

Aerosolization of mycotoxins after growth of toxinogenic fungi on wallpaper

Brankica Aleksic^{1,2}, Marjorie Draghi³, Sebastien Ritoux³, Sylviane Bailly¹, Marlène Lacroix¹,
Isabelle P. Oswald¹, Jean-Denis Bailly^{1*}, Enric Robine³

¹Toxalim, Université de Toulouse, INRA, ENVT, INP-Purpan, UPS, F-31000 Toulouse, France.

²French Environment and Energy Management Agency, Angers, France

³Scientific and Technical Centre for Building, Airborne Pollutants and Bioaerosol Division, Marne-la-Vallée, France

*Corresponding author: jd.bailly@envt.fr

SUPPLEMENTARY DATA 1 –determination of air velocities applied on substrates

The air velocity applied on the substrate has been determined from experimental measures and numerical approach. This approach consisted in the simulation of the speed of the air near the surface of the substrate thanks to a Reynolds Stress Model. This model applies to aeraulic flows with fast direction changes, which is the case with the blowing system that was used in this experimentation.

The comparison of the results coming from the numerical approach and the experimental measures permitted to choose the best configuration of the model and then to determine for each air velocity applied by the blowing device, the corresponding speed of the air on the fungal structures. The experimental measures have been done with an air speed measurement system (sensor FA Höntzsch, Th-industrie, Sarrigne, France) that was placed in different places of the substrate (figure S1A).

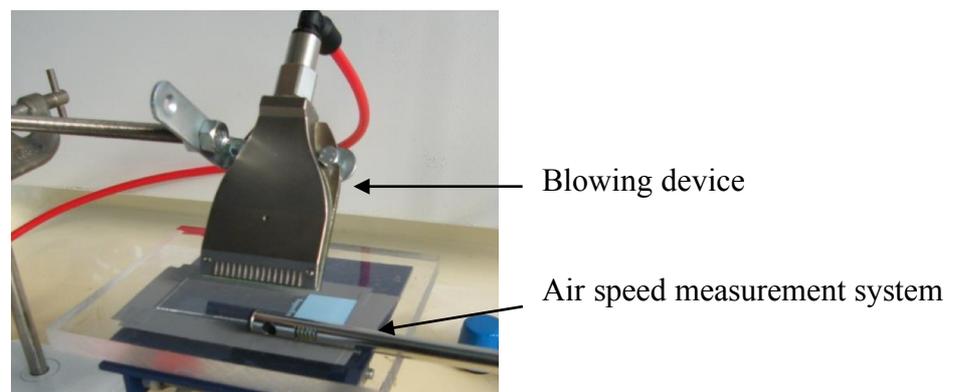


Figure S1A: measurement of air speed applied on the substrate

Obtained results are represented on figure S1B.

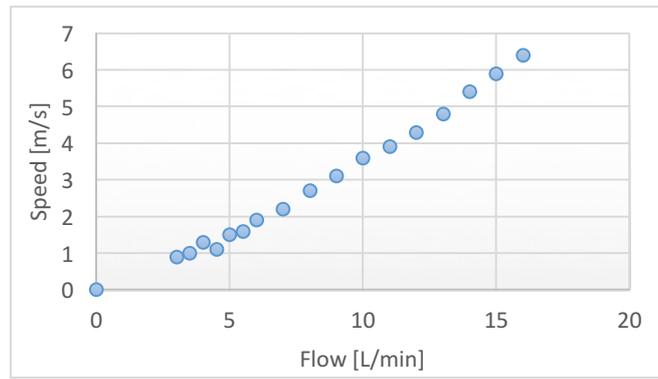


Figure S1B: Air velocity as a function of air flow applied by the blowing device

36
37
38
39

40 **Aerosolization of mycotoxins after growth of toxinogenic fungi on wallpaper**

41
42 Brankica Aleksic^{1,2}, Marjorie Draghi³, Sebastien Ritoux³, Sylviane Bailly¹, Marlène Lacroix¹,
43 Isabelle P. Oswald¹, Jean-Denis Bailly^{1*}, Enric Robine³

44
45 ¹Toxalim, Université de Toulouse, INRA, ENVT, INP-Purpan, UPS, F-31000 Toulouse, France.

