

A cross-scale trophic cascade from large predatory fish to algae in coastal ecosystems

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APPENDIX A

List of all fish species collected with gillnets (total 129) in 32 bays along the Swedish central Baltic Sea coast (see Methods). Total weight (Kg) is the sum of fish individual biomass estimated using species-specific length:weight conversion factors (in the Swedish national database for coastal fish, <http://www.slu.se/kul>). When we could not retrieve conversion factors, NA is reported for total weight. Total abundance (number of individuals) is given for all species.

Scientific name	Common name	Total weight (Kg)	Total abundance (n)
<i>Abramis brama</i>	Bream	32.33	57
<i>Alburnus alburnus</i>	Bleak	24.07	2946
<i>Ammodytes tobianus</i>	Lesser sandeel	NA	4
<i>Blicca bjoerkna</i>	White bream	10.86	283
<i>Carassius carassius</i>	Crucian carp	1.25	1
<i>Clupea harengus</i>	Baltic herring	7.17	233
<i>Coregonus lavaretus</i>	Whitefish	1.04	1
<i>Esox lucius</i>	Pike	70.86	40
<i>Gasterosteus aculeatus</i>	Three-spined stickleback	29.73	13854
<i>Gobius niger</i>	Black goby	0.05	7
<i>Gymnocephalus cernuus</i>	Ruffe	18.91	910
<i>Hyperoplus lanceolatus</i>	Great sandeel	0.12	9
<i>Leuciscus idus</i>	Ide	7.65	16
<i>Myoxocephalus scorpius</i>	Shorthorn sculpin	1.71	10
<i>Nerophis ophidion</i>	Straightnose pipefish	0.01	34
<i>Osmerus eperlanus</i>	Smelt	1.77	121
<i>Perca fluviatilis</i>	Perch	209.16	1847
<i>Phoxinus phoxinus</i>	Minnnow	NA	46

<i>Platichthys flesus</i>	Flounder	0.03	2
<i>Pomatoschistus minutus, P. microps</i>	Sand goby, common goby	0.04	55
<i>Pungitius pungitius</i>	Ninespined stickleback	0.32	447
<i>Rutilus rutilus</i>	Roach	117.96	3498
<i>Salmo trutta</i>	Trout	1.86	2
<i>Sander lucioperca</i>	Pike-perch	0.20	1
<i>Scardinius erythrophthalmus</i>	Rudd	2.28	25
<i>Spinachia spinachia</i>	Fifteen-spined stickleback	0.01	1
<i>Sprattus sprattus</i>	Sprat	0.27	27
<i>Syngnathus typhle</i>	Broadnosed pipefish	0.01	3
<i>Tinca tinca</i>	Tench	17.00	15
<i>Zoarces viviparus</i>	Eelpout	0.31	9

APPENDIX B

Predation intensity assay - Methods

To complement our correlative survey data with measurements of a key ecological process that links the mobile fish communities with the more sessile benthic community, we used a tethering assay [1] in 17 of the bays to estimate predation intensity on gammarids; one of the most efficient grazers on epiphytic macroalgae [2,3]. Tethering is a standard ecological method that can be successfully used to estimate relative differences in predation intensity across space and/or time [4]. Gammarids (2-9 mm long) collected in the field were individually glued to the tip of a 12 cm long transparent monofilament line (0.04 mm) attached to the end of a 30 cm-long acrylic transparent rod. Laboratory tests showed that gluing did not affect gammarid survival over 24h (unpublished data). In each bay, 10 to 14 rods were vertically inserted into the sediment 1-2 m apart, along a transect at 0.5-1.8 m depth. The tethered gammarids were deployed at 4-7 pm, and retrieved and scored for predation at 7-10 am the following morning. We estimated predation intensity per bay (PI) as:

$$PI = 100 * (N_E + N_{PE}) / (N_{TOT} - N_M - N_D)$$

where N_E and N_{PE} are the numbers of prey eaten and partially eaten, respectively, N_{TOT} is the total number of prey deployed in the bay and N_M and N_D are the numbers of prey moulted and dead.

