Supplementary Methods

Study Subjects

All 1112 Chinese ICP patients were of Han origin. A clinical diagnosis of chronic pancreatitis was based on two or more of the following criteria as previously described [Zou et al., 2016]: (i) presence of a typical history of recurrent pancreatitis; (ii) radiological findings such as pancreatic calcification and/or pancreatic irregularities revealed by endoscopic retrograde pancreatography or by magnetic resonance imaging of the pancreas; and (iii) pathological sonographic findings. ICP was defined as having neither reported family history of the disease nor recognized precipitating factors such as alcohol abuse, trauma, medication, infection and metabolic disorders.

Cell Culture and Transfection

HEK 293T cells (GenHunter, Nashville, TN) were cultured in 6-well tissue culture plates at a density of 1.5×10^6 cells per well, in Dulbecco's Modified Eagle Medium (DMEM) (Life Technologies, Grand Island, NY) supplemented with 10% fetal bovine serum, 4 mM glutamine and 1% penicillin/streptomycin at 37°C. Transfections were performed with 4 µg plasmid DNA and 10 µL Lipofectamine 2000 (Life Technologies) in 2 mL DMEM medium. After overnight incubation, the transfection medium was removed, cells were rinsed and covered with 2 mL OptiMEM reduced serum medium (Life Technologies). Conditioned medium was harvested 48 hours after the addition of OptiMEM.

Measurement of Procarboxypeptidase Secretion

Protein levels of CPA1 zymogen in the conditioned medium were measured by SDS-PAGE, Coomassie staining and densitometry. An aliquot (200 μ L) of the medium was precipitated with 10% trichloroacetic acid (final concentration), the precipitate was recovered by centrifugation, dissolved in 20 μ L Laemmli sample buffer containing 100 mM DTT, and heatdenatured at 95°C for 5 min. Samples were electrophoresed on 15% SDS-polyacrylamide mini gels and proteins were stained with Coomassie Blue (Brilliant Blue R-250). Quantitation of bands was carried out with the GelDocXR+ gel documentation system and Image Lab 3.0 software (Bio-Rad, Hercules, CA).

Measurement of CPA1 Activity

CPA1 zymogen was activated with 100 nM human cationic trypsin and 50 nM human CTRC, for 1 h at 37°C. The 40 μ L activation mixture contained 20 μ L conditioned medium, 0.1 M Tris-HCl (pH 8.0), 1 mM CaCl₂, and 0.05% Tween 20 (final concentrations). Carboxypeptidase activity was then measured by adding 50 μ L assay buffer (0.1 M Tris-HCl (pH 8.0), 1 mM CaCl₂, 0.05% Tween 20) and 10 μ L of 600 μ M N-[4-methoxyphenylazoformyl]-L-phenylalanine substrate to the activation mix (100 μ L final volume, 60 μ M final substrate concentration). The decrease in absorbance was followed at 350 nm for 2 min. Rates of substrate cleavage were calculated from fits to the initial linear portion of the curves and were expressed as percent of the wild-type rate.

Exon 1

Exon1_F	CGGTCCTGGGAGGGTTTAAAA
Exon1_R	TCTGTCTGTCTCCTCTACTGGTTG

Exon 2

Exon2_1_F	<u>ACAGCGTCTAGAAGTTTCTAAAGATGG</u>
Exon2_1_R	AGCTCCTTCACCTTCTGTACCT
Exon2_2_F	CAGGTGCTCCGAATCTCTGTAG
Exon2_2_R	AGAGTCAATTCCATGTCTCCTTGTTG

Exon 3

Exon3_1_F	CCAATTATATGAGAACTTCTGGCACCAA

Exon3_1_R CTCGATCATGGTCTCATAGCTGATG

- Exon3_2_F CCAGGCGGTCAAGATCTTTCTG
- Exon3_2_R TAGGTGGCGTAGTTAAAAGTGTCG
- Exon3_3_F <u>CAGGAGCAGATGTTCGCCTT</u>
- Exon3_3_R CAATGGAGCTACGACCACACA

