

## Supporting Information

### Reconfigurable Three-Dimensional Gold Nanorod Plasmonic Nanostructures on DNA Origami Tripod

#### **Content:**

- I. Materials and methods
- II. Theoretical calculations
- III. Supporting figures
- IV. DNA sequences
- V. References

## I. Materials and methods

**Synthesis of gold nanorods.** A two-step method was used to synthesize gold nanorod according to reference.<sup>1</sup>

a. Synthesis of seeds: A 50  $\mu\text{L}$  of 2% (w/v)  $\text{HAuCl}_4$  solution and 9.5 mL of 100 mM CTAB solution were added to a 20 mL flask. A 600  $\mu\text{L}$  of ice-cold  $\text{NaBH}_4$  solution (10 mM) was added to the flask under vigorous stirring for 2 minutes. The color of the reaction solution changed to yellowish brown quickly. The resulting solution will stay 2 hours for the next step. The resulting solution will act as nucleation points of gold nanorods in the next step.

b. Synthesis of AuNR: In a 20 mL flask, 10 mL of 100 mM CTAB (hexadecyl-trimethyl-ammonium bromide) was first added, followed by the addition of 78  $\mu\text{L}$  of 2% (w/v)  $\text{HAuCl}_4$  solution. A 80  $\mu\text{L}$  of 10 mM  $\text{AgNO}_3$  was then added into the mixture sequentially to produce light yellowish color. Then 48  $\mu\text{L}$  of 0.1 M ascorbic acid was injected into the solution with moderate 5s shaking. Upon the addition of ascorbic acid, the yellowish solution gradually became colorless. Finally, 16  $\mu\text{L}$  of seed was injected and the flask was stirring for 1-2 minutes. The colorless solution turned to red, purple, and brown. The resultant mixture was left undisturbed at 30°C for 12 h for NR growth. The final products were isolated by centrifugation at 8,000 rpm for 10 min followed by removal of the supernatant. No size or shape-selective fractionation was performed. The pellet was suspended in 100  $\mu\text{L}$  water. The concentration was measured by UV spectra (UV2550, Shimadzu) using extinction coefficient of  $1 \times 10^9 \text{ M}^{-1} \text{ cm}^{-1}$  for the longitudinal plasmon resonance of the AuNRs.

**Functionalization of AuNRs with DNA.** Functionalization of AuNRs with thiolated DNA

(5'-TTTTTTTTTTTTTTTTTAGCG-SH-3' for handle-1;

5'-TATAATAATAATAATATTTT-SH-3' for handle-2;

5'-TTATAACTATTCCTAAAAA-SH-3' for handle-3)

was carried out following the low pH route.<sup>2</sup> Four milliliter of 1 nM AuNRs was mixed with 40  $\mu\text{L}$  of 1% sodium dodecyl sulfate (SDS), 400  $\mu\text{L}$  of 10 $\times$  TBE, and 100  $\mu\text{L}$  of 100  $\mu\text{M}$  DNA. Hydrochloric acid (HCl) was used to adjust the pH value of the solution to 3. The disulfide bond in the thiolated oligonucleotides was reduced to monothiol using Tris(carboxyethyl) phosphine hydrochloride (TCEP) (20 mM, 1 hr) in water. The oligonucleotides were purified using size exclusion columns (G-25, GE Healthcare) to get rid of the small molecules. The purified DNA was added to AuNR solution ( $\text{OD} \sim 1$ ) containing 0.01% (w/v) SDS with a molecular ratio of 4000: 1. After incubation of the AuNRs with DNA for 1 hour, 5 M NaCl was added to bring the final concentration of NaCl to 500 mM. The solution was then gently shaken overnight. After that, AuNR-DNA conjugates were centrifuged at 8000 rpm for 25 minutes. The pellet was suspended in 1 mL 0.5 $\times$ TBE buffer containing 200 mM NaCl while the supernatant was discarded. The same centrifugation was repeated three times

to remove the excessive thiolated DNA completely. The final concentration of AuNRs was estimated with UV-Vis absorption spectroscopy using extinction coefficient of  $1 \times 10^9 \text{ M}^{-1} \text{ cm}^{-1}$  for the longitudinal plasmon resonance of the AuNRs.

**Agarose gel electrophoresis.** Annealed DNA origami samples and AuNR decorated tripod structures were subjected to agarose gel electrophoresis for purification. Samples were run in 1% agarose gel in a  $0.5 \times \text{TBE-Mg}^{2+}$  buffer (45 mM Tris, 45 mM Boric acid, 1 mM EDTA, 10 mM  $\text{MgCl}_2$ ) at 60V for 3 h within an ice-water bath.

## II. Theoretical calculations

We present a theoretical study of the optical response of the symmetric triple nanorods (TNRs) based on mode-coupling analysis and the finite-difference time-domain (FDTD) simulation. To gain the basic mechanism, we first perform a mode-coupling analysis. Each nanorod (NR) is modeled by an oscillator (with oscillation amplitude  $a$ ,  $b$ , or  $c$ ). The coupling among the oscillators is described by the following equations

$$\ddot{a} + \omega_0^2 a + \gamma \dot{a} + tb + tc = E_a,$$

$$\ddot{b} + \omega_0^2 b + \gamma \dot{b} + tc + ta = E_b,$$

$$\ddot{c} + \omega_0^2 c + \gamma \dot{c} + tb + ta = E_c,$$

Where oscillation frequency  $\omega_0$  is the plasmon resonance frequency (depending on the aspect ratio of the NR), the  $\gamma$  is the relaxation constant,  $E_a$ ,  $E_b$  and  $E_c$  depend on the incident field polarization. We find that there are one eigenmode with frequency  $\sqrt{\omega_0^2 + 2t}$  (with polarization along the central symmetry axis) and two degenerate modes with frequency  $\sqrt{\omega_0^2 - t}$  (with polarizations perpendicular to the central symmetry axis) [we have neglected  $\gamma$  for the eigenmodes analysis]. Therefore, we may observe two peaks (one with wavelength shorter than that of the single NR, the other with wavelength longer than that of the single NR) in the scattering spectra, though there should be three modes in general.

The FDTD method is applied to simulate the scattering spectrum of the structure. The length and the radius of the rods are 38 nm and 6 nm. The interface distances among the ends of the NRs are 6nm for  $\theta=90^\circ$ , 9 nm for  $\theta=60^\circ$ , and 12.5 nm for  $\theta=30^\circ$ , respectively. The background is air. The optical response of the TNR depends on the incident field polarization (relative to TNR). We first consider the case of  $\theta=90^\circ$  as an example. Similar discussions can be applied to the cases with  $\theta=60^\circ$  and  $\theta=30^\circ$ . There are mainly three types of contributions to the scattering spectrum. The first type of contribution (we call it type A) to the peak with wavelength longer than that of the single NR

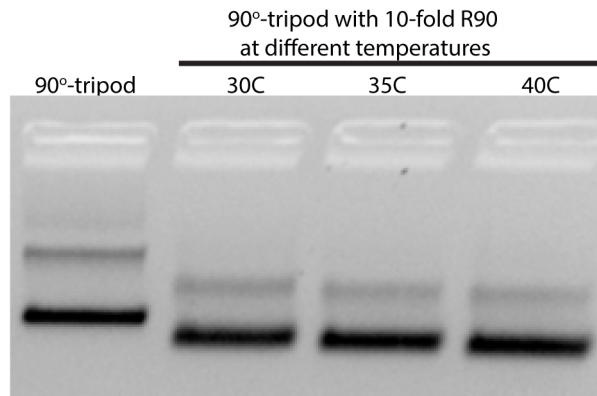
comes mainly from the incident field with polarizations outside the cone formed by the TNRs. The second type of contribution (type B) to the peak with wavelength shorter than that of the single NR comes mainly from the incident field with polarizations within cone formed by the TNRs. The third type (type C) contributes to both peaks. This type of scattering spectrum is mainly due to the plasmonic analogue of electromagnetically induced transparency (EIT), for example from the incident field with direction along one of the NR as shown in Figure S14.

The total scattering spectrum observed in experiment is the summation of the three types of contributions. The type A scattering spectrum is from the fields with the incident angles (with respect to the central symmetry axis)  $0 \leq \alpha < 35.3^\circ$ , which ensures that the incident field polarization outside the core. We use the scattering spectrum of one typical incident field with  $\alpha = 0^\circ$  and the field amplitude proportional to the corresponding solid angle to mimic the type A scattering spectrum. The type B scattering spectrum is from the fields with incident angles  $35.3^\circ < \alpha \leq \alpha_{\max} = 70^\circ$  (we have assume the maximum incident angle to be  $\alpha_{\max} = 70^\circ$ ), which ensures that the incident field polarization within the core. We use the scattering spectrum of one typical incident field with  $\alpha = 70^\circ$  and the field amplitude proportional to the corresponding solid angle to mimic the type B scattering spectrum. The type C scattering spectrum is calculated based on the incident direction along the NR. The relative weights of type A and type B scattering spectrum are determined by the corresponding solid angle related field intensity. The relative weight of type C scattering spectrum to that of type B is chosen as 4. Thus, the weights are  $0.151A+B+4C$  for  $\theta=90^\circ$ ,  $3.20A+B+4C$  for  $\theta=60^\circ$ , and  $A+4C$  for  $\theta=30^\circ$ . The scattering spectra are shown in the following figures. We see two peaks as predicted by the mode-coupling analysis (Fig. S15). And we have a qualitative agreement between the theory and the experiment. (Note that for the case of  $\theta=30^\circ$ , the type B scattering vanishes since there is no polarization within the cone for all incident fields with  $0^\circ \leq \alpha \leq \alpha_{\max} = 70^\circ$ .)

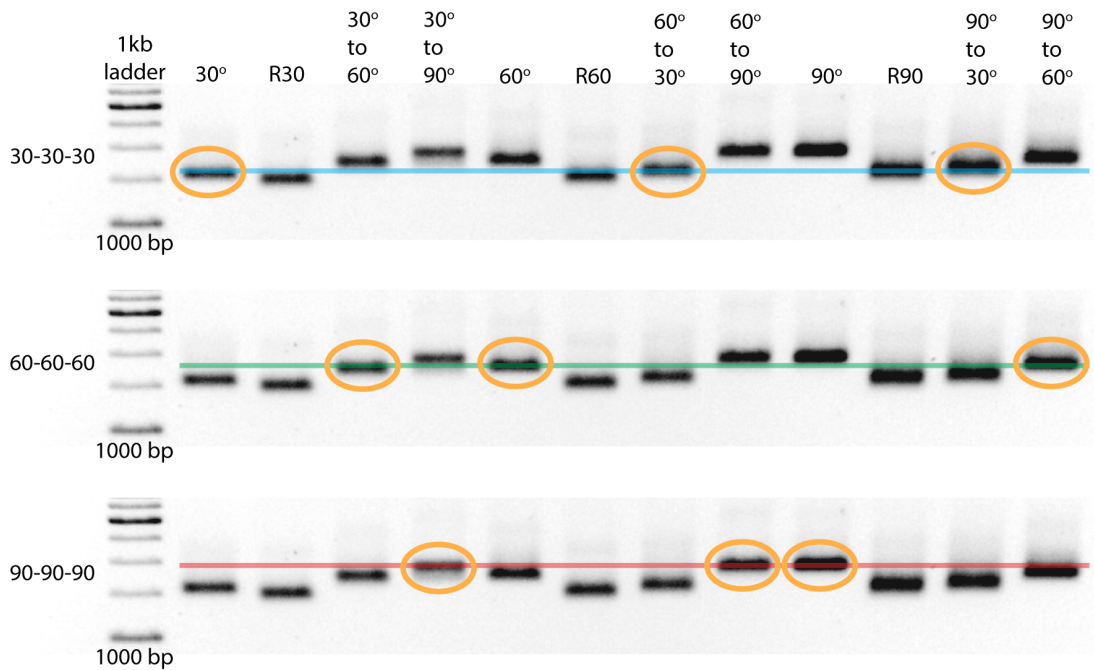
This type of scattering spectrum is mainly due to the plasmonic analogue of electromagnetically induced transparency (EIT).<sup>S3,S4</sup> Figure S14 shows the scattering spectra of type C contribution for  $\theta=30^\circ, 60^\circ, 90^\circ$ , where we see clearly dips in the scattering intensity of the spectra. To further clarify the mechanism, we show the near field maps for  $\theta=90^\circ$  in S16 (The main feature is the same for the configurations of  $\theta=30^\circ, 60^\circ$ ). In this case, the incident field direction is along Z axis (the long axis of one nanorod, nanorod I) and the polarization of the electric field is along the Y axis. One can see that the incident field mainly generates net dipole moments of the nanorods II and III, which lead to the bright field modes. The near field interaction between the nanorods II/III and nanorod I induces the quadrupole excitation of nanorod I, which forms the dark mode. The interference among the fields from bright

modes and dark mode leads to a dip in the scattering intensity of the spectra. This plasmonic analogue EIT is similar as that in the T-shaped nanorod dimer.<sup>S5</sup> In general, the angle among the nanorods ( $\theta$ ) and the distance between the ends of the nanorods ( $d$ ) have impact on the dip in the scattering spectra. Our calculations have also shown that the dip becomes deeper with decreasing the inter-nanorod end distance  $d$  (i.e., increase of the interaction among the nanorods) as shown in figure S17.<sup>S4</sup>