46 ²French Environment and Energy Management Agency, Angers, France

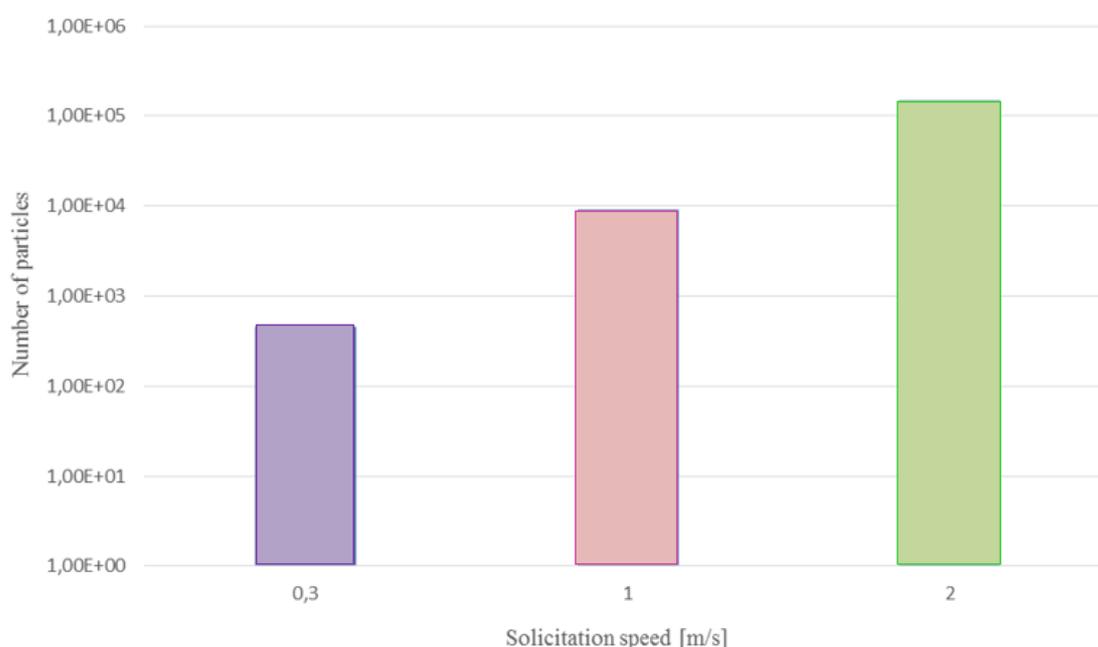
47 ³Scientific and Technical Centre for Building, Airborne Pollutants and Bioaerosol Division, Marne-la-Vallée,
48 France

49
50 *Corresponding author: jd.bailly@envt.fr

51 52 **SUPPLEMENTARY DATA 2 – Characterization of aerosolization conditions for** 53 ***Penicillium brevicompactum***

54
55 The propensity of fungal species to emit particles was studied by applying increasing speeds on
56 highly contaminated culture media. Results obtained for *Penicillium brevicompactum* are
57 shown on figure S2A. Below 0.3 m/s, the particle emission was sporadic with a maximum of
58 10 particles measured in the experimental volume (10.5 L). The number of particles increased
59 with air velocity (figure S2A).