Exon 4

Exon4_F	TCTGGGTGCCCAGAAGCTATTA		
Exon4_R	TGTTCCCTGGTACCATGATTTCC		

GGGCTGCTAAGGGAAGCA**TCTGGGTGCCCAGAAGCTATTA**AGGCCAGTGGTCTCTTCTTCACA CCTCAGATCTATGACTTCCTGGACCTGCTGGTGGCGGAGAACCCGCACCTTGTCAGCAAGATCC AGATTGGCAACACCTATGAAGGGCGTCCCATTTACGTGCTGAAGGTAACATCCACATGTGGACAT ACACAGGGGAGAATGGACCCACACGTGGCATCCGTGATGGGCGTGGGCTCTCCCGG<u>GGAAATC</u> <u>ATGGTACCAGGGAACA</u>CGCTGTTAAATGGACTCCCCATGCAGACATTTGGAAAGGCCTGAGTCT CCACCCTGGTCTTGGCGTGTGCACTCCTGCCCAT

Exon 5

Exon5_1_F	AAGCAGAGCCTCTACCTGAGAT
Exon5_1_R	GCCTTACCTTCTTTGCAAACCAGA
Exon5_2_F	CAGTAAGCGTCCAGCCATCT
Exon5 ² R	CCACAGCAATGGACACCTTTC

GCCCCACAAGCAGAGCCTCTACCTGAGATACACAGAGAGCAGGTGGTCTCTGGCCCAGGTCGG GGTCTCCTTCAGGGCAGCAAGATGAGGCCTCAGCTGTGAAATTGCCTCTGATCACTCCCCTGCC TCCTCTCCAGTTCAGCACGGGGGGCAGTAAGCGTCCAGCCATCTGGATCGACACGGGCATCCA TTCCCGGGAGTGGGTCACCCAGGCCAGTGGGGTCTGGTTTGCAAAGAAGGTAAGGCCGGGGA GGTGAGGAGGGCTCTCACCTGGTGGGGCCATTGGTGTCCAAGGCCCACAGAAGCCCGGGCCTCC CTTTGCCCATCCAGAAGCAGTGACCACAGAGGACATGGGGAAAGGTGTCCATTGCTGTGG GCAGATGCCTGGCCCAGCCTGCGCTGCCCCTCTGCTCC

Exon 6

Exon6_F	<u>GGGAAAGGTGTCCATTGCTGT</u>
Exon6_R	<u>GGGTGAACTACTTGTGCAGCTT</u>

ACAGAAGCCCGGGCCTCCCTTTGCCCATCCAGAAGCAGTGACCACAGAGGACATGGGGAAAGG <u>TGTCCATTGCTGT</u>GGCTTGGCAGATGCCTGGCCCAGCCTGCGCCCCTCTGCTCCTCTAACC CCCCAGATCACTCAAGACTACGGGCAGGATGCAGCTTTCACCGCCATTCTCGACACCTTGGACAT CTTCCTGGAGATCGTCACCAACCCTGATGGCTTTGCCTTCACGCACAGCACGGTACCGGCCTTCT CCTGTCCTTGGGGGAAGCAGGATGGGCCTCTGGCTTCT<u>AAGCTGCACAAGTAGTTCACCC</u>CTAA TCTCAAGCCCCAGAAGTCAAGGGAGGGGCAATCAGA

