Supplementary Information

Assessing endgame strategies for the elimination of lymphatic filariasis: A model-based evaluation of the impact of DEC-medicated salt

Morgan E Smith¹, Brajendra K Singh¹, Edwin Michael^{1*}

1 Department of Biological Sciences, University of Notre Dame, Notre Dame, IN 46556, USA

*Corresponding author (emichael@nd.edu, 349 Galvin Life Science Center, University of Notre Dame, Notre Dame, IN 46556)

Supplementary Results

Supplementary Figure S1. Estimates of site-specific drug efficacies using model fits to intervention community trial data. This figure accompanies Figure 2 in the main text, reporting the results of the remaining trial sites. The 500 parameter vectors fitted to baseline conditions were used to project the impact of DEC-medicated salt with each vector simulated 500 times via sampling from an initial range of plausible drug parameter values. Intervention survey data used to accept or reject a parameter vector are represented by red crosses with 95% binomial error bars in the main plot axes. Gray curves are the model fits from which monthly worm and mf kill rates per site were calculated. Inset plots show the posterior relative frequency distributions of the monthly worm and mf kill rates for each site.

Supplementary Table S1. Median estimates of DEC-medicated salt monthly worm and mf killing rates as predicted by model fits to intervention community trial data.

Supplementary Table S2. Model-predicted site-specific mf breakpoints under ABR and TBR conditions for three elimination probabilities.

Supplementary Table S3. Mean differences between intervention times under Regimen A and Regimen B used in the posthoc Tukey's HSD multiple comparison tests. Highlighted cells indicate there were significant differences between the group means being compared (p-values < 0.05).

Supplementary Table S4. Test statistics, degrees of freedom, and p-values for pairwise two sample F-tests testing the differences in variation of the time required to reach elimination under different treatment regimens. One-tailed two-sample F-tests were performed to test whether the variance of Regimen A (column label) was less than the variance of Regimen B (row label). Significant p-values are highlighted. In all comparisons, $df1 = 11078$ and $df2 = 11078$.

The mathematical model of LF transmission dynamics

We employed a *Culex* mosquito-vectored transmission model of LF to carry out the modelling work in this study¹⁻⁷. Briefly, the state variables of this hybrid coupled partial differential and differential equation model vary over age (*a*) and/or time (*t*), representing changes in the prepatent worm burden per human host($P(a,t)$), adult worm burden per human host($W(a,t)$), the microfilariae (mf) level in the human host modified to reflect infection detection in a 1 mL blood sample $(M(a,t))$, the average number of infective L3 larval stages per mosquito (L), and a measure of immunity $(I(a,t))$ developed by human hosts against L3 larvae. The state equations comprising this model are:

$$
\frac{\partial P}{\partial t} + \frac{\partial P}{\partial a} = \lambda \frac{V}{H} h(a) \Omega(a, t) - \mu P(a, t) - \lambda \frac{V}{H} h(a) \Omega(a, t - \tau) \zeta
$$

\n
$$
\frac{\partial W}{\partial t} + \frac{\partial W}{\partial a} = \lambda \frac{V}{H} h(a) \Omega(a, t - \tau) \zeta - \mu W(a, t)
$$

\n
$$
\frac{\partial M}{\partial t} + \frac{\partial M}{\partial a} = \alpha s \phi[W(a, t), k] W(a, t) - \gamma M(a, t)
$$

\n
$$
\frac{\partial I}{\partial t} + \frac{\partial I}{\partial a} = W_T(a, t) - \delta I(a, t)
$$

\n
$$
\frac{dL}{dt} = \lambda \kappa g \int \pi(a) (1 - f[M(a, t)]) da - (\sigma + \lambda \psi_1) L
$$

\n
$$
L^* = \frac{\lambda \kappa g \int \pi(a) (1 - f[M(a, t)]) da}{\sigma + \lambda \psi_1}
$$

The above equations involve partial derivatives of four state variables (*P* - pre-patent worm load; *W* - adult worm load; *M* - microfilaria intensity; *I* - immunity to acquiring new infection due to the pre-existing total worm load where $W_T = W(a,t) + P(a,t)$. Given the faster time scale of infection dynamics in the vector compared to the human host, the infective L3-stage larval density in mosquito population is modelled by an ordinary differential equation essentially reflecting the significantly faster time-scale of the infection dynamics in the vector hosts. This allows us to make the simplifying assumption that the density of infective stage larvae in the vector population reaches a dynamic equilibrium (denoted by L^{*}) rapidly^{1, 2, 5, 8, 9}. This basic coupled

immigration-death structure of the model as well as its recent extensions has been extensively discussed previously^{1-3, 5, 8, 9}. The effects of worm patency are captured by considering that at any time *t*, human individuals of age less than or equal to the pre-patency period, *τ*, will have no adult worms or Mf, and the rate at which pre-patent worms survive to become adult worms in these individuals at $a > r$ is given by $\zeta = \exp(-\mu \tau)$. The term $f(M)$ enables us to account for the different establishment and development rates of the incoming L3-stage larvae as adult worms depending on the genus of mosquito vectors. For culicine-mediated LF, the functional form reflecting a negative-density dependent development of L3 larvae from ingested mf was deployed in the model¹. See Supplementary Table S5 for the description of all the model parameters and functions.