60

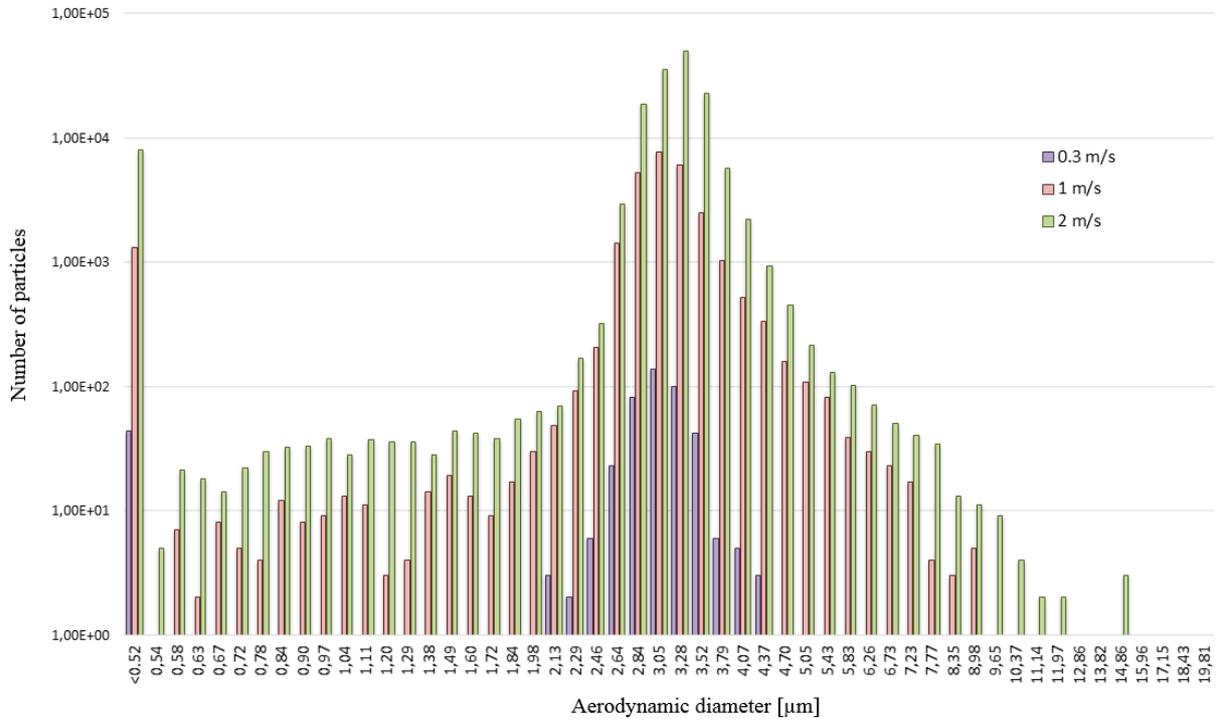


61

Figure S2A: Number of particles aerosolized from *P. brevicompactum* culture, by applying air stream 0.3, 1 and 2 m/s during 5 s.

62
63
64
65
66

The granulometric profiles of the emitted particles, for each air velocity is showed on figure S2B.



67

Figure S2B: granulometric profiles of particles aerosolized from *Penicillium brevicompactum* contaminated substrate following application of increasing air velocities for 5 s.

68

69 For the lowest speed, the particles are divided into two groups:

- 70 - The sub-micron particles of aerodynamic diameter less than 523 nm
- 71 - The super-micron particles of aerodynamic diameter ranging between 2 and 4.4 µm, which
- 72 may correspond to spores of *Penicillium brevicompactum*.

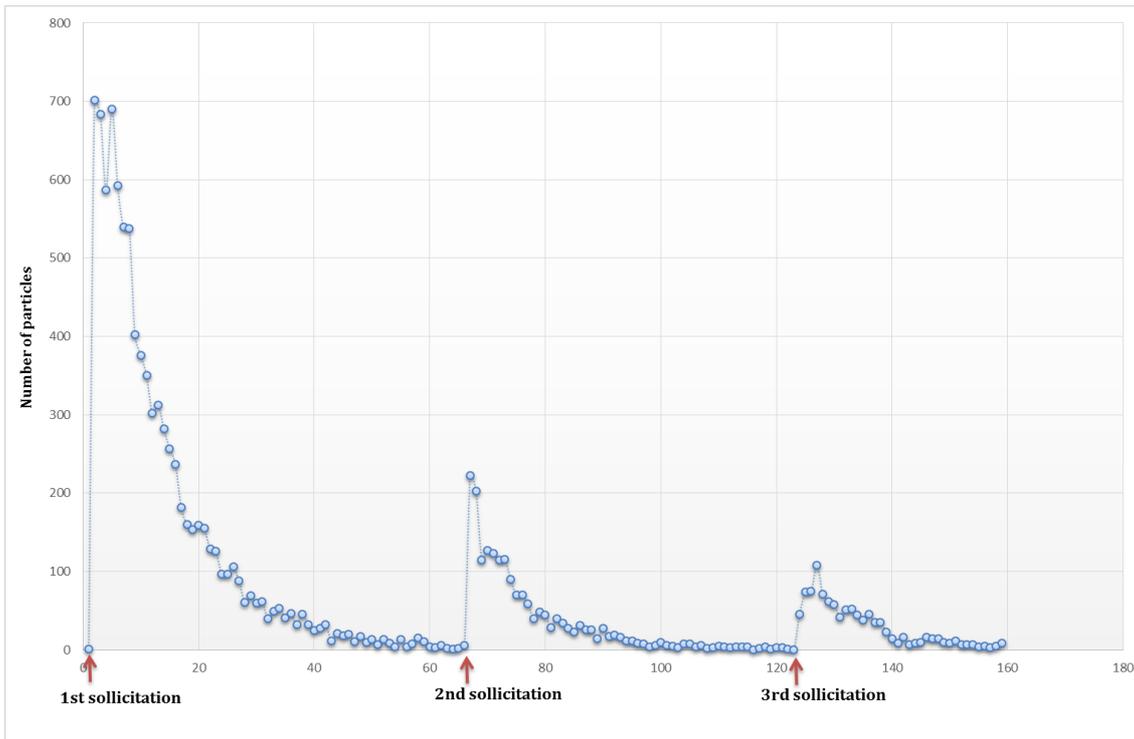
73 For blowing speed of 1 m/s, the particle size profiles is poly-dispersed with a main group made
74 of particles whose size is comprised between 2 and almost 10 µm. Once again, this group is
75 probably made of spores, groups of spore and mycelium fragment emitted from the substrate.
76 Increasing air velocity to 2 m/s did not modify the granulometric profile but allowed the
77 emission of few larger particles with size between 10 and 14 µm.