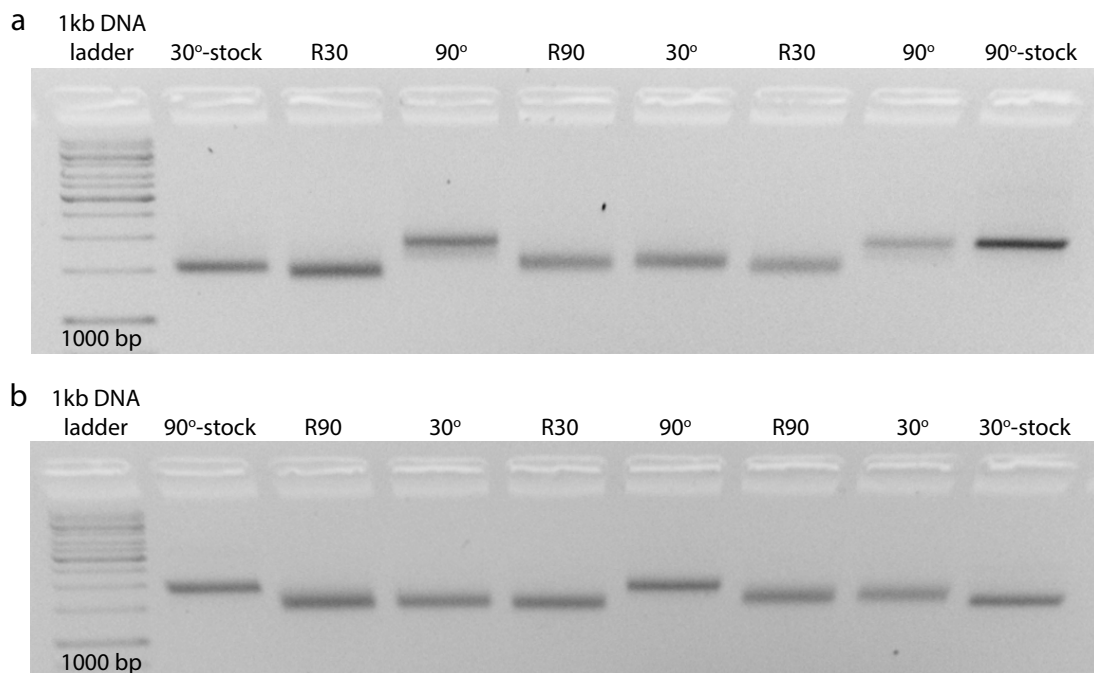
### III. Supporting figures



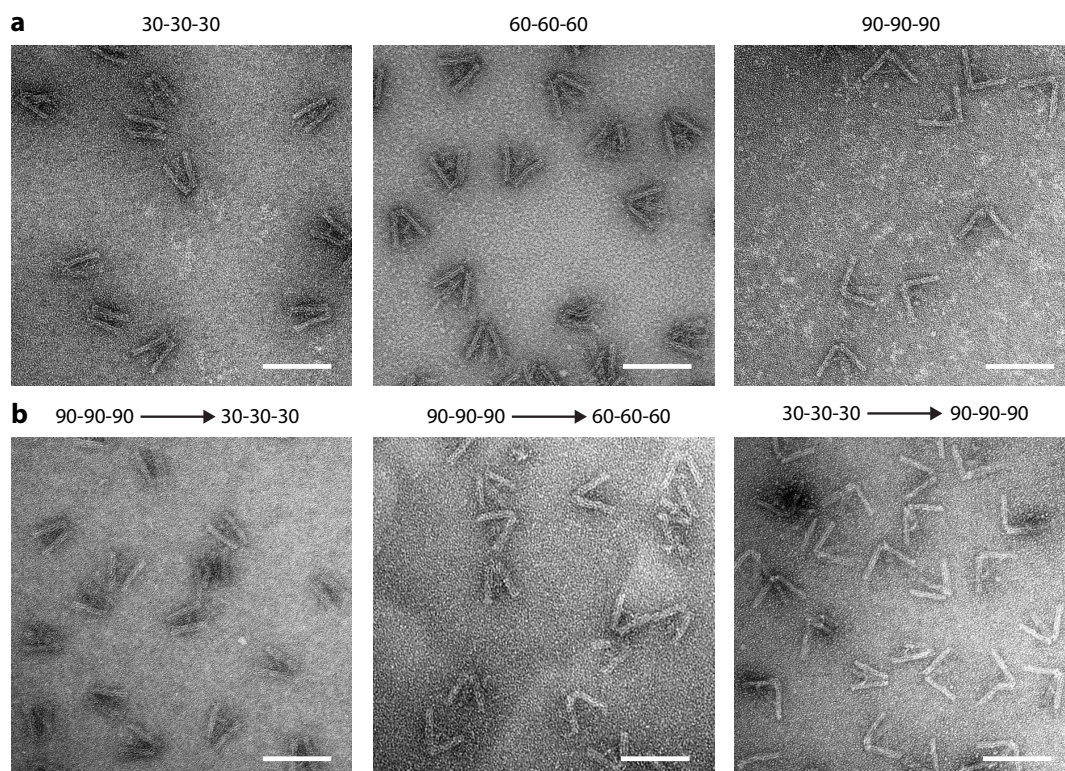
**Figure S1.** Agarose gel electrophoresis image of 90-90-90 tripod (90°-tripod) treated with 10-fold excess of R90 (releasing strands) at various annealing temperatures (at 30°C, 35°C and 40°C) for 4 hours followed by keeping at room temperature for overnight.



**Figure S2.** Agarose gel image showing conversion of one tripod conformation to the other two conformations. Here same gel image is presented three times to highlight individual tripod conformations (30°, 60° and 90°). Bands R30, R60 and R90 correspond to the released conformation (no locking state) from 30°, 60° and 90°, respectively.

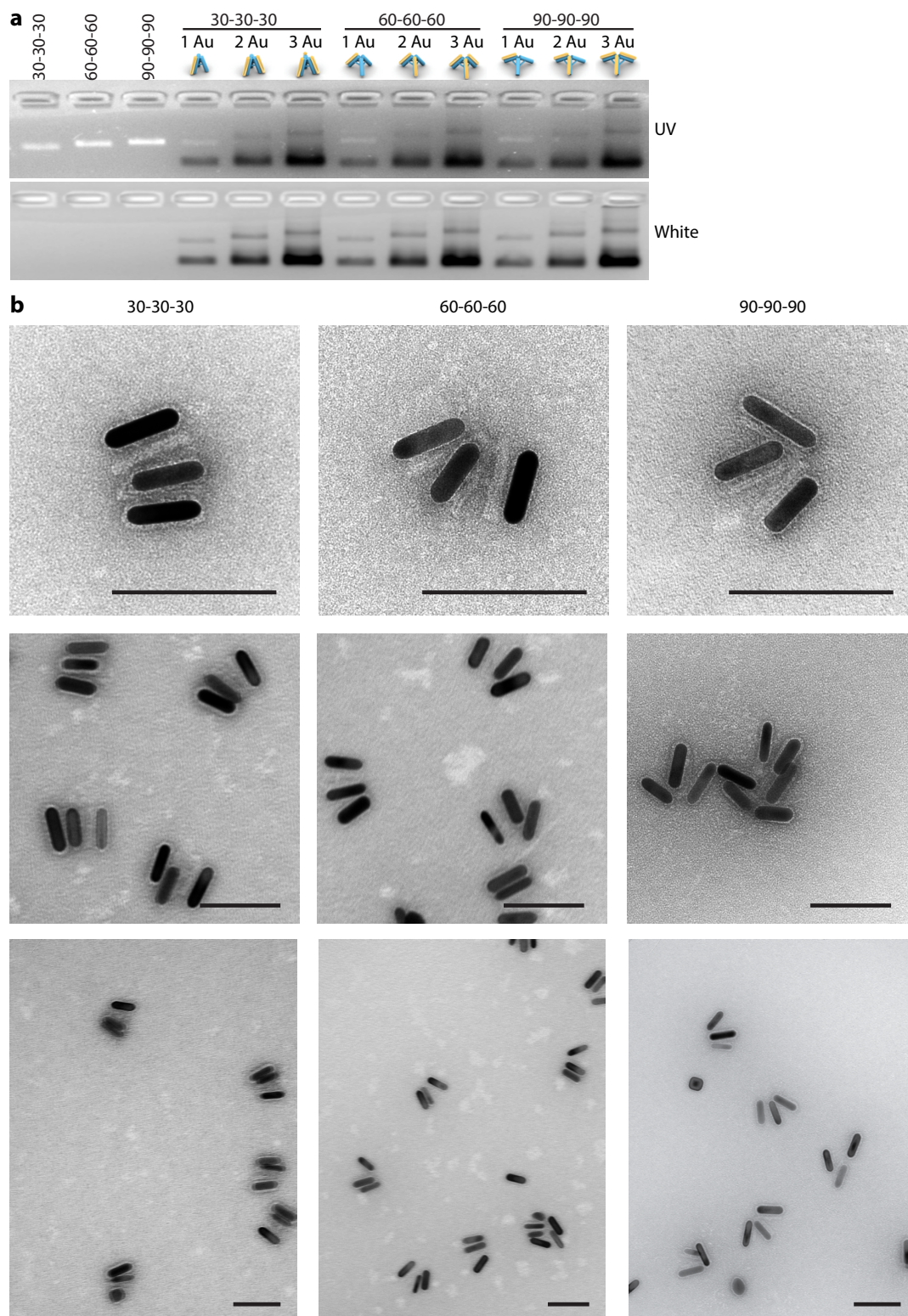


**Figure S3.** (a) Agarose gel electrophoresis image showing back and forth conversion of 30-30-30 tripod ( $30^\circ$ ) to 90-90-90 tripod ( $90^\circ$ ). The bands for R30 and R90 correspond to strut released conformation (no locking state) achieved from  $30^\circ$ - and  $90^\circ$ -tripod, respectively. (b) Agarose gel electrophoresis image showing back and forth conversion of 90-90-90 tripod ( $90^\circ$ ) to 30-30-30 tripod ( $30^\circ$ ).