Supplementary Table S5 - **Description the basic LF model parameters and functions used in the model.**

¹The proportion of L3-stage larvae infecting human hosts that survive to develop into adult worms².
²The gradient of Mf uptake *r* is a measure of the initial increase in the infective L3 larvae uptake by vector as

³The facilitated establishment rate of adult worms due to parasite-induced immunosuppression in a heavily infected human host ⁴The initial rate of increase by which the strength of immunosuppression is achieved as *W*

Note MBR (monthly biting rate) serves as an input to initialize the model, measured as mosquito bites per person per month, the value of which may be obtained from entomological surveys conducted in study sites. In the absence of the observed MBR value, the model has been adapted to estimate it from the community-level Mf prevalence data.

Mf age profile construction in the absence of age-stratified infection data

The Bayesian Melding (BM) procedure for calibrating our deterministic LF model with data relies on baseline age profiles of microfilaria (mf) prevalence, but, in some cases, only the overall community level mf prevalence was available for a particular site in this study. This therefore required the translation of the overall prevalence into theoretical age infection profiles. This was done by firstly fitting equations to datasets (Supplementary Table S6 and Supplementary Table S7) of age-stratified mf infection data which qualitatively follow either plateau or convex profiles (Supplementary Figure S2). Specifically, plateau and convex age prevalence curves were defined by the following equations where *P* is the mf prevalence as a function of age *a*:

+ (a) = $\frac{0.0018a}{2}$ for plateau-type age profiles, and 1 11.99 $P(a) = \frac{0.0018a}{1+a}$

 $P(a) = 0.029a * \exp[-0.033a]$ for convex-style age profiles.

The number of infected individuals in each age class was then derived from the observed overall mf prevalence by applying these equations to the overall prevalence, while subdividing the total population into age-classes in each site according to their respective national agedemographic patterns.

Supplementary Table S6. Plateau-style age profile data

Site Age No. Examined No. mf Positive Reference Bhagalpur 0-5 345 26 29 6-10 601 45 11-20 204 62 21-30 334 56 31-40 234 32 41-50 144 21 51+ 78 10 Gonda Town 0-1 13 0 30 2-5 205 5 6-10 443 23 11-20 807 65 21-30 773 88 31-40 509 57 41-50 206 19 50+ 142 14 Pondicherry 0-5 121 4 31 6-10 211 29 11-15 224 43 16-20 195 52 21-30 328 60 31-40 215 38 41-50 121 22 51-60 82 16 61+ 52 12 Gorakhpur 0-1 11 0 32 2-5 1678 56 6-10 2867 236 11-20 4070 675 21-30 5212 778 31-40 3045 429 41-50 4051 590 50+ 1400 180 Chaliyum Ward I 1-4 283 0 33 5-14 926 92 15-24 509 77 25-34 324 38 35-54 346 27 55+ 158 9

Supplementary Table S7. Convex style age profile data

Supplementary Figure S2. Fits to (A) plateau- and (B) convex-style age infection profiles.

References

1. Gambhir, M. & Michael, E. Complex ecological dynamics and eradicability of the vector borne macroparasitic disease, lymphatic filariasis. *PLoS One* **3**, e2874 (2008).

2. Gambhir, M. *et al*. Geographic and ecologic heterogeneity in elimination thresholds for the major vector-borne helminthic disease, lymphatic filariasis. *BMC biology* **8**, 1 (2010).

3. Michael, E. *et al*. Mathematical modelling and the control of lymphatic filariasis. *The Lancet infectious diseases* **4**, 223-234 (2004).

4. Michael, E., Malecela-Lazaro, M. N., Kabali, C., Snow, L. C. & Kazura, J. W. Mathematical models and lymphatic filariasis control: endpoints and optimal interventions. *Trends Parasitol.* **22**, 226-233 (2006).

5. Singh BK and Bockarie MJ and Gambhir M and Siba PM and Tisch DJ and Kazura J and others. in *Sequential Modelling of the Effects of Mass Drug Treatments on Anopheline-Mediated Lymphatic Filariasis Infection in Papua New Guinea* (PLoS One, 2013).

6. Singh, B. K. & Michael, E. Bayesian calibration of simulation models for supporting management of the elimination of the macroparasitic disease, Lymphatic Filariasis. *Parasit vectors* **8**, 1-26 (2015).

7. Michael, E. & Singh, B. K. Heterogeneous dynamics, robustness/fragility trade-offs, and the eradication of the macroparasitic disease, lymphatic filariasis. *BMC medicine* **14**, 1 (2016).

