142 47 Nielsen, S. L., Nicolaisen, M. & Kryger, P. Incidence of acute bee paralysis virus, black queen cell virus, chronic bee paralysis virus, deformed wing
143 virus, Kashmir bee virus and sacbrood virus in honey bees (*Apis mellifera*) in Denmark. *Apidologie* **39**, 310-314, doi:10.1051/apido:2008007 (2008).
144