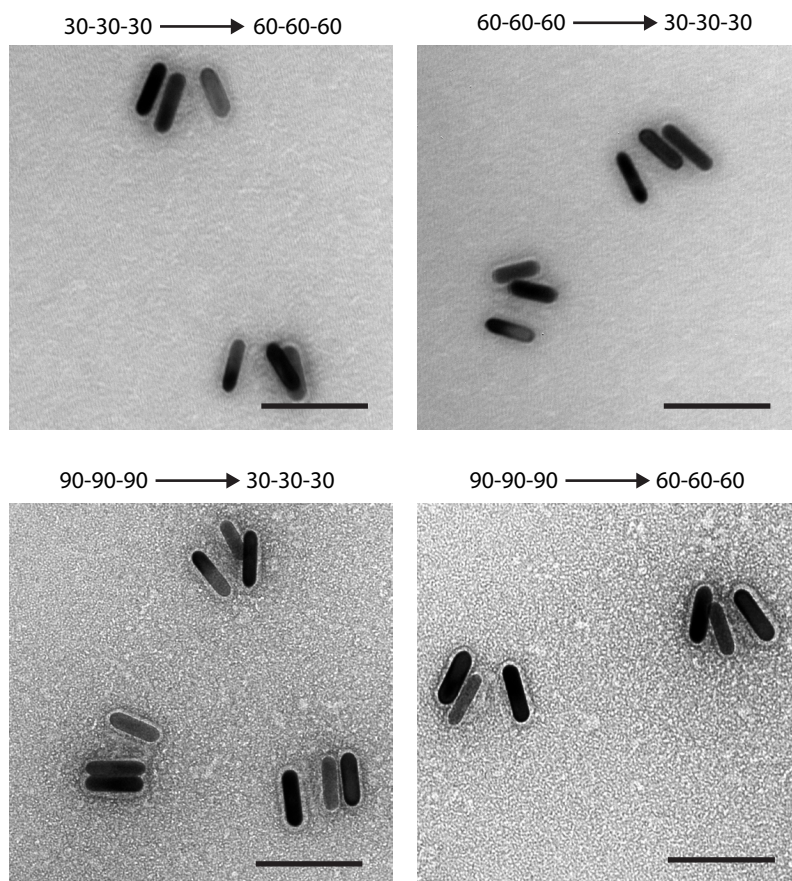
**Figure S4.** (a) TEM images of 30-30-30, 60-60-60 and 90-90-90 tripod. (b)

TEM images of 30-30-30 converted from 90-90-90, 60-60-60 converted from 90-90-90 and 90-90-90 converted from 30-30-30. Scale bar, 100 nm.

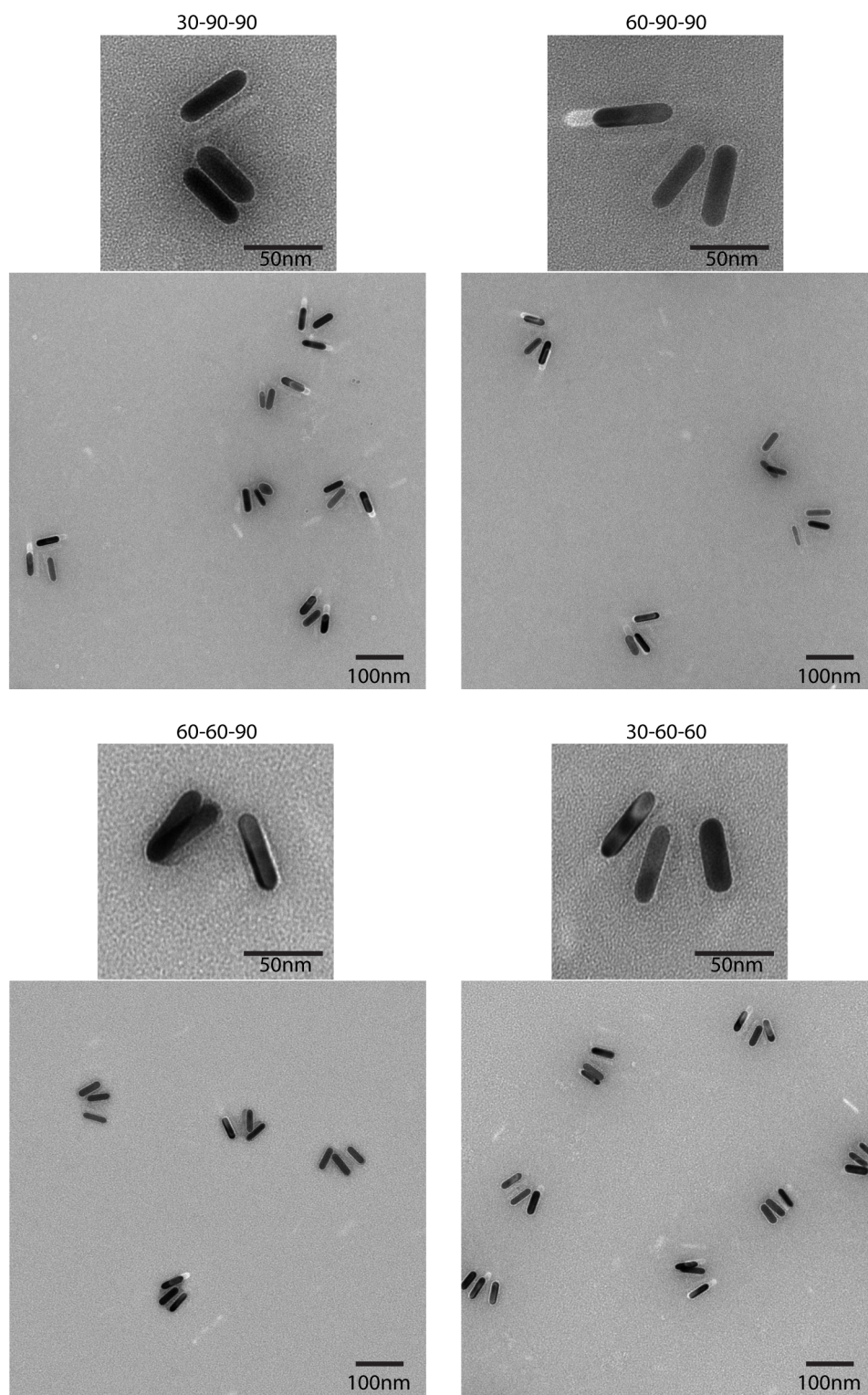


**Figure S5.** (a) Agarose gel electrophoresis image (UV and white light illuminated) of tripods with one, two and three AuNRs. (b) TEM images of tripods with AuNRs. Scale bars, 100 nm.

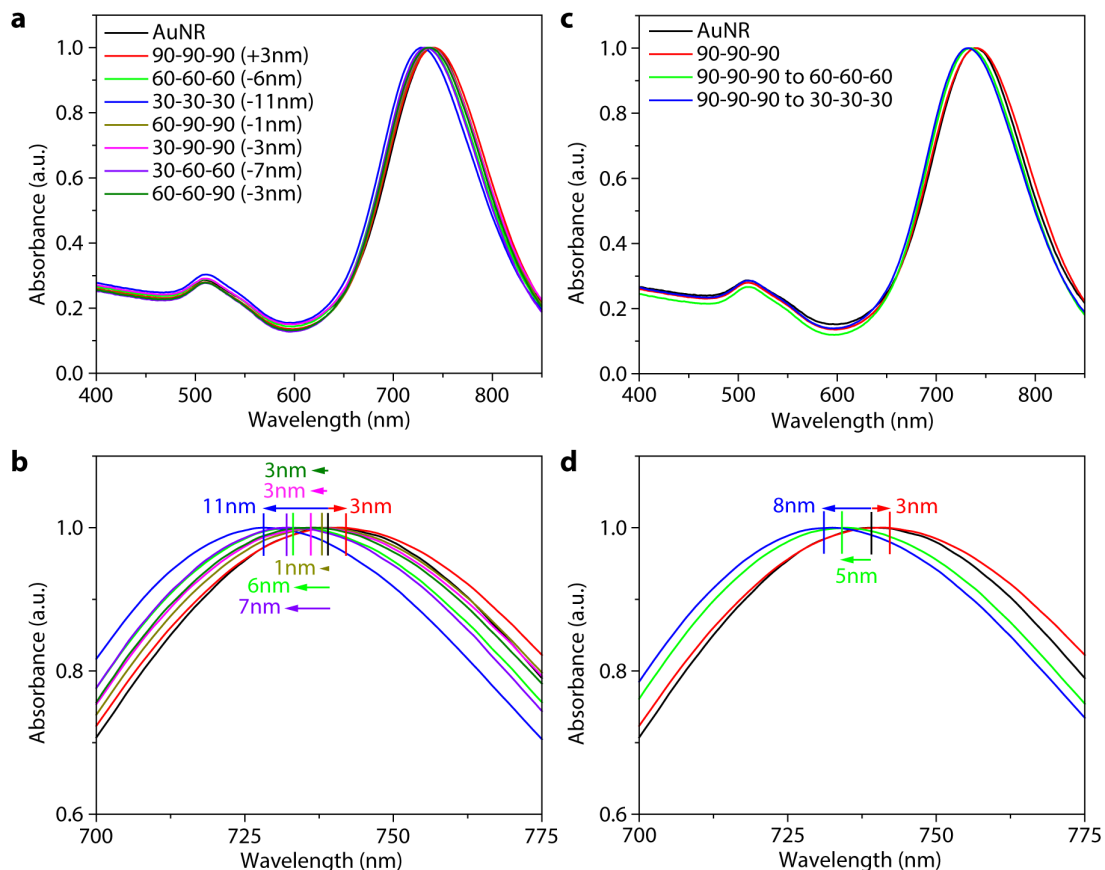




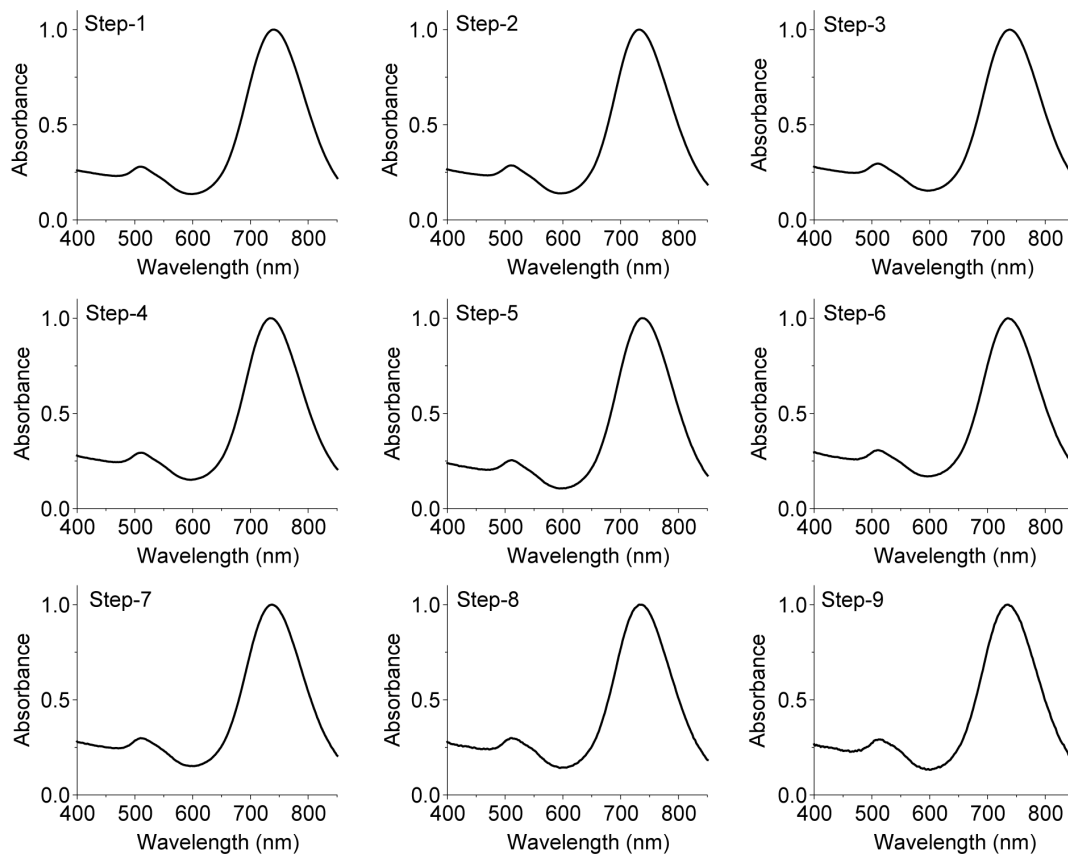
**Figure S6.** TEM images of 60-60-60, 30-30-30, 30-30-30 and 60-60-60 converted from 30-30-30, 60-60-60, 90-90-90 and 90-90-90, respectively. Scale bars, 100 nm.