Previous studies show that not only three-spined stickleback but also small perch feed on gammarids [5]. Furthermore, predation intensity may be influenced by vegetation density, which affects predator and prey abundances as well as predator-prey encounter rate [6,7]. To test effects of different predators, vegetation cover and their interaction on predation intensity across bays, we fitted multiple linear regressions with square-root transformed values of predation intensity as the response variable. As explanatory factors we included bay averages of total vegetation cover, biomass of either sticklebacks or perch (which were negatively collinear and could not be included in the same model), and their interaction with vegetation cover.

Predation intensity assay - Results

Bay-level stickleback biomass explained 41% of predation intensity on tethered gammarids ($r = -0.68$, $n = 17$, $P = 0.003$, adjusted $R^2 = 0.41$), strongly supporting the idea that stickleback predation contributes to control grazer densities in shallow coastal bays. Bay-level perch biomass and vegetation cover did not explain predation intensity, nor did the interaction between vegetation cover and either perch or stickleback biomass.

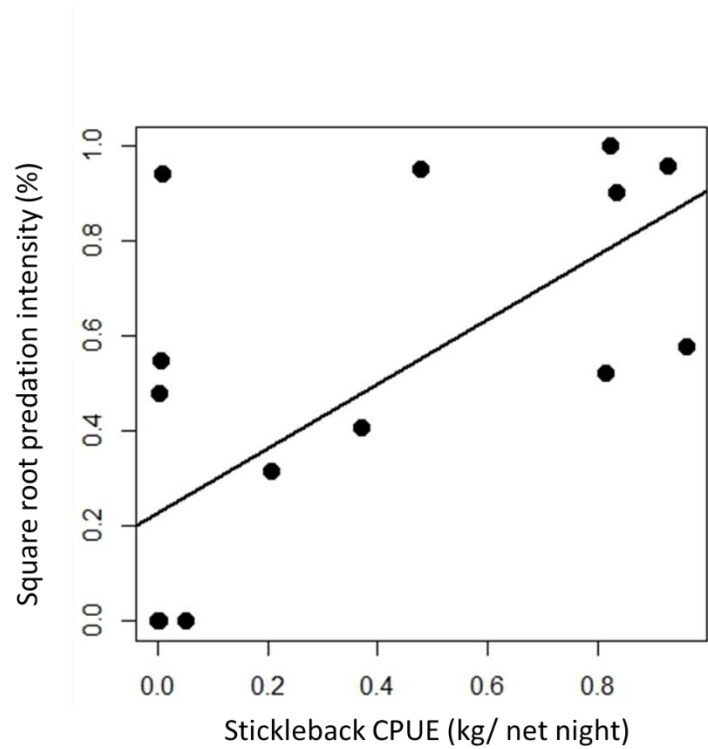


Fig. B1. Scatter plot of bay-level stickleback biomass and predation intensity on tethered gammarids and regression line ($r = 0.68$, $P = 0.003$, $n = 17$, adjusted $R^2 = 0.41$).

