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## **Supplemental Information**

## Activation of Nrf2 Signaling Augments Vesicular

### Stomatitis Virus Oncolysis via Autophagy-Driven

### **Suppression of Antiviral Immunity**

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### **Supplementary Materials**

#### Supplemental Figure Legends

Figure S1. Sulforaphane potentiates VSV $\Delta$ 51 infectivity in various cancer cell lines. Human prostate DU145, human breast MDA-MB, and murine breast TS-A were pretreated for 24h with SFN and then challenged with VSV $\Delta$ 51. 24 hours later, viral infectivity was determined by flow cytometry. Data are the means  $\pm$  SEM from at least two experiments performed in duplicate or triplicate on each cell line.

**Figure S2. Sulforaphane treatment does not affect PC-3 viability** *in vitro.* Cytotoxic effect of SFN alone was tested in PC-3 cells by treating the cells with increasing concentrations of SFN for 24h and 48h. (a) SFN toxicity on PC-3 cells was visualized 48h after treatment by light microscopy. (b-c) The percentage of viable and apoptotic cells was assessed by flow cytometry using a 7AAD/annexin-V staining as previously described.

Figure S3. Sulforaphane does not increase VSV $\Delta$ 51 infectivity and oncolysis in non-cancerous human fibroblasts. (a-b) Human prostate PC-3 cancer cells and human MRC-5 normal fibroblasts were pre-treated with SFN (15 $\mu$ M) for 24h and were subsequently infected with VSV $\Delta$ 51-GFP (MOI 0.1). Infectivity (a) and oncolytic activity (b) were determined by flow cytometry at 48h post-infection. Data are the means ± SEM from two independent experiments performed in quadruplicate for MRC-5 and triplicate for PC-3.

Figure S4. Diethylmaleate stimulates Nrf2 promoter activity and increases VSVA51 infectivity in resistant prostate cancer cells. (a) HEK 293T cells were pre-treated for 24h with increasing doses of diethylmaleate (DEM) and ARE promoter activity was assessed using a luciferase assay. (b) PC-3 cells were pre-treated with sulforaphane (SFN) ( $20\mu$ M) or diethylmaleate ( $100\mu$ M) for 24h and were subsequently infected with VSVA51-GFP (MOI 0.1 or 1). Viral infectivity was determined by flow cytometry based on GFP expression. **Figure S5. SFN activates Nrf2 and increases VSV** $\Delta$ **51 infectivity independently of ROS. (a)** ROS generation in SFN-treated PC-3 cells was monitored by flow cytometry using CM-H2DCFDA (1µM) at 4 and 24h following treatment. H<sub>2</sub>O<sub>2</sub> (500µM) and pyocyanin (100µM) were used as positive internal controls of ROS accumulation. (b) ROS generation was also monitored 24h after treatment with H<sub>2</sub>O<sub>2</sub> (500µM) or pyocyanin (100µM) in presence or absence of L-NAC (2mM) and SFN (20µM). (c) HEK 293T cells were pre-treated with L-NAC (10mM) for 30 min which was subsequently washed away. Cells were then treated with SFN (10µM) for 18h. ARE promoter activity was assessed using a luciferase assay. (d) PC-3 cells were pre-treated with L-NAC (10mM) for 30 min. L-NAC was removed and cells stimulated or not with SFN (20µM) for 24h prior to challenge with VSV $\Delta$ 51 (MOI 1) for 24h. Viral infectivity was determined by flow cytometry based on GFP expression.

Figure S6. IRF3 does not bind to DNA following SFN stimulation in VSVA51-infected PC-3 cells. IRF3-nuclear binding activity was assessed using an IRF3-binding ELISA TransAM kit after treatment of PC-3 cells with SFN ( $20\mu$ M) for 24h and infection with VSVA51 (MOI 1) for 8 hrs. The data are expressed as relative arbitrary units. Data are the means from two independent experiments.

Figure S7. Abrogating autophagy reduces VSV $\Delta$ 51 infection and replication in A549 lung cancer cells. A549 cells were transfected with control or Atg7 siRNA and 48h later were challenged with VSV $\Delta$ 51-GFP (MOI 0.01) for an extra day. (a) VSV $\Delta$ 51 infection was determined by flow cytometry and pictures were taken using the ZOE fluorescent cell imager. Data are the means $\pm$  SEM from three independent experiments. (b) Viral replication was assessed by plaque assay. Data are the means $\pm$  SEM from one experiment performed in triplicate.





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