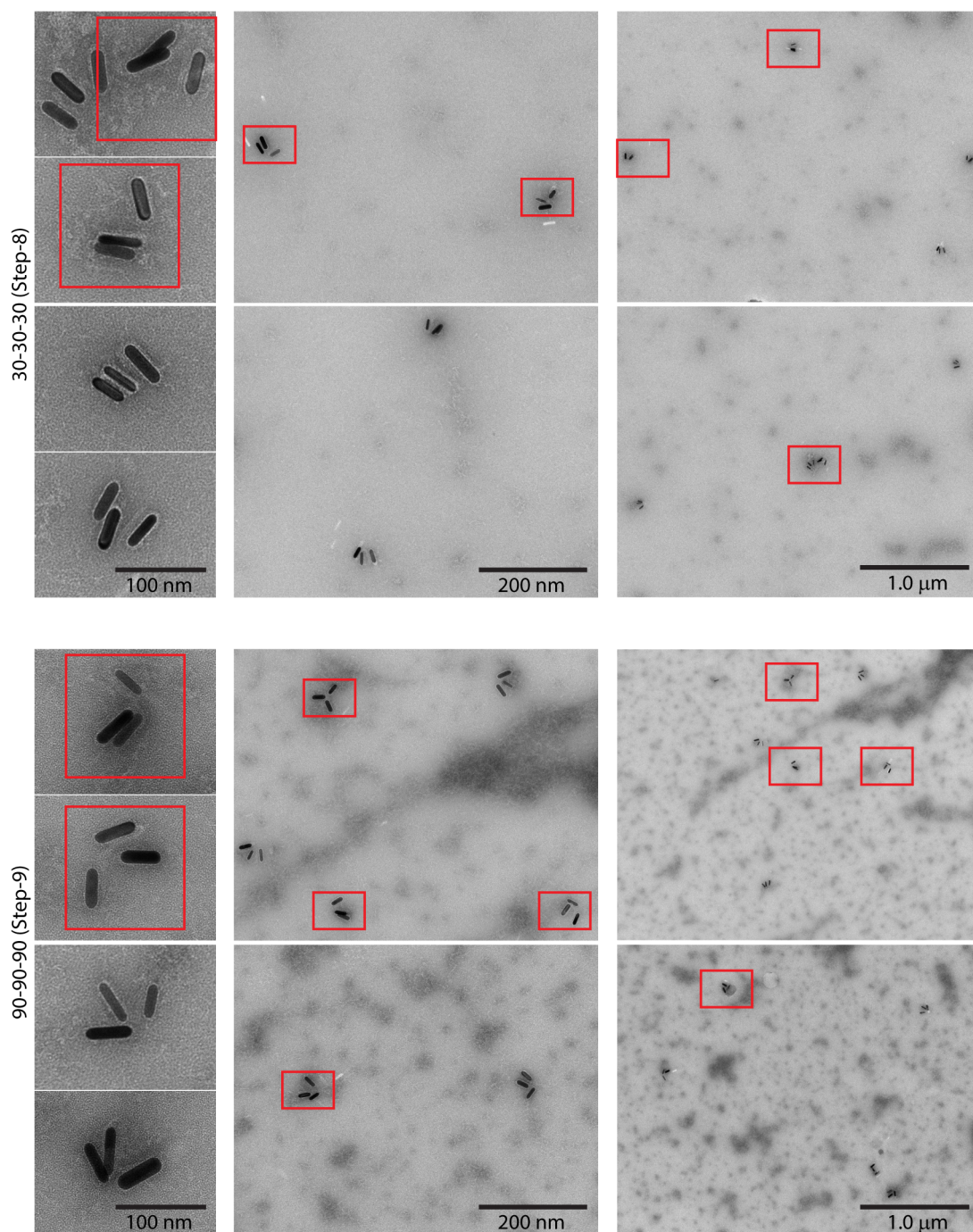
**Figure S7.** TEM images of AuNR conjugated 30-60-60, 60-60-90, 30-90-90 and 60-90-90 tripods.



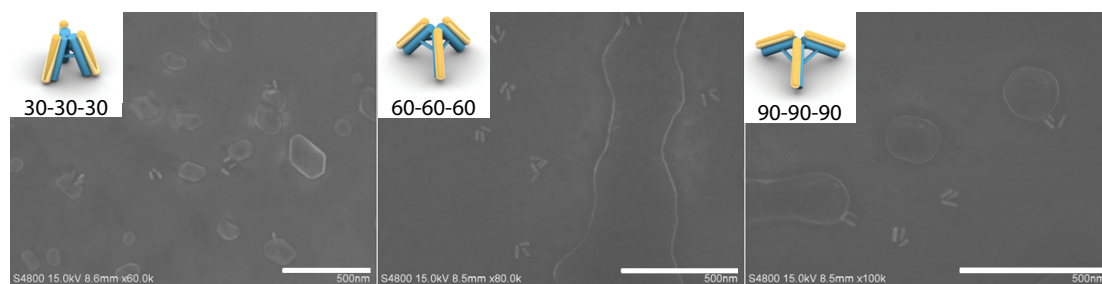
**Figure S8.** (a) Absorbance spectra of free DNA conjugated AuNR, and AuNR modified 30-30-30, 60-60-60, 90-90-90, 30-60-60, 60-60-90, 30-90-90 and 60-90-90 tripod with corresponding LSPR peak shifts. (b) Zoomed in spectra of (a) showing LSPR peak shifts of various tripod conformations compared to free DNA conjugated AuNR. (c) Absorbance spectra of free DNA conjugated AuNR, AuNR assembled 90-90-90 tripod, and 30-30-30 and 60-60-60 tripod converted from 90-90-90 tripod. (d) Zoomed in absorbance spectra of (c) showing shifts in absorbance maxima.



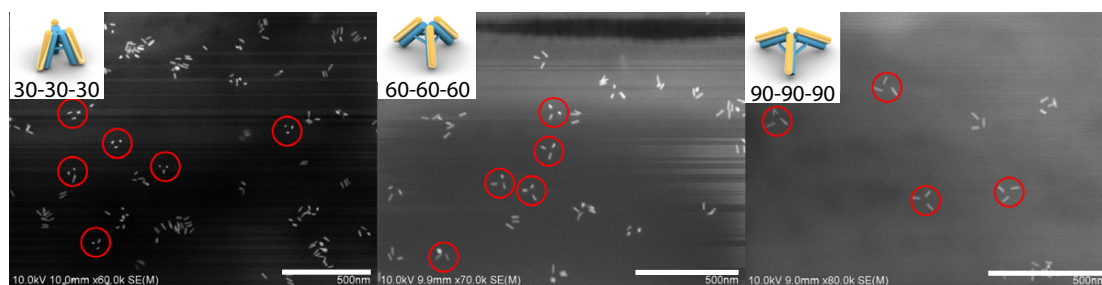
**Figure S9.** Absorbance spectra of each steps from reconfiguration cycle experiment (Figure 3f). Step-1 corresponds to 90-90-90 tripod (red sphere, Figure 3f).



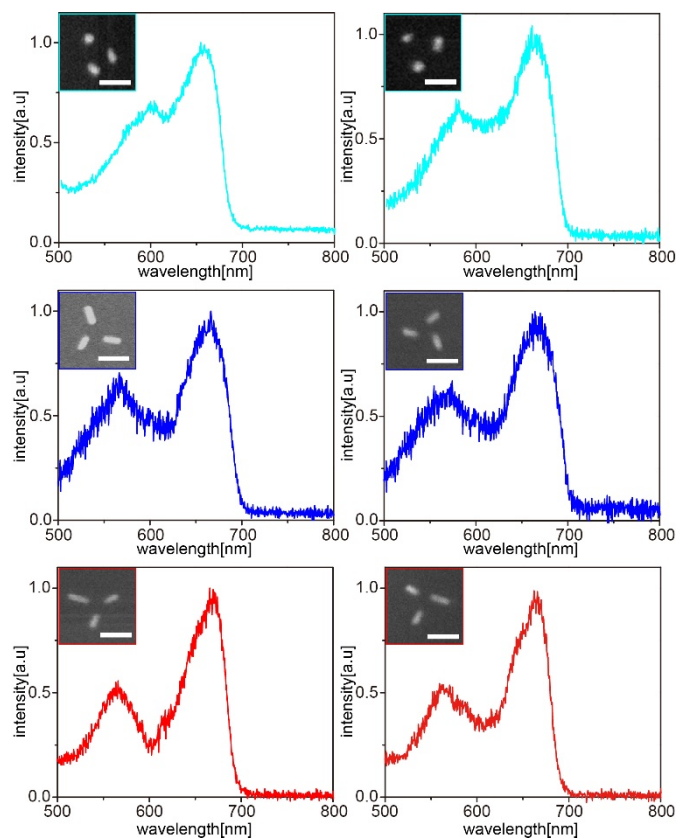
**Figure S10.** TEM images of the 30-30-30 AuNR-tripods from Step-8 and the 90-90-90 AuNR-tripods from Step-9 from reconfiguration cycle experiment (main text Figure 3f). Some of the partially transformed AuNR-tripods are highlighted with red boxes. Images of the 30-30-30 AuNR-tripods from Step-8 show that some angles are larger than  $30^\circ$ . Similarly, images the 90-90-90 AuNR-tripods from Step-9 show that some angles are smaller than  $90^\circ$ .



**Figure S11.** Additional SEM images of AuNR-tripod nanostructures without biotin modifications. Scale bar 500 nm.

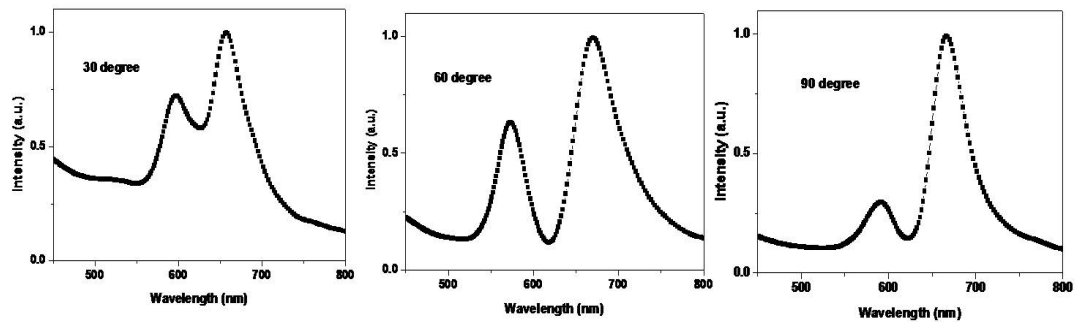


**Figure S12.** Additional SEM images of AuNR-tripod nanostructure with biotin modifications. The AuNR-tripods show stand-up configuration on streptavidin modified substrate due to the biotin-streptavidin interactions.

