

Proteomic analysis of cerebrospinal fluid from children with central nervous system tumors identifies candidate proteins relating to tumor metastatic spread

SUPPLEMENTARY MATERIALS

Supplementary Data 1: List of proteins and peptides identified by mass spectrometry after the SEQUEST searcher for each sample

See Supplementary Data 1

Supplementary Data 2: Spectral count of the 558 identified proteins

See Supplementary Data 2

Supplementary Data 3: List of the 558 NCBI annotated proteins identified in both case and control groups after label free spectral counting (scaffold analysis) of the LC-MS/MS results

See Supplementary Data 3

Supplementary Data 4: Protein identifiers in column A were downloaded from CSF proteome database www.biosino.org/bodyfluid. This database provides International Protein Index (IPI) identifiers corresponding to 1286 unique proteins. As our proteins were identified by NCBI GI accession numbers, we used David gene ID conversion tool (<https://david.ncifcrf.gov>) to convert the NCBI GI accession numbers to IPI identifiers in order to compare our and Biosino lists (column B). We were able to confidently (status: identical) convert 504 of the 558 NCBI GI accession numbers to 1926 IPI identifiers. In column C the total list of IPI identifiers (3212, both databases); in column D 2878 unique identifiers. Thus 334 identifiers overlapped between the two lists, 1592 identifiers were unique (column E) for our study and 948 unique in Biosino database

See Supplementary Data 4

Supplementary Data 5: After the conversion from IPI identifiers to UniProt accession numbers we compared through PANTHER system Uniprot annotations included in Biosino database (column A) with those identified by us (Column B): 793 and 487 annotations, respectively (total annotations in column C, unique in column D). 212 uniprot accession number overlapped; 275 out of 487 annotations (56%) did not overlap Biosino database

See Supplementary Data 5

Supplementary Table 1: Cellular components according to the PANTHER database

See Supplementary File 1

Supplementary Table 2: Protein classes according to the PANTHER database

See Supplementary File 1

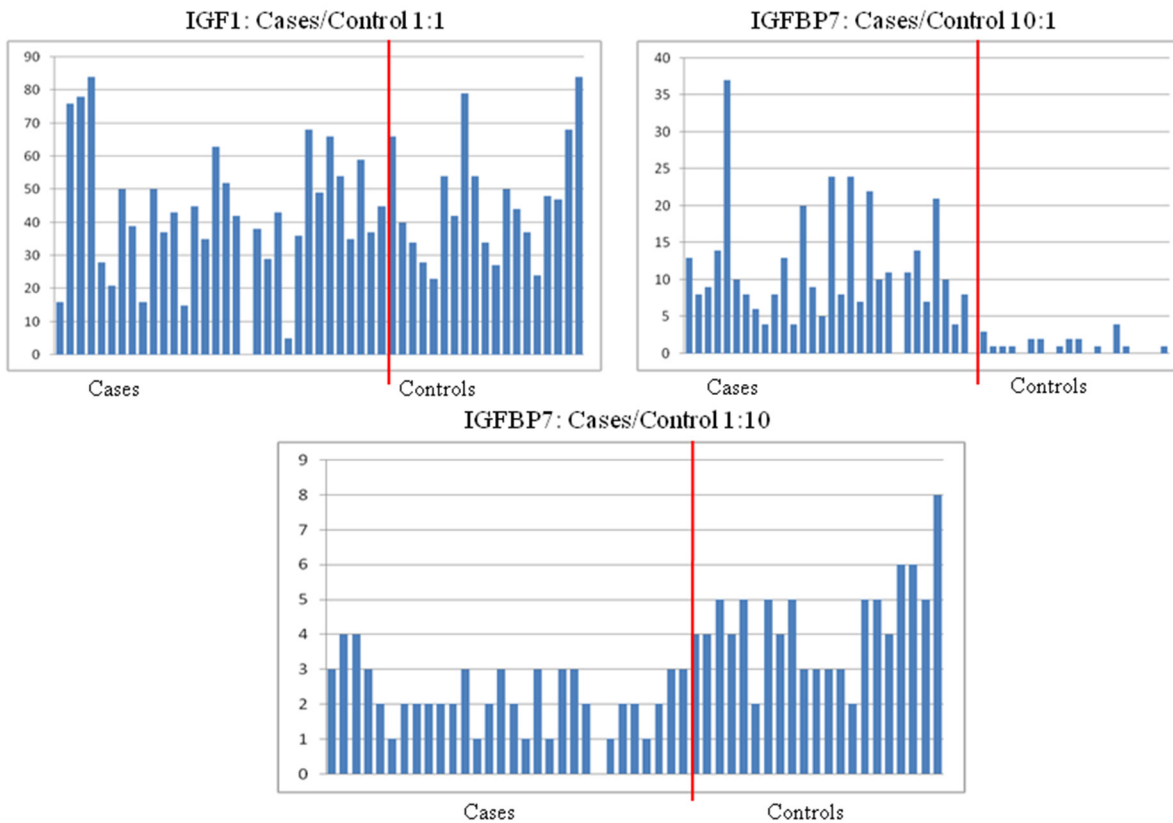
Supplementary Table 3: Molecular pathways according to the PANTHER database