1. Duffy, J. E., Ziegler, S. L., Campbell, J. E., Bippus, P. M. & Lefcheck, J. S. 2015 Squidpops: a simple tool to crowdsource a global map of marine predation intensity. *PLoS One* 10, e0142994. (doi:10.1371/journal.pone.0142994)
2. Moksnes, P. O., Gullström, M., Tryman, K. & Baden, S. 2008 Trophic cascades in a temperate seagrass community. *Oikos* 117, 763–777. (doi:10.1111/j.0030-1299.2008.16521.x)
3. Sieben, K., Rippen, A. D. & Eriksson, B. K. 2011 Cascading effects from predator removal depend on resource availability in a benthic food web. *Mar. Biol.* 158, 391–400. (doi:10.1007/s00227-010-1567-5)
4. Heck, K. L. & Thoman, T. A. 1981 Experiments on predator-prey interactions in vegetated aquatic habitats. *J. Exp. Mar. Bio. Ecol.* 53, 125–134. (doi:10.1016/0022-0981(81)90014-9)
5. Reiss, K., Herriot, M. & Eriksson, B. 2014 Multiple fish predators: effects of identity, density, and nutrients on lower trophic levels. *Mar. Ecol. Prog. Ser.* 497, 1–12. (doi:10.3354/meps10622)
6. Eklöv, P. 1997 Effects of habitat complexity and prey abundance on the spatial and temporal distributions of perch (*Perca fluviatilis*) and pike (*Esox lucius*). *Can. J. Fish. Aquat. Sci.* 54, 1520–1531. (doi:10.1139/cjfas-54-7-1520)
7. Scheinin, M., Scyphers, S. B., Kauppi, L., Heck, K. L. & Mattila, J. 2012 The relationship between vegetation density and its protective value depends on the densities and traits of prey and predators. *Oikos* 121, 1093–1102. (doi:10.1111/j.1600-0706.2011.19941.x)

APPENDIX C

List of species included as epiphytic algae in the analyses.

Chlorophyta
<i>Cladophora</i> spp. ^a
<i>Mougeotia</i>
Unidentified uniseriate spp. ^b
Cyanophyceae
Unidentified uniseriate spp. ^c
Phaeophyceae
<i>Dictyosiphon foeniculaceus</i>
<i>Ectocarpus siliculosus</i>
<i>Elachista fucicola</i>
<i>Pylaiella littoralis</i>
Rhodophyta

<i>Ceramium tenuicorne</i>
<i>Polysiphonia fucoides</i>

^a *Cladophora glomerata*, *C. rupestris* and *C. fracta*, ^b *Ulothrix* spp., *Urospora* spp. and *Spirogyra* spp., ^c *Lyngbya* spp., *Tolypothrix* spp. and *Rivularia* spp. previously observed in the system [1,2].

1. Eriksson, B. K., Ljunggren, L., Sandström, A., Johansson, G., Mattila, J., Rubach, A., Råberg, S. & Snickars, M. 2009 Declines in predatory fish promote bloom-forming macroalgae. *Ecol. Appl.* **19**, 1975–88. (doi:10.1890/08-0964.1)
2. Hansen, J. P., Wikström, S. A. & Kautsky, L. 2008 Effects of water exchange and vegetation on the macroinvertebrate fauna composition of shallow land-uplift bays in the Baltic Sea. *Estuar. Coast. Shelf Sci.* **77**, 535–547. (doi:10.1016/j.ecss.2007.10.013)

APPENDIX D

List of species identified as macroalgal grazers, i.e. shredders and gatherers whose diet includes large proportion of plant material.

Crustacea
<i>Asellus aquaticus</i>
<i>Gammarus</i> spp.
<i>Gammarus duebeni</i>
<i>Gammarus locusta</i>
<i>Gammarus oceanicus</i>
<i>Gammarus salinus</i>
<i>Gammarus zaddachi</i>
<i>Idotea</i> spp.
<i>Idotea balthica</i>
<i>Idotea chelipes</i>
Hexapoda

<i>Agraylea multipunctata</i> (larvae)
<i>Cataclysta lemnata</i> (larvae)
Chironomidae (larvae, pupae)
Chrysomelidae (adults, larvae) ^a
Curculionidae (larvae) ^b
<i>Hydroptila</i> sp. (larvae)
<i>Haliphus</i> sp. (larvae)
Limnephilidae (larvae)
Pyralidae (larvae)

^a Donaciinae spp. and ^b *Bagous* sp. previously observed in the system [1].