78 So it appeared that air velocity influence more the number of emitted particles rather than the
79 granulometric profile of bioaerosol.

80

81 To evaluate the impact a repeated air stresses on the substrate, three independent air stresses
82 were performed on same substrate. Figure S2C illustrates the evolution of the overall number

83 of particles emitted from a substrate contaminated with *Penicillium brevicompactum* following
84 three successive aeraulic stresses done with an air velocity of 1 m/s.
85



86

Figure S2C: Evolution of the total number of particles emitted from a substrate contaminated with *P. brevicompactum* following three successive air stresses of 5 s done at 1 m/s.

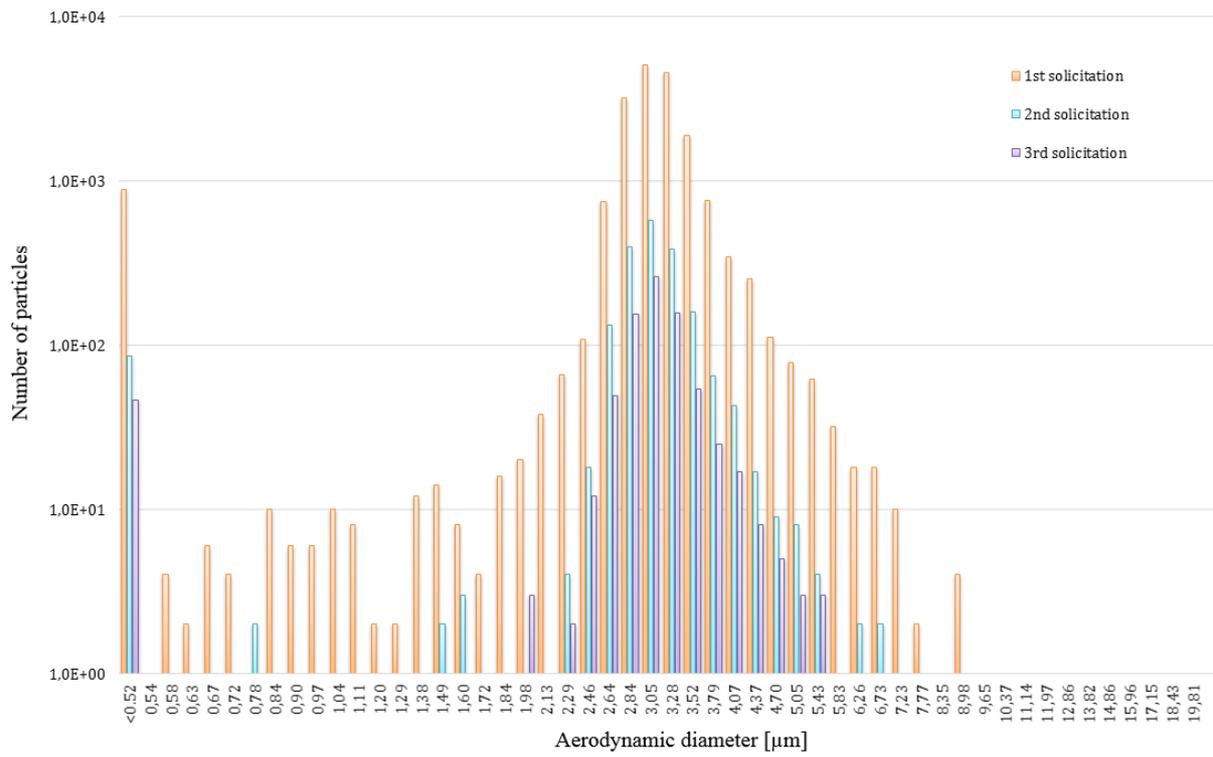
87

88 The first sollicitation allowed the aerosolization of a large amount of particles which represented
89 70% of the total number of emitted particles. The repetition of blowing procedure led to the
90 progressive exhaustion of the substrate. For aerosolization experiments, air stresses were
91 repeated until emitted particle number decreased to 1 particle/L

92 Figure S2D show granulometric profiles obtained during three consecutive air stresses done at
93 1 m/s on a substrate contaminated with *P. brevicompactum*. The repetition of blowing
94 procedure led to a progressive decrease of the number of particles in each of the measurement
95 channels, particles of size ranging between 2 and 5 μm being always predominant.

96

97



98

Figure S2D: Granulometric profiles of aerosols generated from *Penicillium brevicompactum* contaminated substrate during 3 successive air stresses done at 1 m/s for 5 s.

99