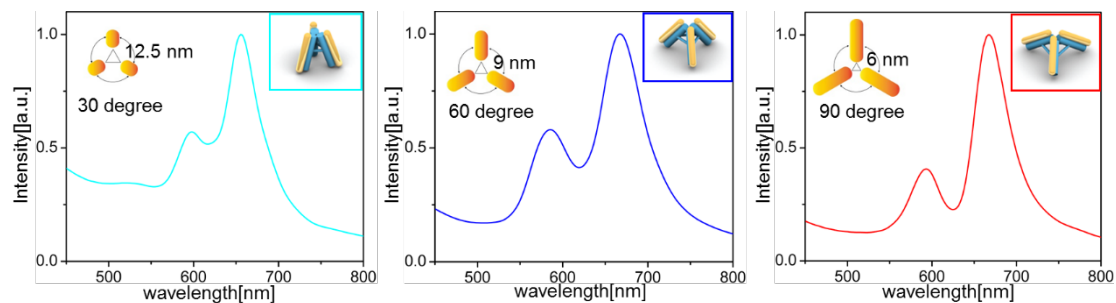


**Figure S13.** Additional light scattering spectra for individual three AuNRs arranged (top) in a 30-30-30 configuration, (middle) in a 60-60-60 configuration, (bottom) in a 90-90-90 configuration, all on silicon wafer and in air. Insets in

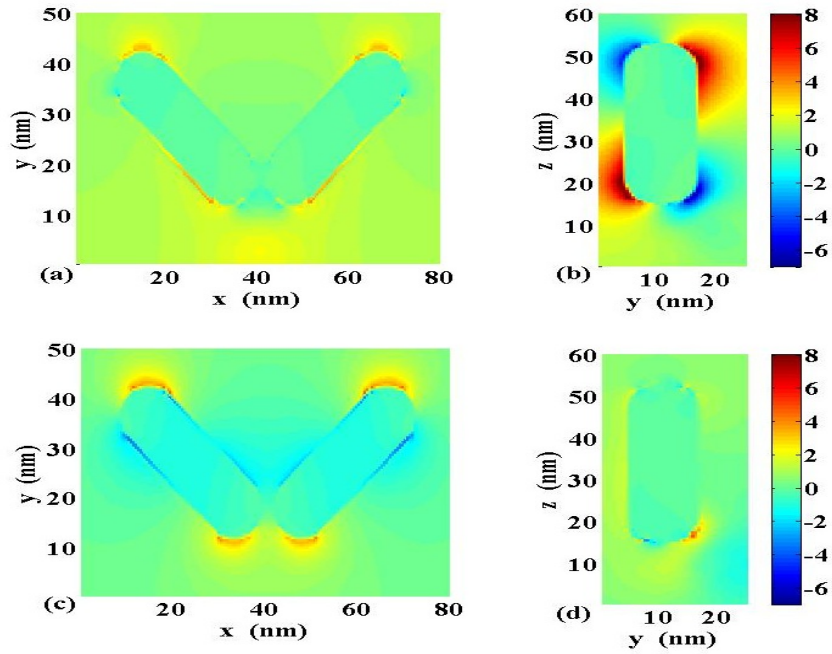
left-side show the SEM images of the particles giving rise to each scattering spectrum. Scale bar = 50 nm. Insets in right-side show the designated configuration for simulation, the size of AuNR is  $\sim 38\text{nm} \times 12\text{nm}$ .



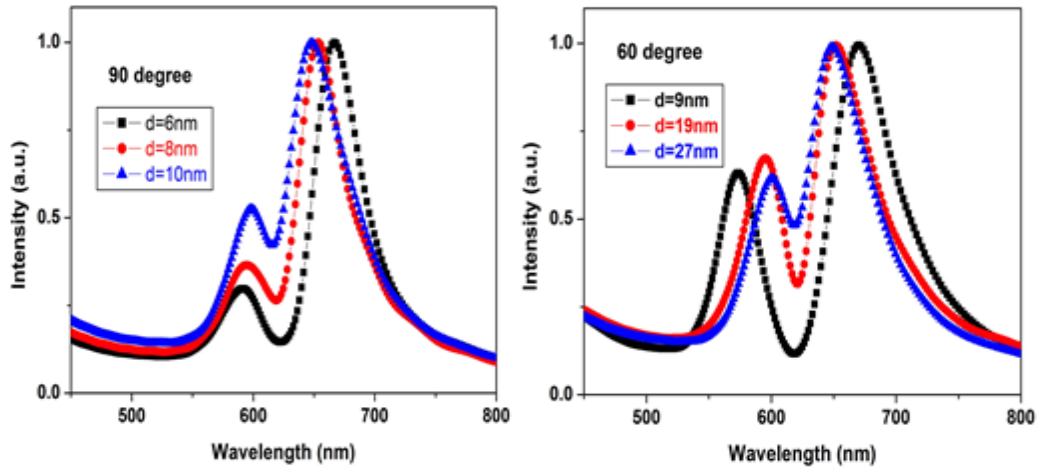
**Figure S14.** The scattering spectra of type C contribution for  $\theta=30^\circ$ ,  $60^\circ$ ,  $90^\circ$ .



**Figure S15.** The normalized scattering spectra of the TNRs for  $\theta=30^\circ$ ,  $60^\circ$ ,  $90^\circ$ . Notes on calculation details: The cone angle  $2\alpha$  is calculated by the formula  $\alpha = \arcsin(2 \sin(\theta/2) / \sqrt{3})$ . For the case of  $\theta=90^\circ$ , the cone angle is  $2 \times 54.7^\circ$ . Then the incident field angle should satisfy  $0 \leq \alpha < 35.3^\circ$  to ensure that the incident field polarization outside the core. The solid angle corresponding to the cone angle  $2\alpha$  is calculated by the formula  $\Omega = 2\pi(1 - \cos(\alpha))$ .



**Figure S16.** The near field distribution of  $\text{Re}[E_y]$  ((a),(b)) and  $\text{Im}[E_y]$  ((c),(d)) for the case of type C with  $\theta=90^\circ$ . The wavelength of the incident field is 620nm.



**Figure S17.** The scattering spectra of type C configurations for  $\theta=90^\circ$  ( $d=6\text{nm}$ ,  $8\text{nm}$ ,  $10\text{nm}$ ) and  $60^\circ$  ( $d=9\text{nm}$ ,  $19\text{nm}$ ,  $27\text{nm}$ ).



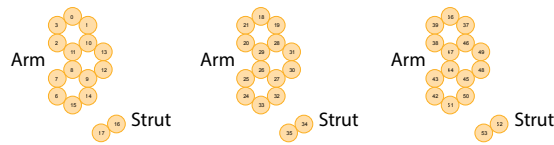
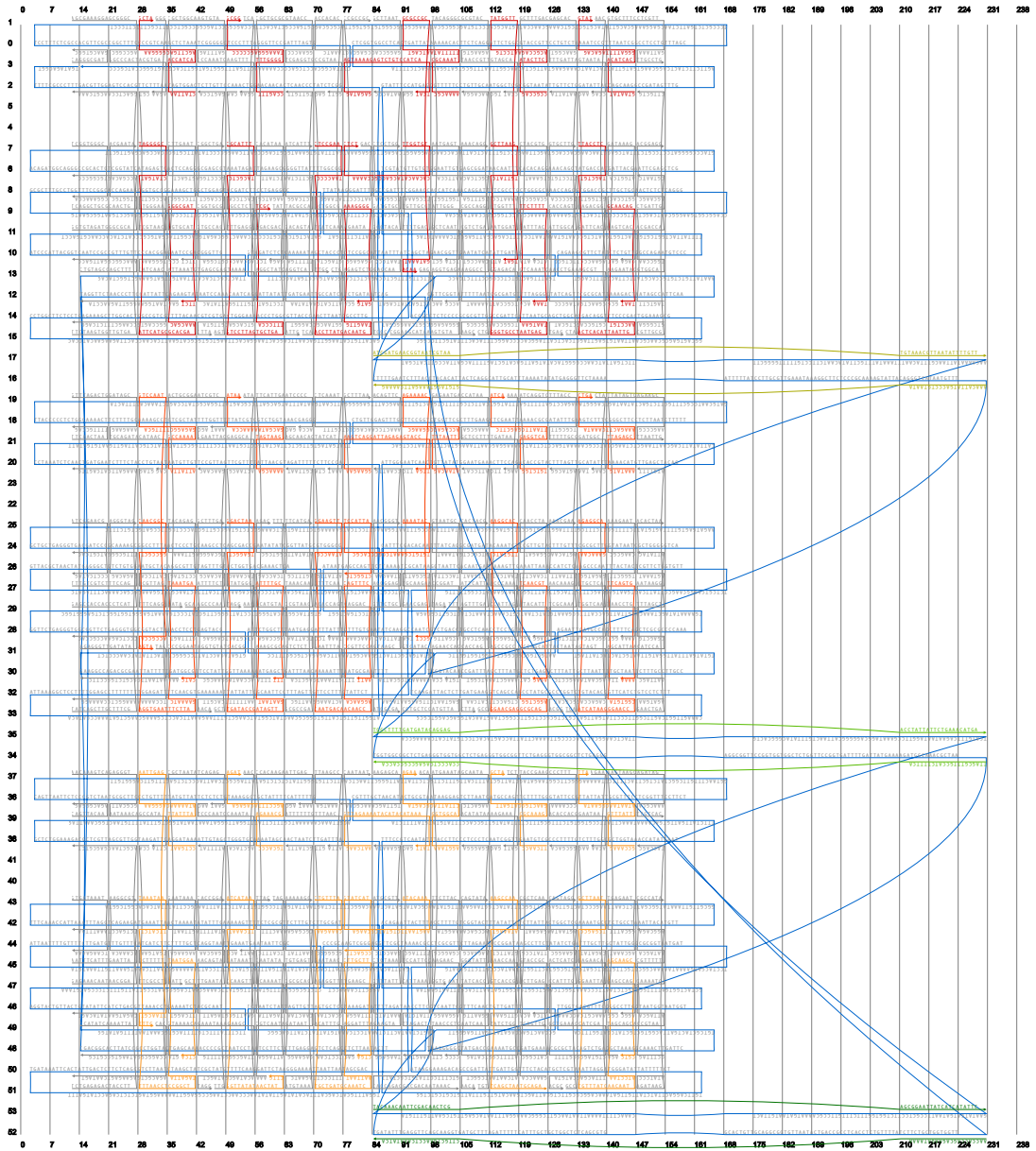


Figure S18. Strand diagram of 30-30-30 tripod.

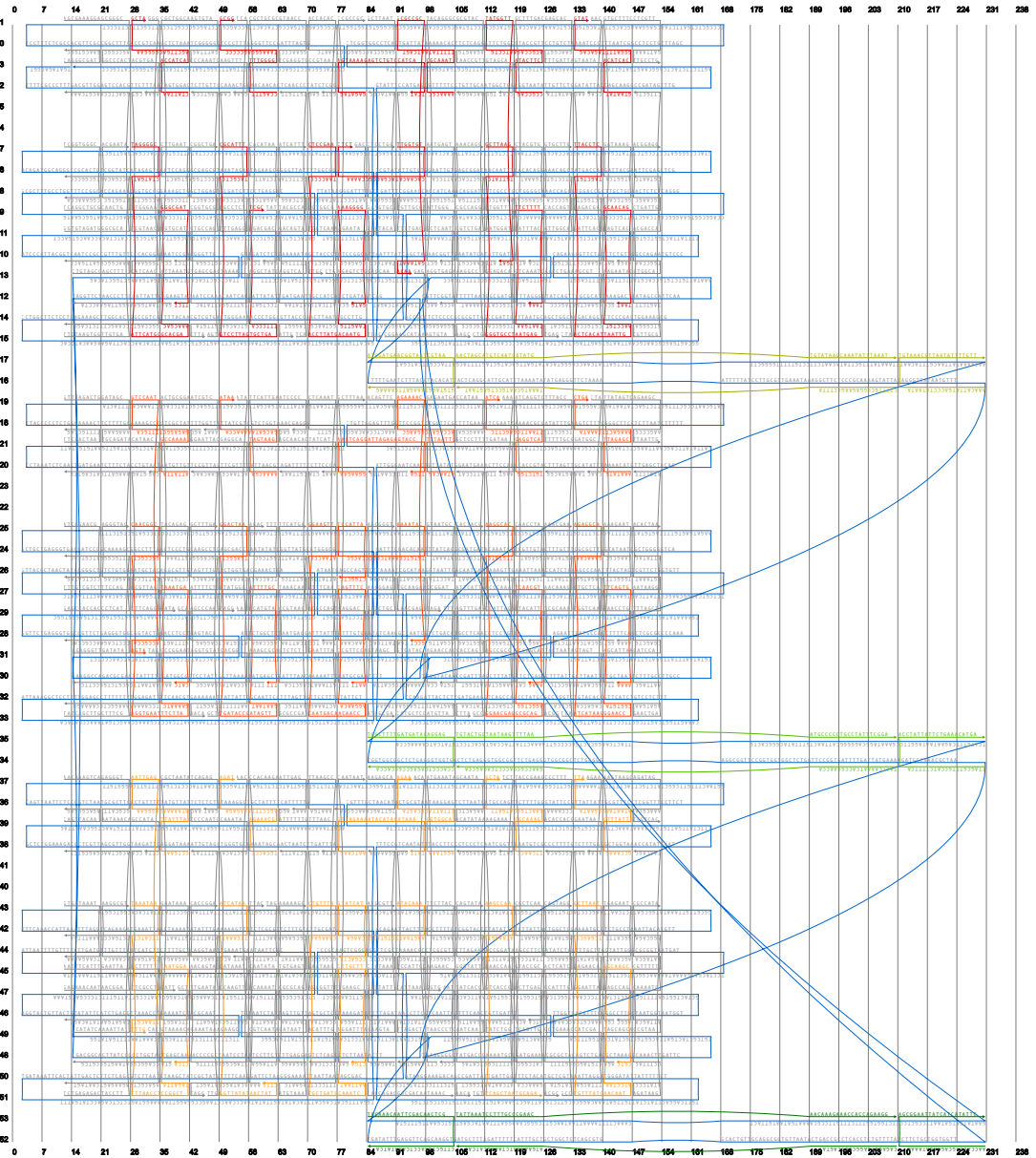


Figure S19. Strand diagram of 60-60-60 tripod.