1. Hansen, J. P., Wikström, S. A. & Kautsky, L. 2008 Effects of water exchange and vegetation on the macroinvertebrate fauna composition of shallow land-uplift bays in the Baltic Sea. *Estuar. Coast. Shelf Sci.* 77, 535–547.
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APPENDIX E

Effects of piscivores on grazer assemblage composition

Methods

To test whether piscivores biomass influenced the composition of the grazer assemblage across the 32 bays, we first calculated the community-weighted biomass of each of 12 grazer taxa found (i.e. their relative contribution to total biomass) at the bay scale. Second, we used a permutational multivariate analysis of variance (PERMANOVA, 9999 permutations) to test whether piscivores biomass affected grazer composition (based on Bray-Curtis dissimilarities), and included salinity, plant cover and bay openness as covariates, using the *adonis* function in the vegan R package, version 2.4-1 [1]. Finally, we visualized changes in the grazer community composition along gradients of piscivores biomass and plant cover (which were significant predictors in the PERMANOVA analysis, see below) using contour plots on a non-metric multidimensional scaling ordination model (nMDS) [2].

Results

The bay-level grazer composition was influenced by piscivores biomass ($F = 4.66$, $P = 0.007$) and plant cover ($F = 4.81$, $P = 0.007$), while salinity and bay openness had no effects ($P \gg 0.05$). In bays with low piscivores biomass, the grazer assemblage was dominated by *Idotea* spp. and *Gammarus* spp., while in bays with high piscivores biomass, the assemblages were dominated by a mix of freshwater taxa including *Asellus aquaticus*, *Hydroptila* sp. and Chironomidae (Fig. E1). In bays with low plant cover, a mix of *Hydroptila*, *Asellus aquaticus* and Chironomidae dominated, while in bays with high plant cover, *Idotea* spp. and *Gammarus* spp. were prevalent (Fig. E2).