See Supplementary File 1

Molecular pathways	Protein N°		
Alzheimer disease-amyloid secretase pathway (P00003)	2	Interleukin signaling pathway (P00036)	1
Alzheimer disease-presenilin pathway (P00004)	4	Ionotropic glutamate receptor pathway (P00037)	1
Alzheimer disease-presenilin pathway (P00004)	1	Metabotropic glutamate receptor group I pathway (P00041)	1
Angiogenesis (P00005)	2	Metabotropic glutamate receptor group II pathway (P00040)	1
Angiotensin II-stimulated signaling through G proteins and beta-arrestin (P05911)	2	Metabotropic glutamate receptor group II pathway (P00040)	1
Angiotensin II-stimulated signaling through G proteins and beta-arrestin (P05911)	1	Metabotropic glutamate receptor group III pathway (P00039)	1
Apoptosis signaling pathway (P00006)	1	Methylmalonyl pathway (P02755)	1
Axon guidance mediated by netrin (P00009)	2	Muscarinic acetylcholine receptor 1 and 3 signaling pathway (P00042)	1
Axon guidance mediated by semaphorins (P00007)	1	Nicotine pharmacodynamics pathway (P06587)	1
Axon guidance mediated by Slit/Robo (P00008)	2	Nicotinic acetylcholine receptor signaling pathway (P00044)	3
B cell activation (P00010)	2	Nicotinic acetylcholine receptor signaling pathway (P00044)	1
Blood coagulation (P00011)	19	Opioid prodynorphin pathway (P05916)	1
Blood coagulation (P00011)	14	Opioid prodynorphin pathway (P05916)	1
BMP signaling pathway-drosophila (P06211)	1	Opioid proenkephalin pathway (P05915)	1
Cadherin signaling pathway (P00012)	6	Opioid proenkephalin pathway (P05915)	1
Cadherin signaling pathway (P00012)	3	p38 MAPK pathway (P05918)	1
CCKR signaling map (P06959)	4	p53 pathway (P00059)	2
CCKR signaling map (P06959)	2	Parkinson disease (P00049)	3
Cytoskeletal regulation by Rho GTPase (P00016)	9	PDGF signaling pathway (P00047)	2
Cytoskeletal regulation by Rho GTPase (P00016)	1	Plasminogen activating cascade (P00050)	7
De novo purine biosynthesis (P02738)	1	Plasminogen activating cascade (P00050)	4
Dopamine receptor mediated signaling pathway (P05912)	1	Pyruvate metabolism (P02772)	1
DPP signaling pathway (P06213)	1	Ras Pathway (P04393)	2
DPP-SCW signaling pathway (P06212)	1	SCW signaling pathway (P06216)	1
EGF receptor signaling pathway (P00018)	4	T cell activation (P00053)	2
Endothelin signaling pathway (P00019)	1	TGF-beta signaling pathway (P00052)	2
Enkephalin release (P05913)	1	Toll receptor signaling pathway (P00054)	2
Enkephalin release (P05913)	1	Toll receptor signaling pathway (P00054)	1
FAS signaling pathway (P00020)	1	Transcription regulation by bZIP transcription factor (P00055)	1
FAS signaling pathway (P00020)	1	Vasopressin synthesis (P04395)	2
FGF signaling pathway (P00021)	3	Vasopressin synthesis (P04395)	1
General transcription by RNA polymerase I (P00022)	1	VEGF signaling pathway (P00056)	2
General transcription regulation (P00023)	1	Vitamin D metabolism and pathway (P04396)	1
Glycolysis (P00024)	3	Vitamin D metabolism and pathway (P04396)	1
Glycolysis (P00024)	1	Wnt signaling pathway (P00057)	8
Gonadotropin releasing hormone receptor pathway (P06664)	8	Wnt signaling pathway (P00057)	3
Gonadotropin releasing hormone receptor pathway (P06664)	2		
Huntington disease (P00029)	8		
Huntington disease (P00029)	3		
Huntington disease (P00029)	3		
Inflammation mediated by chemokine and cytokine signaling pathway (P00031)	13		
Inflammation mediated by chemokine and cytokine signaling pathway (P00031)	3		
Insulin/IGF pathway-mitogen activated protein kinase kinase/MAP kinase cascade (P00032)	1		
Integrin signalling pathway (P00034)	15		
Integrin signalling pathway (P00034)	3		
Interferon-gamma signaling pathway (P00035)	1		

Supplementary Figure 1: Overview of the distribution of molecular functions in control (red) and cases (black) and shared (highlighted in orange) according to the PANTHER database.

Analysis of the efficiency of the nanoparticles-ESI-MS/MS workflow



Supplementary Figure 2: Two chemokines IGFBP7 and IGF-1 were spiked in the CSF samples before nanoparticles incubation as control standards in order to evaluate the performance of nanoparticles harvesting. These chemokines were spiked in cases versus controls in a ratio of 10:1, 1:1, respectively. After Scaffold analysis, they were found to be differentially abundant with a ratio similar to the expected one, (actual ratio of 11.1 and 0.92) in brain tumor samples versus controls.