Exon 7

Exon7_F	<u>CCAATCAGGGCACTTGTGTTG</u>
Exon7_R	CATCCTTGCTCCCAAGACACT

Exon 8

Exon8_1_F	GGTTATAGAAGGCCTTTGGGCTT			
Exon8_1_R	GGTCCTTCACAAAGTCTACAATGGA			

- Exon8_2_F GGCAAGTTTGCCAATTCCGAAG
- Exon8_2_R_TGTGCCTTAAGCAGGTCTGATG

CCCCA**GGTTATAGAAGGCCTTTGGGCTT**TCCTGAATCCAGGGGTGGGAGTGAGCCCTTCCATAC CACCTCACCCCCAACTCCATGCAAAGAACTGGATTCCAGAAGCCACAGAAGCTGGAGGAGCCAC ACCGCCATGCCCTCTGTCCCCCACAGTGTCCGGAGCCAGCAGTAACCCCTGCTCGGAGACTTA CCAC**GGCAAGTTTGCCAATTCCGAAG**TGGAGGTCAAG**TCCATTGTAGACTTTGTGAAGGACC**AT GGGAACATCAAGGCCTTCATCTCCATCCACAGCTACTCCCAGCTCCTCATGTATCCCTATGGCTA CAAAACAGAACCAGTCCCTGACCAGGATGAGCTGGTAGGCACTGACCTCGGCTTGCCCCCTCGT CCCCAAGGTGGCTTCGGACAGGCCCAGGCTTTCCCCC**CATCAGACCTGCTTAAGGCACA**GACAC CTCCAGAGTACCTGACACCCTTCCTTCCCTGATGGCTTGGGAAGACCAGCGGGTGGACTAACCA TTTTTTCTGGGAAACTGAGGCACAGGAGTGATGGAGTCACTTCTGTAATGTGACAGAGCAAGGGG CAGGGCTAGACTTCGAT

Exon 9

Exon9_FGACTACCCTGGACATGCTGTTCExon9_RGATACTCCCTCGGTCAGGAAGA

Exon 10

Exon10_1_F	CTCTTTGGACCTCTTGGCTTAGA
Exon10_1_R	AGCTCGAAGGTGAAGGAGTACTT
Exon10_2_F	GAAGCACTATTGACTGGACCTACA
Exon10_2_R	<u>CTCAAACTTTATTTGGTTGCCTGGAT</u>

Supplementary Figure S1. Sequence information pertaining to the targeted sequencing of the *CPA1* gene. The sequences of the 16 primer pairs are first provided, followed by an illustration of their respective locations within the *CPA1* genomic sequence using different underlines, in the context of the 10 exons. Coding sequences are highlighted in blue. The *CPA1* genomic sequence was obtained from human GRCh37/hg19.

Exons 1-2

cccgctgaccctcaggccccgctggccccagatggtcgggggtggagctctggcttatctctccagctgcccagttccctgccactttatcatgga gggtgagaggtgtcagagctcAGAACTCCCACCCCAGCCTCCCCGTGGGACAGGACCCAGGTGCTGGG GGAGAACAGACCTCGGGAGCAGCCAGGAGTCCTCGGTCCTGGGAGGGTTTAAAAGCCAGGGGG CCGTCTCGACCTCAGTCTGACCTTCCCTCCCGGCAGCAGCATGCGGGGGTTGCTGGTGTTGAGT GTCCTGTTGGGGGGCTGTCTTTGGCAAGGAGGACTTTGTGGGGTAGGGATGTGGAGAGGGGG TACATGGCAACACGCCAGAGAGGGATGGCCTGGCACCACCTGGGACAGCTGGCAGATAAGCAA CCAGGGTTGGGAAGAGGTTGTGTCTCCCAAGACAGTCTCCTGAGCCCTGGAGAAACCTGAGCTC TCCAAACTGGGATTGAGATAGTAACAGCGTCTAGAAGTTTCTAAAGATGGGGAGAGAAGAGGTGA GGGAGCCCAGGCCTCTCCTTGTTGAGACCCTTGGTGCTGTCCCCTGCCCAGCCCAGAGGACGA GGCCCAGTCTGGGGATTCTGGGCTGCTGCCTCCCTCCTGCACCTGGGGAGTGCTTGTGGCCAG GGCAGGTGCAGCTTGGGTGCTCACTCCCCACTCCCCACTCTGCCCAGGCATCAGGTGCTCCGAAT CTCTGTAGCCGATGAGGCCCAGGTACAGAAGGTGAAGGAGCTGGAGGACCTGGAGCACCTGCA **G**GTCAGAAGAGGGGAGAAGGGCTCTCTGAGGCCCCAGGGTATCAGCTGGGGCCACCCCAGGTC CCCAGCGGCCAACTGTGCCTGGGCTGTCTCCACCCAACAAGGAGACATGGAATTGACTCTGTTA GGAAGCGACTTCAAGCCCCCAGCTCCAGAGCCTTGCTGCCCTTGAGCACCTGGGCAAGGGGGCC GGGCCCCTTTCCAGTTCCTCCCTCTGCCTTGCTGTCTCAAAGGCCCCTCACTCTGCTCCTGGGC CATTTC**CCTGACCACACTGGACTCTC**