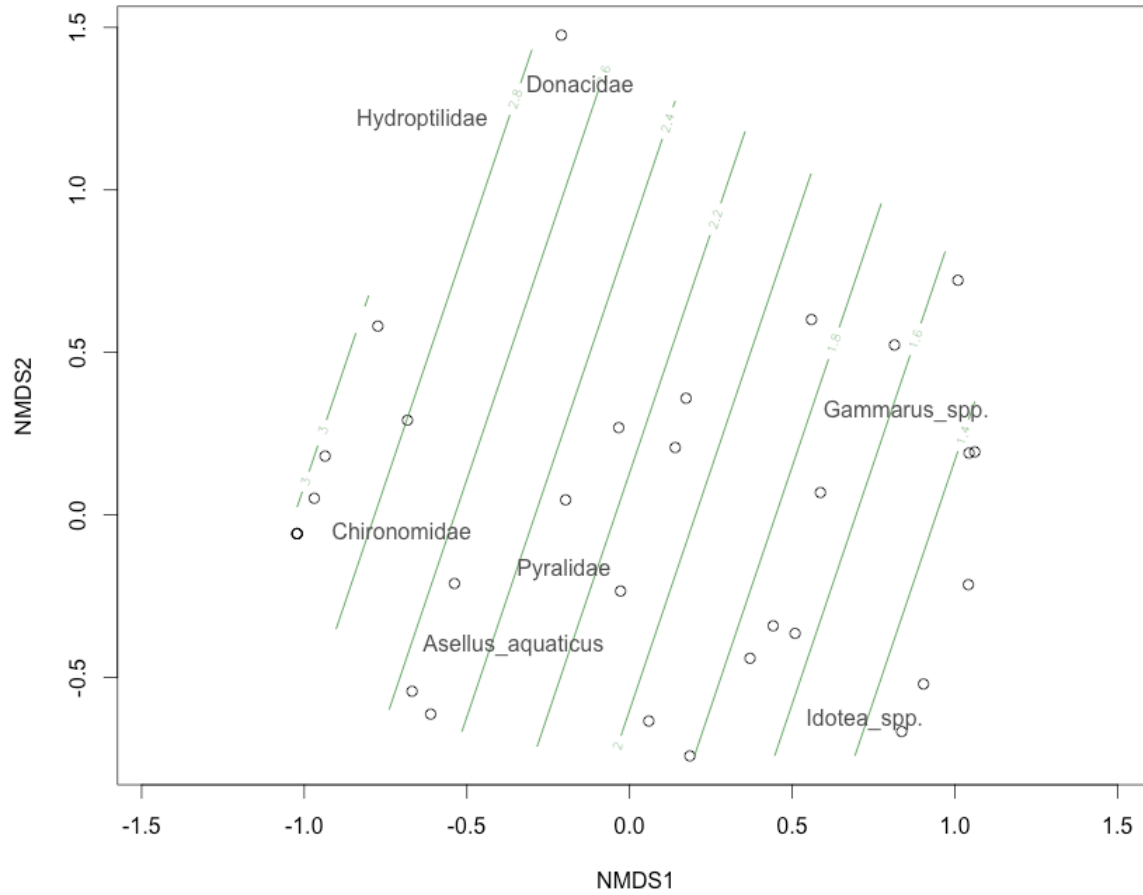


Fig. E1. NMDS plot of the grazer community composition with contours showing piscivores biomass modelled as a function of the ordination scores. Stress = 0.12. A stress value below 0.20 indicates a reliable ordination.