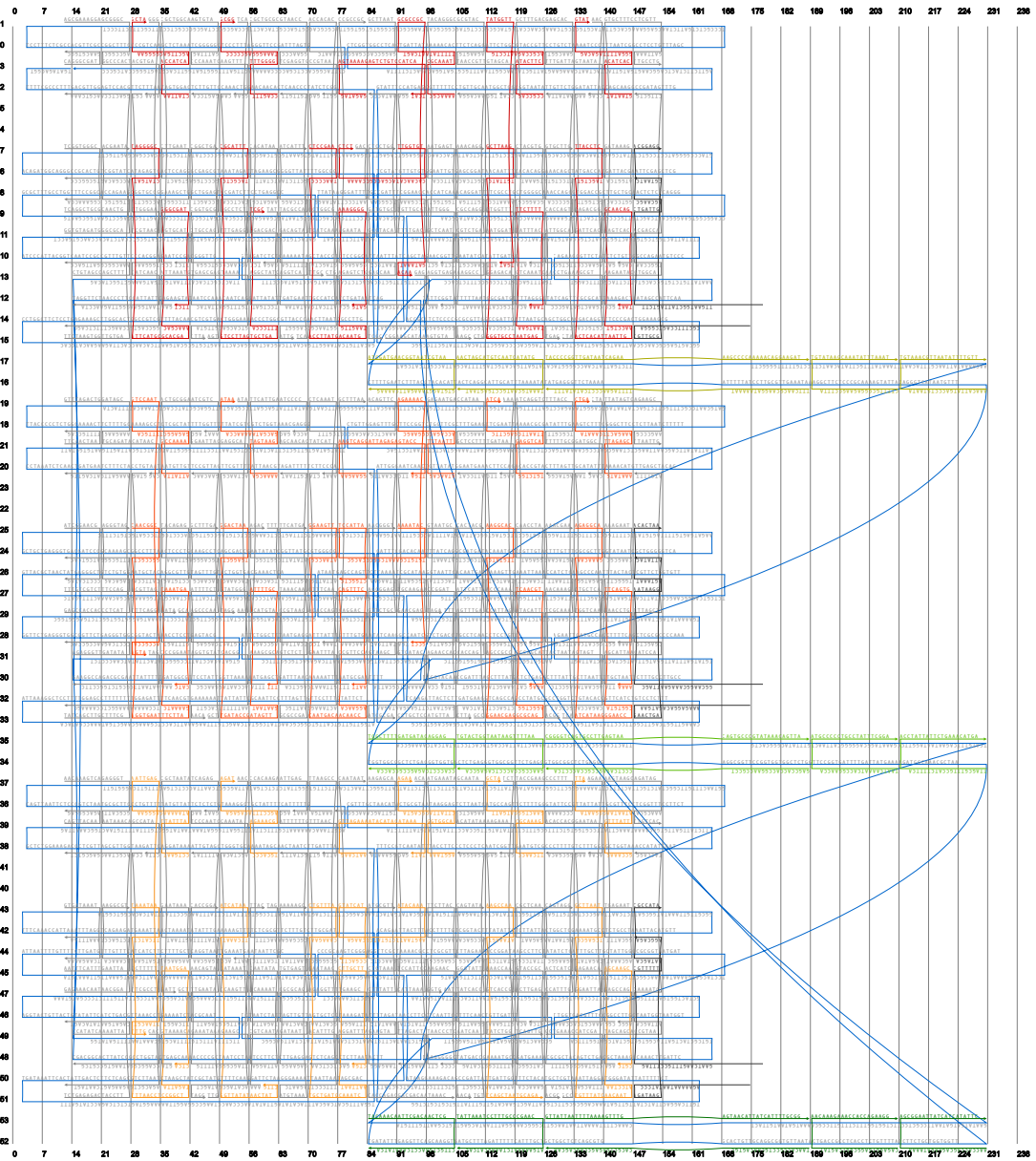


Figure S20. Strand diagram of 90-90-90 tripod.

## IV. DNA Sequences

core-1 GCAAGAGTCTGGAGCAATAATGCCGCTACAAATACCC  
core-2 GCGAAGAATACGTGGCATGGCCAAACGACCACTTGCTG  
core-3 GATGTGCTTTCCTCGTTGATTAAATTGCCTG  
core-4 ATAGCTTTGACGAGCACGAACGGTTTTGATTAGTAATACCAG  
core-5 AACAGGTGAGAAAGGCCAACCGTCGTCTGAAACAGGA  
core-6 ACAGTCAAATCAACCTGAAAGCGTCGAACTGATAGCCCCACCAGTTGCCCCAGTGCTTG  
core-7 TCAGCTTCTAATCTATTACATAAAAGATCGGGACGACGAGA  
core-8 CGGAAGCATAAAGTGTAGGGAGAGGCGGTCAAGGCGATTTCG  
core-9 CTGCGCTCACAATTCCAAATGAGTTGGTGGTGCTCAATTCTA  
core-10 CTATTCTGTGTGAAATCTACGTGGCAGGCGTATTTACATCA  
core-11 GAAATTAATGTGAGCGAGTAAATGTTTGGATTATACCGGTGCGGCCAGCTCGGCTGA  
core-12 GAGGCTGATTATCAGATGATGTGCTCGGCAACTCCTGG  
core-13 CTGATTCATCAATATAATACGCCACCTCAGGATCATTT  
core-14 AGTGCCCTGGAGTGACTCTTGAATTTCCGGCCGTGCATTTCT  
core-15 TTCTAAGTGGTTGTGAACCGACAGTGGGCC  
core-16 TTCCCATAGTAATCATAACCCAACTGACCTTCATCAACTTA  
core-17 CGTTCGTAACACCGCTTTTGGGAAGGCCAGTGCCAAGCTTCTCAGG  
core-18 CAATTTGAGGCACTCCAGGCCTCTGTCACGACGTTGTACTTA  
core-19 ACAGTTCAAACCTCCAACAGCCCTCGTTTCTCAAATAAA  
core-20 AAATTGAGGCAGCCCAAAGCTAAATCGGTATTCATTTGTGAATCACTACG  
core-21 CTAGGCATCAATTTCTGATCGCAAAAGTACGG  
core-22 AGCGGAATTACGAGGCAACTAACGTAGG  
core-23 GGGTGGGAACCATCAACTAATGGAAGGTTAGAACCTA  
core-24 TCAACCAACCCGTCGGACTGCCAGGAGTCCA  
core-25 GTAGTTTCAGGCATTCACACGTTAGTCCAAAAAAGGCTCCAAAAGG  
core-26 ATTTGAGACATCGCCATTAATAATCGCCAGGTGTTTGAAAACAGG  
core-27 GCTTACAGGGCGGTACAGAAGTGAACCGTTGTAGCACCAT  
core-28 CTAACACCAGTCAGGACAAACAAAGCTTTAA  
core-29 TCTCGCTGCGCGTAACCGGAACCCTCGAGGTGCCGTAAGGTT  
core-30 TTCGAACGAGTGCATTTGCAAGAACCGGATTGTA  
core-31 CCGCGCTGGCAAGTGTAGATTTAGCCCAAATCAAGTTGGAA  
core-32 CTGTAGCCAGCTTTAAACGGCGGATTGACC  
core-33 GGTGTAGATGGGCGCAGACTCCAACGTCAA  
core-34 TAATGGTCAAGAGATGGCTGCTCACAGACCAGGCGCATACGG  
core-35 TATTCCTGATAGGCTATCAGGTCATTTTGGAGACAGTATGTTGTT  
core-36 ATCGGAACGCCCTCATAG  
core-37 TCAGGCTGCGCAACTGCTGGTGCACGAATA  
core-38 ATGGTCAGATAAAGGTTCCACCAGTCACCAGA  
core-39 TTTTCACCAGCCATGTAAGCCAGATTCATTGAATCCCCACCA  
core-40 GACAAATCCCTTATAAGGGCTGGCGGTAACGCCAGGTTATTG  
core-41 TCGGTGGGCCGAAACCA  
core-42 AACGATTCACGCTGGTTGAGACGGCTGCCAGCTGCATTGAG

core-43 TGCAATGGATAAAAATCCGTGGTTTTTCGGCCAACGCGGAAGC  
 core-44 CAAGCGCAAAGAATTGGGCTTTAACCTGAAAA  
 core-45 ATTATGCTTTCAAATAAAGGAATGCGCGCCGAATATATTCGGTCGCAGAC  
 core-46 TTTGTCGTCTTTCCAGCAGACAGAGGGTAG  
 core-47 GAGCCACCACCTCATAAAGATTCATCAGTTG  
 core-48 AGCGAAAGGAGCGGGCAGCCGGCGCCACTACGTGAAGAA  
 core-49 AACTACAAACGTATGGGAATTTTTTCACGTAAACA  
 core-50 TTTACGATTGGAGAATGACCATAAAAAGCGAGCTCCTTTTGATAAAAAGT  
 core-51 TCCGAATTTAGTTAAATCAGCTCATAAACAAAGTTTCGTTTCATGA  
 core-52 TGAGTCATACAAAAATTCGCATTAAGCGGAGTTAAGAAAACGGGT  
 core-53 AGGATCAGGTCTTTTACCTCGCGTTTTTTTTGCGGATGGCCAAC  
 core-54 GGTTTATAGTCAGAAGCAAGCCCGTTAATTG  
 core-55 AGAGGGTTGATATACCCTCAGAACCGCCAC  
 core-56 TAGAATAATTCGCGTCTGGCCTTC  
 core-57 AAACCCGGAATAGGTGTATCACGGACCAATAGGAACGCATTTTCTTACAACGGCTTTGA  
 core-58 TTTAAAAGCGCAGTCTTTCATTAACCGTAACCGTTAAT  
 core-59 TTTCCGTTCCAGTAAGCTATTCACGTTCTGCCAATG  
 core-60 CCAAGAACCACCACCAGAGGTCAGAGTTTGAATTCCAT  
 core-61 CATGCCAGCATTACTAATAGTAGTAGCAATAAAGCCTCAACAAAGTTAATTAACGAA  
 core-62 TTAAGCATTAACATCCACGAGCTGTTTAGCTATGTTTT  
 core-63 GGATTTGACTCCGAAATGTTGCGTATTGGGACCG  
 core-64 GAGTCATCAAAGAATAGGAGAGGCCGCCGC  
 core-65 TATCAGCTTGCTTTTCGTTTTCGGGATCGTCAC  
 core-66 GCTTGCAGGGAGTTAAATACAGAGCCTGTAGGATA  
 core-67 CCTGCTCCATGTTAGAGTAATCTTGAAGAATAGAAATCGCGA  
 core-68 GCCATCGCCTGATAAATGTAATGCTACCTTATAGA  
 core-69 TCAAGTACAACGGAGATCAACCTATCAACTTATACATTCAGG  
 core-70 GCTTAATGAGGCCACCGAGCACTAAATCACACACGTAGCTATTGC  
 core-71 CAGGGCGATGAACGTGGC  
 core-72 AACAGAACCCTTCTGCCATTGGCAAATATTA  
 core-73 GTTTAGACTGGATAGCAAAGAAGTGCAGATACATAACCAG  
 core-74 ATAATCGTACTCAGGAGAGCCCAAGAACAAC  
 core-75 TTCAACTAATTTTGCCAG  
 core-76 AACAAAGTCAGAGGGTGCGCATTAATAAACAGCCATACTTA  
 core-77 AACGAATTGCGTAGATTTTGAATAATTTTAT  
 core-78 AAGAGCATGTTAGCAAACGCAA  
 core-79 ACATTATCTATTAGACTAAAGTAT  
 core-80 TCTGTTGAAATATCTGGATTTTCGGTCATAGGGGTATTAAGAACGCAGTATA  
 core-81 AGAAAAACGTCACCAAACCATTTGGGCGACA  
 core-82 AACTCCCAATCCAAATAAGCTACACCAAGTTAATT  
 core-83 AGCATTTTTTGTTTAACATTA  
 core-84 AAATGATTAAGCCGCACTAACATTTGTCTCAA  
 core-85 CAGTTACAAAGACGGGAG  
 core-86 CCATTCGCCTTGATGAACCTTTTTAGACGCTGAGAAGAGTCAATAGT