Exons 3-4

CTTGCTGTCTCAAAGGCCCCTCACTCTGCTCCTGGGCCATTTCCCTGACCACACTGGACTCTCCA GTCCCCAGGGTTTCCCCGTATTTCTCTGGCCCTATTACCCCTGACCTATCCCCCAATTATATGAGAA CCAGCCCGCTGTGACCGTGCCGGCTCTTGTCCTCCCCAGCTGGACTTCTGGCGGGGGCCTGCC CACCCTGGCTCCCCATCGACGTCCGAGTGCCCTTCCCCAGCATCCAGGCGGTCAAGATCTTTC TGGAGTCCCACGGCATCAGCTATGAGACCATGATCGAGGACGTGCAGTCGCTGGACGAGG AGCAGGAGCAGATGTTCGCCTTCCGGTCCCGGGCGCGCTCCACCGACACTTTTAACTACGCCAC CTACCACACCCTGGAGGAGGTGAGGGCGCCCCTAGCGGCCGCTCCCTGCAGCCACCAGCTCTT CATCATGGCTGGTAGAACGCGGTAGGGCCAAGGCCAGGGCCAGCCTGGGTGTGCGCAGCGCCT GCTCTGTTTCCATGTGGCCTGTGTGGTCGTAGCTCCATTGCAGGGCTCGCAGCAGGCTGGGACG GTGGGGCTGCTAAGGGAAGCATCTGGGTGCCCAGAAGCTATTAAGGCCAGTGGTCTCTTCTTC ACACCTCAGATCTATGACTTCCTGGACCTGCTGGTGGCGGAGAACCCGCACCTTGTCAGCAAGA TCCAGATTGGCAACACCTATGAAGGGCGTCCCATTTACGTGCTGAAGGTAACATCCACATGTGGA CATACACAGGGGGAGAATGGACCCACACGTGGCATCCGTGATGGGCGTGGGCTCTCCCGGGGAA ATCATGGTACCAGGGAACACGCTGTTAAATGGACTCCCCATGCAGACATTTGGAAAGGCCTGAGT CTCCACCCTGGTCTTGGCGTGTGCACTCCTGCCCAT

Exons 5-6

<u>GCCCCACAAGCAGAGCCTCT</u>ACCTGAGATACACAGAGAGCAGGTGGTCTCTGGCCCAGGTCGG GGTCTCCTTCAGGGCAGCAAGATGAGGCCTCAGCTGTGAAATTGCCTCTGATCACTCCCCTGCC TCCTCTCCAGTTCAGCACGGGGGGGCAGTAAGCGTCCAGCCATCTGGATCGA