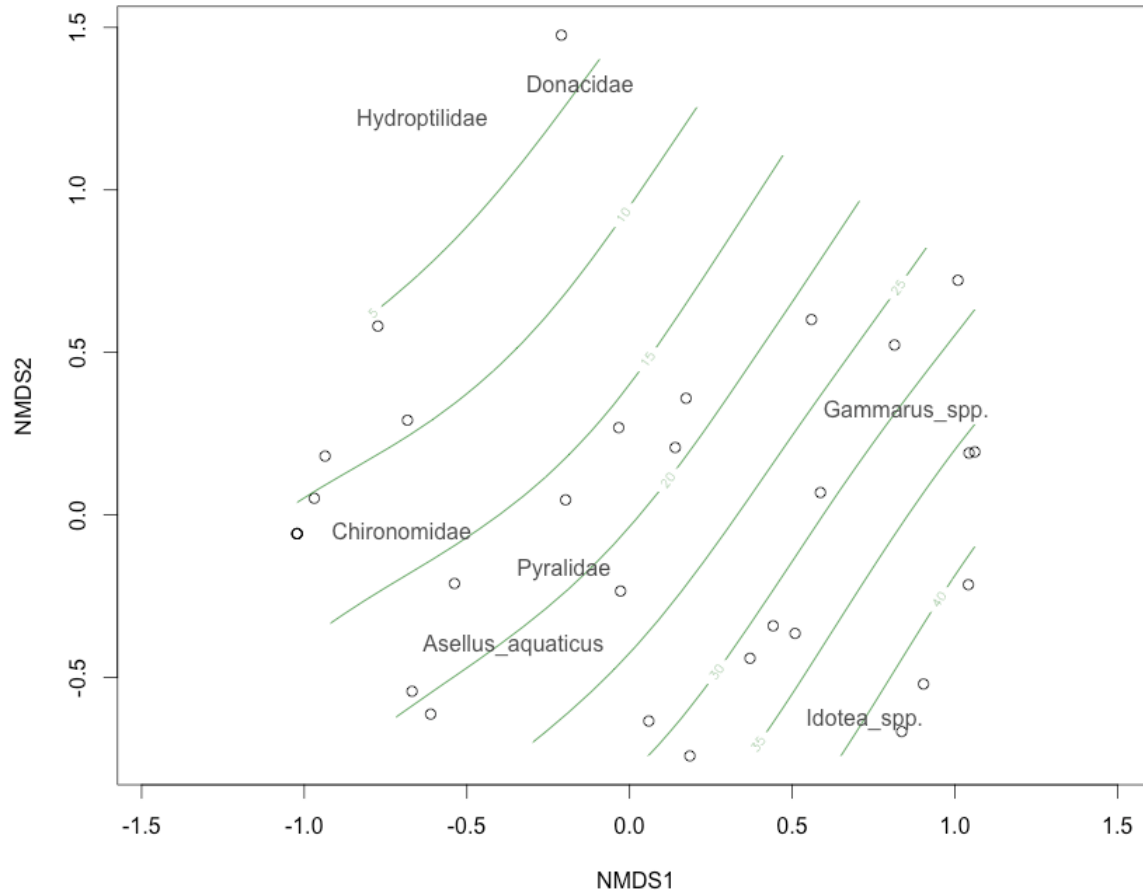


Fig. E2. NMDS plot of the grazer community composition with contours showing plant cover modelled as a function of the ordination scores. Stress = 0.12. A stress value below 0.20 indicates a reliable ordination.

1. Oksanen et al. 2016. Vegan: community ecology package. R package version 2.4-1.
2. Kruskall, J.B. & Wish, M. 1978. Multidimensional Scaling. Sage, Beverly Hills, CA, USA.

APPENDIX F

Three-spined stickleback in stomachs of perch and pike

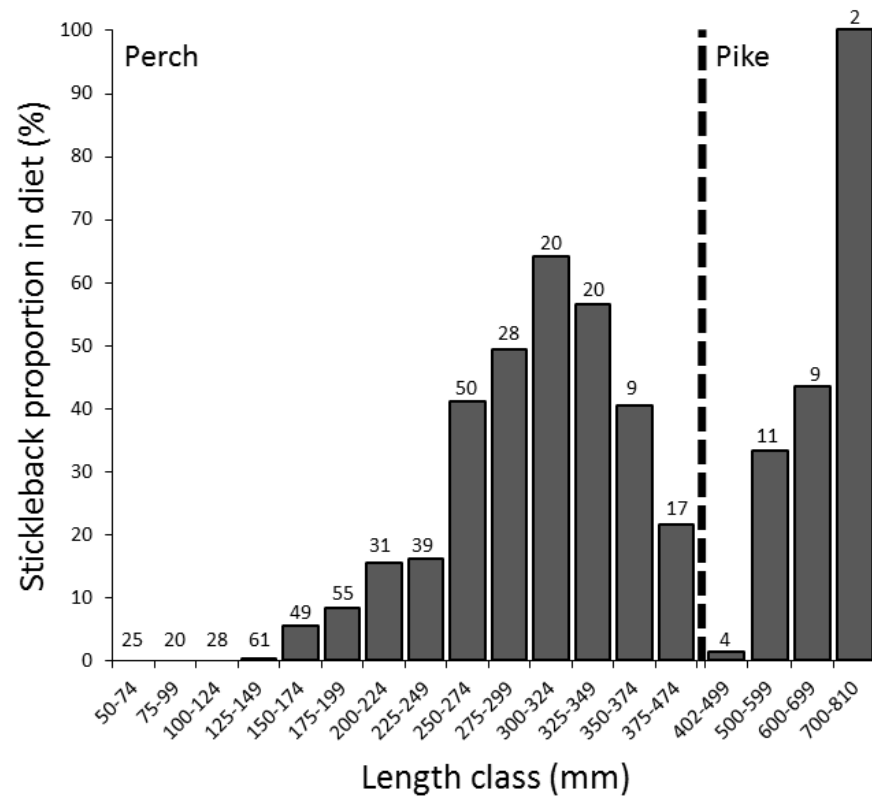


Fig. D1. Three-spined sticklebacks in stomachs of perch (left) and pike (right) expressed as percentage of total stomach content volume (following [1]). Data are shown for perch total length classes of 25 mm and pike total length classes of 100 mm. Numbers of individuals dissected for each length class are shown above bars.

1. Hyslop, E. J. 1980 Stomach contents analysis -a review of methods and their application. *J. Fish Biol.* 17, 411–429. (doi: 10.1111/j.1095-8649.1980.tb02775.x)