core-87 GACAGCAACACTATCATAAAT  
 core-88 GCAGTTTAGTACTGCGGAATCGTCAAAT  
 core-89 GTTAGAGGTTTTGAAGCTTTAGGACAATAAT  
 core-90 CCATATCAAAATTACAGATGAATATACAGT  
 core-91 GACCATTAAATTTTAGCTCCATTCCAAGAACCCCC  
 core-92 CACCCAGGCGGATAAGTGCCGTCG  
 core-93 GAGGTCACCGGTATTTCTAAACCAAATCAATAATCGGCTAACA  
 core-94 GCTGAAACCATCGACTGTAGCGCGTTTTACTCATCAGAAGGCCAGTAGG  
 core-95 AAGGGAATTACAGATATGAGAACATTTACGAGCATGTAACGC  
 core-96 GTAGTAAACAGAAATAAGAAGCTTAGCGGGGTTTTGAACAGTATGAGCAACACCGGA  
 core-97 GTGATAAATTCAGAAAA  
 core-98 GAGAAGGTGTGAGTGAATTATAAAAAGC  
 core-99 ACATACCGAAGCCCTTTCCCAAAAACACCAGGAATAAACAA  
 core-100 TAGTCATTTCCAAAAATTAGATTACCACAAGAATTGAGAAAT  
 core-101 GCCAGTAATAAGAGAATCGCTCAATTATCCGACTTGAGATCA  
 core-102 TGTACCGACAAAAGGTATTCTTACCAGGCGGGTG  
 core-103 TTGATATTTTAGTTAATGAATAAAAAGAAGAGATT  
 core-104 TCTGAGAGACTACCTTTCTGACCTAAATTTAA  
 core-105 GGCATTTTGAGAATAGCAAATGAGCCAGAGGC  
 core-106 ACCCATAAATACATAGCGATAGCTTAGG  
 core-107 GCGGGGCGAATAACTTTCCCTTAGAATCATGTAACCGGAGAAAACTTTTTAC  
 core-108 AATTTCAATTTGAATTAACAAACAAAGGCGT  
 core-109 GAGAAACAATAACGGAACGCTAACGAGCGTCT  
 core-110 GCTTTCAGGTCGCTAATATCAGAGGAAT  
 core-111 AATAGGAAGGATGAAATAGCAATAAAGACTCACATATAAAAAGAAAGATT  
 core-112 GTCTTTCCTTATGTGCGTATTCACCAGACGACGACAATAAAC  
 core-113 CGGAAGTAAGCAGATAGAAACGCATTGTCCAC  
 core-114 GACAGGATTTAGAAGTAAATATCCTTTATTTGAAATA  
 core-115 TAAGAGGCTGTAAAGAACCCTCATGCGTT  
 core-116 GAAACCAGTACCGCCATCGGCTCAGTTG  
 core-117 AGATAGCAGCACCGTAAATTAGCACAAAATCTACCAGC  
 core-118 AAACAATATAATTAGGACGTCAATAGATAATCAACTAACGCGCAGGCTATTT  
 core-119 CTTTAACCCCTCAATCAAGGAATTGTATCACCGGAGGGA

**Biotin binding handles**

Handle-bio-1 TCTCTCTCTCCGCTTTCCAGTCGGGACGTTGCGCGTAATC  
 Handle-bio-2 TCTCTCTCTCAGAAAAATAATATCCAGATAAGAGGCAGA  
 Handle-bio-3 TCTCTCTCTCTTGAATGGCTATTAGTCTTCTGATTGAGCAAGCACGGAGG  
 Handle-bio-4 TCTCTCTCTCAGAAATCAAGTTGCTTTAGCGTTTTTAATAGCACGCCATA  
 Handle-bio-5 TCTCTCTCTCAGCAAGGCAAAGAATTAGCAATAAGGAGTAAATACACTAA  
 Handle-bio-6 TCTCTCTCTCAAAGAGGACAGATGAAGAAGTGAATTATAC

**AuNR capturing strands**

handle1-1 AAAAAAAAAAAAAACCAGTTTTTTGGGGTAAAGGAGCCCCCGCGG  
 handle1-2 AAAAAAAAAAAAAATTACCTCTAGCTGTACTCACATTAATTGAACCTGTGCAACAGTAAT  
 handle1-3 AAAAAAAAAAAAAACGCATTTTACGCTCGTCTTAGTGTGATTTCCCATCGC

handle1-4 AAAAAAAAAAAAAATAGGGGCTATGATATTCATGCGCACGAAAACGACGGGCGATTCT  
 handle1-5 AAAAAAAAAAAAAAGTAATATACATCACGGGATTTTAGACAGGTAT  
 handle1-6 AAAAAAAAAAAAAACCGCCAGATACTTCACGCCAGAATCCTGTATGGTTTGAT  
 handle1-7 AAAAAAAAAAAAAAGCTTAAGTGTATCGGGTGCCTAATGAGTAATGAATCTTTTTTAAA  
 handle1-8 AAAAAAAAAAAAAAGAGATAGAGTAAAAGAGTCTGTCCATCATCAT  
 handle1-9 AAAAAAAAAAAAAAAACGCCGCAATTTTTTTATAATCAGTGCGCCGCGATAAAATACAA  
 handle1-10 AAAAAAAAAAAAAATGGTGTCCACAACATACGACCGCCAAAACCTCT  
 handle1-11 AAAAAAAAAAAAAACTCCGAATAACCCACCTTATGACAATGTAAGTTGAAAGGGGATG  
 handle1-12 AAAAAAAAAAAAAACTATTAAACCATCAAGCTTGACGGGGAAGCTA  
 handle2-1 TAATAATAATAATAGGAAGTTATAACCGCAATGACAACAACCAGGAACACAGTTTCAATT  
 handle2-2 TAATAATAATAATAAAAACGATAGTAAGGACGATAAAAACCAATAA  
 handle2-3 TAATAATAATAATAAGAGGCAGAAACAATCATAAGGAACCCGGTGTATTCAGTGAAAA  
 handle2-4 TAATAATAATAATAATTATTAGCCAAAAGAGAGGCTTTTGCAGTCCAATACCGCCAAGTA  
 handle2-5 TAATAATAATAATAAAGGCACCTGTATCGGAACGAGGCGCAGAGGCTGGTCAACGTAGAG  
 handle2-6 TAATAATAATAATAAACAAGTTTAAATTACCAGACCGGAAGCAGAAAACGCCT  
 handle2-7 TAATAATAATAATAAAAATACTGTGTGCAAAATCCGCCACGCTCCATTACTGGCTC  
 handle2-8 TAATAATAATAATATGTCTGGGAGGTCATTAATTCGAGCTTCATCA  
 handle2-9 TAATAATAATAATAAATAATGTTAGAGCAAAGACTTCAAATACTGA  
 handle2-10 TAATAATAATAATACAACGGCGCCGCTAGGTGAATTTCTTAGAAAACTAAATGACATC  
 handle2-11 TAATAATAATAATAGGAAGAAAAGTCAGGATTAGAGAGTACCTTGA  
 handle2-12 TAATAATAATAATAGGACTAATGAGGCTTGATACCGATAGTTGAATAATATTTTGCTTT  
 handle3-1 AGGAATAGTTATAATGCACCAGAAACGAGCCTTTACAGAGAAGAT  
 handle3-2 AGGAATAGTTATAACCTGAATTTATTAATAAAAACAGGGAAAATGAGTTAACGTTTTG  
 handle3-3 AGGAATAGTTATAAAGGTAAAGGTGGCATTATTACGCAGTAAGAA  
 handle3-4 AGGAATAGTTATAATFCAACCCGCAAAGGAACTGGCATGATFGCTA  
 handle3-5 AGGAATAGTTATAAAATCAAGGTTAGAAAATACATACATAAAATATT  
 handle3-6 AGGAATAGTTATAATAAATAATTCACTTTTAACTCCGGCTTAGATTATAATGGACTCA  
 handle3-7 AGGAATAGTTATAAATCATAATTCAAATGGTTATATAACTATCTTG  
 handle3-8 AGGAATAGTTATAAGCCAAAGGTTTATTATAATAACGGAATATTA  
 handle3-9 AGGAATAGTTATAAGCTTAATTCGAGCCTGTTTATCAACAATATCCTAAAGCAAGCCGTC  
 handle3-10 AGGAATAGTTATAAAAGCCAAATAAAGTTCAGCTAATGCAGA  
 handle3-11 AGGAATAGTTATAACTGTTTACAAAGAATGCTGATGCAAATCAATTAATCTTGCTTCTGA  
 handle3-12 AGGAATAGTTATAAATACAAAAGTAATTCGTATCGCAAGAGTATCATCCGACTT

**90-strut**

90-1-1 TGATGCCATATCTTAAACATTTATGACCCTGTAATATGTAACGTTAATATTTTGGTT  
 90-1-2 AGGTTACCTTGCTTATCGATGAACGGTAATCGTAAGTGTAGGTAAAGATTCAAAG  
 90-1-3 TATTCGCATGGTCCCTTTTGGCGGAGAAGCCTTTATGTATAAGCAAATATTTAAAT  
 90-1-4 GATTTTCAGAACTAACTAGCATGTCAATCATATGTAAATGCAATGCCTGAGTAAT  
 90-1-5 GCTGACTTATTTAATTTCAACGCAAGGATAAAAATTTTLAGAACCCCTCATATATTT  
 90-1-6 ATCTCGATGTAAAGTACCCCGGTTGATAATCAGAAAAGCCCCAAAACAGGAAGAT  
 90-2-1 TTAATGCCGTATGCTTAGCGTTTGCCATCTTTTCAACCTATFATTCGAAACATGA  
 90-2-2 TTGTATAGCCAACCTGGCTTTTGTATGATACAGGAGCCACCCCTCAGAGCCGCCACCA  
 90-2-3 GATTTCTACGCGTATAATCAAAATCACCGGAACCAATGCCCCCTGCCTATTTCCGGA  
 90-2-4 TTACCATTGGTGTGTACTGGTAATAAGTTTAAAGAACCGCCACCCCTCAGAGCCA