Exons 7-9

AGCAAGGATGGGATGGCCTCGAATGGCTCCTCACCACTGCTCTTGCTCCCCGCCTCTCCTGC CCCTGCAGTGAGGGGAGGGTTGGGGGGTGGCAGCTCTGCCTCTGAGGGCTCTTGGGGATGGAG TCTGAAATCCTACCTAGGGGGGCTTCTAGCTTAACCTCAAGTCCTCCCGCCTTGGCCACGTCTCTG AACCAGGCTGGGAGGGCAGTTCCCCCCAGGTTATAGAAGGCCTTTGGGCTTTCCTGAATCCAGGG GTGGGAGTGAGCCCTTCCATACCACCTCACCCCCAACTCCATGCAAAGAACTGGATTCCAGAAG CCACAGAAGCTGGAGGAGCCACACCGCCATGCCCTCTGTCCCCCACAGTGTCCGGAGCCAGC AGTAACCCCTGCTCGGAGACTTACCACGGCAAGTTTGCCAATTCCGAAGTGGAGGTCAAGTCCAT TCCTCATGTATCCCTATGGCTACAAAACAGAACCAGTCCCTGACCAGGATGAGCTGGTAGGCACT GACCTCGGCTTGCCCCCTCGTCCCCAAGGTGGCTTCGGACAGGCCCAGGCTTTCCCCCATCAGA CCAGCGGGTGGACTAACCATTTTTCTGGGAAACTGAGGCACAGGAGTGATGGAGTCACTTCTGT AATGTGACAGAGCAAGGGGCAGGGCTAGACTTCGATCTTAGGCACCACATGTGGAGGTGCATGT GCCTGAAGGGCAGGGAAGGCCAGCCGGACACCCTGAAGGGCCAGAAAACTGTGCTGACAGGCT CCCGGGCTGAGCTTCCAGGCTAAGCAGGCTCTGTGCTCCCTGCTGGAGGCCCACTTCTGCAGG GTGCTTCTCCTAGGCTCCCCTGCCAGCCCCCAGACTACCCTGGACATGCTGTTCCAGGAGCCTG GCCATGACAGGTGGCTTTGCTTGGTGTTTTGTCCAGGATCAGCTTTCCAAGGCTGCTGTGACAGC CCTGGCCTCTCTCTACGGGACCAAGTTCAACTATGGCAGCATCATCAAGGCAATTTGTAAGTGGC CGTAGGGTCTCTCTTGATGGGCCTGCGAGGAACATCTGCTGGCTCTTCCTGACCGAGGGAGTAT CCCTCATGGAAGGAAGTAGCCAAGGGCACCCAGATCTG

Exon 10

Supplementary Figure S2. Sequence information pertaining to the validation of targeted sequencing-derived *CPA1* variants by Sanger sequencing. Primer sequences used for amplifying the different exons are underlined. These primers were often used as sequencing primers. Further primers used for sequencing are indicated in red (forward) or green (reverse). Coding sequences are highlighted in blue. The *CPA1* genomic sequence was obtained from human GRCh37/hg19.



Supplementary Figure S3. Distribution of functionally impaired *CPA1* missense variants within the CPA1 amino acid sequence. Variants included those reported in this study as well as by Witt et al. [2013] irrespective of their occurrence in patients or controls. Variants found in different populations are indicated by different symbols, with their respective affected amino-acid positions highlighted in bold and blue. Short vertical bars in red indicate the exon boundaries. The long vertical line in green indicates the midpoint of the primary amino-acid sequence. The amino-acid sequence was derived from GenBank reference sequence NM_001868.3. Ger., E.r., Jap., Ind., and Chi. refer to the previously studied German, European replication, Japanese and Indian populations [Witt et al., 2013] and the currently studied Chinese population, respectively.

Controls in Director i optimions						
Population	Disease	Patients (%)	Controls (%)	OR	95% CI	P value
	subtype					
German	NACP	37/944 (3.92)	53/3,938 (1.35)	2.99	1.95-4.58	1.3×10 ⁻⁷
Eur. rep.	NACP	15/600 (2.50)	39/2432 (1.60)	1.57	0.86-2.87	0.14
Indian	NACP	10/230 (4.35)	0/264 (0)	Undefined	Undefined	0.002
Japanese	NACP	6/247 (2.43)	0/341 (0)	Undefined	Undefined	0.005
Chinese	ICP	20/1112 (1.80)	24/1580 (1.52)	1.19	0.65-2.16	0.57

Supplementary Table S1. All Rare *CPA1* Variants in Chronic Pancreatitis Patients and Controls in Different Populations

Data newly obtained in the Chinese population are highlighted in bold. Data from the other populations were derived from Witt et al. [2013].

Abbreviations: CI, confidence interval; Eur. rep., European replication; ICP, idiopathic chronic pancreatitis; NACP, nonalcoholic chronic pancreatitis; OR, odds ratio.