90-2-5 TGCTCGTAAGGTTGAGCCACCACCGGAACCGCTCCCTCAGAGCCGCCACCCTCA  
90-2-6 GTTATTTCGACTAGCCGGGGTCAGTGCCTTGAGTAACAGTGCCCGTATAAACAGTTA  
90-3-1 AGTTCCTTGAGGTAAACCACCAGCAGAAGATAAAAAGCGGAATTATCATCATATTC  
90-3-2 GACTTGCTATAAAGTACAAACAATTCGACAACCTCGCTTGCTGAACCTCAAATATCA  
90-3-3 CTTAAGCTGTCTCTCAGAGGTGAGGCGGTGAGTATAACAAAGAAACCACCAGAAGG  
90-3-4 AGATTGTCTCGTGTATTAAATCCTTTGCCCGAACATGAAAAATCTAAAGCATCAC  
90-3-5 TCTATGTAGCGTCATAACACCGCTGCAACAGTGCCACGCTGAGAGCCAGCAGCAA  
90-3-6 CTAGGATTTCCGCGGTTATTAATTTAAAAGTTTGAGTAACATTATCATTTTGCGG

**90-strut release**

R-90-1-1 AACAAAATATTAACGTTTACATATTACAGGGTCATAATGTTTAAAGATAGGCATCA  
R-90-1-2 CTTTTGAATCTTTACCTACACTTACGATTACCGTTTCATCGATAAGCAAGGTAACCT  
R-90-1-3 ATTTAAATATTTGCTTATACATAAAGGCTTCTCCCGCAAAGGGACCATGCGAATA  
R-90-1-4 ATTACTCAGGCATTGCATTTACATATGATTGACATGCTAGTTAGTTCTGGAAAATC  
R-90-1-5 AAATATATGAGGGTCTAAAAATTTTATCCTTGCGTTGAAATTAATAAGTCAGC  
R-90-1-6 ATCTTCCTGTTTTTGGGGCTTTTCTGATTATCAACCGGGTACTTTACATCGAGAT  
R-90-2-1 TCATGTTTCAGAATAATAGGTTGAAAAGATGGCAAACGCTAAGCATACGGCATTAA  
R-90-2-2 TGGTGGCGGCTCTGAGGGTGGCTCCTGTATCATCAAAGCCAGGTTGGCTATACAA  
R-90-2-3 TCCGAAAATAGGCAGGGGCATTGGTTCCGGTGATTTTGATTATACGCGTAGAAAATC  
R-90-2-4 TGGCTCTGAGGGTGGCGGTTCTTAAAACCTTATTACCAGTACACACCAATGGTAA  
R-90-2-5 TGAGGGTGGCGGCTCTGAGGGAGGCGGTTCCGGTGGTGGCTCAACCTTACGAGACA  
R-90-2-6 TAACTGTTTTATACGGGCACGTACTCAAGGCACGTACCCCGGCTAGTCGAATAAC  
R-90-3-1 GAATATGATGATAATTCGCTTTTTATCTTCTGCTGGTGGTTTACCTCAAGGAACT  
R-90-3-2 TGATATTTGAGGTTTCAGCAAGCGAGTTGTGCAATTGTTTGTACTTTATAGCAAGTC  
R-90-3-3 CCTTCTGGTGGTTTCTTTGTTATACTGACCGCTCACCTCTGAGAGACAGCTTAAG  
R-90-3-4 GTGATGCTTTAGATTTTTTCATGTTCCGGCAAAGGATTTAATAACACGAGACAATCT  
R-90-3-5 TTGCTGCTGGCTCTCAGCGTGGCACTGTTGCAGGCGGTGTTATGACGCTACATAGA  
R-90-3-6 CCGCAAATGATAATGTTACTCAAACCTTTTAAAATTAATAACCGGAAATCCTAG

**60-strut**

60-1-1 ATTAGTCTGCGCATAAACATTATGACCCTGTAATATGTAAACGTTAATATTTTGT  
60-1-2 TTTAGCTGCACGTAATCGATGAACGTAATCGTAAGTGTAGGTAAGATTCAAAAG  
60-1-3 ATTTATGCGTCACGCTTTTTGCGGGAGAAGCCTTTATAAATGCAATGCCTGAGTAAT  
60-1-4 ATGCTACTTGTGAAACTAGCATGTCAATCATATGTGTATAAGCAAATATTTAAAT  
60-2-1 CTAATGTCTGACGTTTAGCGTTTGCCATCTTTTCAACCTATTTCTGAAACATGA  
60-2-2 CTGCTTTGAACGCGTGGCTTTTGATGATACAGGAGCCACCCTCAGAGCCGCCACCA  
60-2-3 ACCTGAGTTTCTGTAAATCAAAATCACCGGAACCAGAACCGCCACCCTCAGAGCCA  
60-2-4 GTACTTGCTAAAAGTGTACTGGTAATAAGTTTTAAATGCCCCCTGCCTATTTGGA  
60-3-1 TTTACTGCGAGTCCAACCACCAGCAGAAGATAAAAAGCGGAATTATCATCATATTC  
60-3-2 TGCCAGTTTTATCTCTACAAACAATTCGACAACCTCGCTTGCTGAACCTCAAATATCA  
60-3-3 TCTACTGTAGCCACAGAGGTGAGGCGGTGAGTATATGAAAAATCTAAAGCATCAC  
60-3-4 CGTTCCTAAGTCGCTATTAATCCTTTGCCCGAACAAAGAAACCACCAGAAGG

**60-strut release**

R-60-1-1 AACAAAATATTAACGTTTACATATTACAGGGTCATAATGTTTATGCGCAGACTAAT  
R-60-1-2 CTTTTGAATCTTTACCTACACTTACGATTACCGTTTCATCGATTACGTGCAGCTAAA  
R-60-1-3 ATTACTCAGGCATTGCATTTATAAAGGCTTCTCCCGCAAAGACGTGACGCATAAT



R-60-1-4 ATTTAAATATTTGCTTATACACATATGATTGACATGCTAGTTTCGACAAGTAGCAT  
R-60-2-1 TCATGTTTCAGAATAATAGGTTGAAAAGATGGCAAACGCTAAACGTCAGACATTAG  
R-60-2-2 TGGTGGCGGCTCTGAGGGTGGCTCCTGTATCATCAAAGCCACGCGTTCAAAGCAG  
R-60-2-3 TGGCTCTGAGGGTGGCGGTTCTGGTTCGGTGATTTTGATTACAGGAAACTCAGGT  
R-60-2-4 TCCGAAATAGGCAGGGGCATTTAAACTTATTACCAGTACACTTTTAGCAAGTAC  
R-60-3-1 GAATATGATGATAATTCGCTTTTTTATCTTCTGCTGGTGGTTGGACTCGCAGTAAA  
R-60-3-2 TGATATTTGAGGTTTCAGCAAGCGAGTTGTGCAATTGTTTGTAGAGATAAACTGGCA  
R-60-3-3 GTGATGCTTTAGATTTTTTATATACTGACCGCCTCACCTCTGTGGGCTACAGTAGA  
R-60-3-4 CCTTCTGGTGGTTTCTTTGTGTTTCGGGCAAAGGATTTAATAGCGACTTAAGAACG

**30-strut**

30-1-1 TGATATCTGCGATTAAACATTATGACCCTGTAATAGTGTAGGTAAAGATTCAAAG  
30-1-2 CGTAATCTTGCTTTATCGATGAACGGTAATCGTAATGTAACGTTAATATTTTGT  
30-2-1 TGGCTTTCAAGTCTTAGCGTTTGGCATCTTTTACCACCCCTCAGAGCCGCCACCA  
30-2-2 CGATTTGCTAGCGTTGGCTTTTGTATGATACAGGAGACCTATTTCTGAAACATGA  
30-3-1 ATATCGTCGCTTACAACCACCAGCAGAAGATAAACTTGCTGAACCTCAAATATCA  
30-3-2 TTCATTTGACCTGGTACAAACAATTCGACAACCTCGAGCGGAATTATCATCATATTC

**30-strut release**

R-30-1-1 CTTTTGAATCTTTACCTACACTATTACAGGGTCATAATGTTTAAATCGCAGATATCA  
R-30-1-2 AACAAAATATTAACGTTTACATTACGATTACCGTTCATCGATAAAGCAAGATTACG  
R-30-2-1 TGGTGGCGGCTCTGAGGGTGGTAAAAGATGGCAAACGCTAAGGACTTGAAAGCCA  
R-30-2-2 TCATGTTTCAGAATAATAGGTTCTCCTGTATCATCAAAGCCAACGCTAGCAAATCG  
R-30-3-1 TGATATTTGAGGTTTCAGCAAGTTTTATCTTCTGCTGGTGGTTGTAGACGACGATAT  
R-30-3-2 GAATATGATGATAATTCGCTCGAGTTGTGCAATTGTTTGTACCAGGTCCAATGAA

**PAINT handle**

vertex CAAAATTAATTACATTTAACTTATCTACATA  
vertex CCTCAGAACCGCCACCCTCATTATCTACATA  
vertex AACAGTACCTTTTACATCGGTTATCTACATA  
vertex TTAGCGTAACGATCTAAAGTTTATCTACATA  
vertex CCTCAGCAGCGAAAGACAGCTTATCTACATA  
vertex GTAATGGGATAGGTCACGTTTATCTACATA  
vertex TGCCATCTGTAAGCAACTCGTTATCTACATA  
vertex GGCAAAGCGCCATTCGCCATTTATCTACATA  
vertex TGGTTTGAATACCGACCGTTTATCTACATA  
End TTAACACCAGAACGAGTCTTGCCCTGACGAGATTATCTACATA  
End TTAACATGTAATTTTCTGACATTATCTACATA  
End TTCCCTGAGAGAGTTGCCCTTCACCGCTGGTTATCTACATA  
End TTGGGACATTCAGACAATATTTTATCTACATA  
End TTCAAATATCGGCGTAATAAAATTTATCTACATA  
End TTTTTGGGGCGATAAATCATACTATCTACATA  
End TTAGAAGGAAACCGAGGCCGAACAAAGTTACCTTATCTACATA  
End TTACCATTACCTCAGTAGCGACATTATCTACATA  
End TTTGACCCCCAGCGCAACTTTGTTATCTACATA  
End TTATTCATATGGTTACCAGTAGCTTATCTACATA  
End TTGCTAAACAGGAGGCCAGAATCAGAGCGGGATTATCTACATA

End TTCTGTAGCTCAACATATTTTCATTATCTACATA  
End TTATCATTACCGCGCCCATTTTCATCGTAGGATTATCTACATA  
End TTAAAAAGATTAAGAGGAAAGCGGATTGCATCTTATCTACATA  
End TTCGAGCTCGAATTCTCACTGCCTTATCTACATA

## V. References

- S1. Jana, N. R.; Gearheart, L.; Murphy, C. J., Seed-Mediated Growth Approach for Shape-Controlled Synthesis of Spheroidal and Rod-Like Gold Nanoparticles Using a Surfactant Template. *Adv. Mater. (Weinheim, Ger.)* **2001**, *13*, 1389-1393.
- S2. Shi, D. W.; Song, C.; Jiang, Q.; Wang, Z. G.; Ding, B. Q., A Facile and Efficient Method to Modify Gold Nanorods with Thiolated DNA at a Low pH Value. *Chem. Commun. (Cambridge, U. K.)* **2013**, *49*, 2533-2535.
- S3. Liu, N.; Langguth, L.; Weiss, T.; Kastel, J.; Fleischhauer, M.; Pfau, T.; Giessen, H., Plasmonic Analogue of Electromagnetically Induced Transparency at the Drude Damping Limit. *Nat. Mater.* **2009**, *8*, 758-762.
- S4. Zhang, S.; Genov, D. A.; Wang, Y.; Liu, M.; Zhang, X., Plasmon-Induced Transparency in Metamaterials. *Phys. Rev. Lett.* **2008**, *101*, 047401.
- S5. Yun, B. F.; Hu, G. H.; Cong, J. W.; Cui, Y. P., Fano Resonances Induced by Strong Interactions between Dipole and Multipole Plasmons in T-Shaped Nanorod Dimer. *Plasmonics* **2014**, *9*, 691-698.