8. Chan, M. S. *et al*. Epifil: a dynamic model of infection and disease in lymphatic filariasis. *Am. J. Trop. Med. Hyg.* **59**, 606-614 (1998).

9. Norman, R. *et al*. EPIFIL: the development of an age-structured model for describing the transmission dynamics and control of lymphatic filariasis. *Epidemiol. Infect.* **124**, 529-541 (2000).

10. Rajagopalan, P. Population dynamics of culex pipiens fatigans, the filariasis vector, in pondicherry: influence of climate and environment. *Proc Ind Nat Science Acad B* **46**, 745-752 (1980).

11. Subramanian, S., Manoharan, A., Ramaiah, K. D. & Das, P. K. Rates of acquisition and loss of Wuchereria bancrofti infection in Culex quinquefasciatus. *Am. J. Trop. Med. Hyg.* **51**, 244-249 (1994).

12. Scott, A. L. & Nutman, T. Lymphatic-dwelling filariae. *Lymphatic filariasis.*, 5-39 (2000).

13. Vanamail, P., Subramanian, S., Das, P. K., Pani, S. P. & Rajagopalan, P. K. Estimation of fecundic life span of Wuchereria bancrofti from longitudinal study of human infection in an endemic area of Pondicherry (south India). *Indian J. Med. Res.* **91**, 293-297 (1990).

14. Evans, D. B., Gelband, H. & Vlassoff, C. Social and economic factors and the control of lymphatic filariasis: a review. *Acta Trop.* **53**, 1-26 (1993).

15. Ottesen, E. & Ramachandran, C. Lymphatic filariasis infection and disease: control strategies. *Parasitology Today* **11**, 129-130 (1995).

16. Vanamail, P. *et al*. Estimation of the fecund life span of Wuchereria bancrofti in an endemic area. *Trans. R. Soc. Trop. Med. Hyg.* **90**, 119-121 (1996).

17. Hairston, N. G. & de Meillon, B. On the inefficiency of transmission of Wuchereria bancrofti from mosquito to human host. *Bull. World Health Organ.* **38**, 935-941 (1968).

18. Subramanian, S. *et al*. The relationship between microfilarial load in the human host and uptake and development of Wuchereria bancrofti microfilariae by Culex quinquefasciatus: a study under natural conditions. *Parasitology* **116**, 243-255 (1998).

19. Subramanian, S., Pani, S., Das, P. & Rajagopalan, P. Bancroftian filariasis in Pondicherry, south India: 2. Epidemiological evaluation of the effect of vector control. *Epidemiol. Infect.* **103**, 693-702 (1989).

20. Das, P. *et al*. Bancroftian filariasis in Pondicherry, south India–epidemiological impact of recovery of the vector population. *Epidemiol. Infect.* **108**, 483-493 (1992).

21. Ho, B. C. & Ewert, A. Experimental transmission of filarial larvae in relation to feeding behaviour of the mosquito vectors. *Trans. R. Soc. Trop. Med. Hyg.* **61**, 663-666 (1967).

22. May, R. M. Togetherness among schistosomes: its effects on the dynamics of the infection. *Math. Biosci.* **35**, 301-343 (1977).

23. Duerr, H., Dietz, K. & Eichner, M. Determinants of the eradicability of filarial infections: a conceptual approach. *Trends Parasitol.* **21**, 88-96 (2005).

24. Jain, D. C., Menon, P. K., Sethumadhavan, K. V., Johney, V. M. & Ghosh, T. K. Epidemiology of bancroftian filariasis in a semi-urban community of Kerala State. *J. Commun. Dis.* **21**, 265-271 (1989).

25. Singh, M., Rastogi, K., Singh, R. & Srivastava, V. Observations on rural filariasis in Sitapur District (Uttar Pradesh). *Indian J. Malariol.* **17**, 303-310. (1963).

26. Basu, P., Rao, V. & Pattanayak, S. Filariasis in greater Bombay-results of a rapid survey conducted in June, 1965. *Bull.Indian Soc.for Malaria & other Communicable Dis.* **4**, 296-317. (1967).

27. Chand, D., Singh, M. & Pathak, V. Filariasis in the District of Ghazipur (Uttar Pradesh). *Indian J. Malariol.* **15**, 21-29 (1961).

28. Rath, R., Das, R., Mishra, G., Mohapatra, B. & Ramakrishna, C. Bancroftian filariasis in two selected rural communities in Puri District: Orissa―a comparative study of filariometric data. *J. Commun. Dis.* **16**, 104-112 (1984).

29. Varma, B., Dass, N. & Sinha, V. Filariasis in the Rural Population around Bhagalpur Town. Parts I. *Indian J. Malariol.* **15**, 285-292 (1961).

30. Chand, D., Singh, M., Gupta, B. & Srivastava, R. A note on filariasis in Gonda Town (Uttar Pradesh). *Indian J. Malariol.* **15**, 39-17 (1961).

31. Rajagopalan, P., Kazmi, S. & Mani, T. Some aspects of transmission of Wuchereria bancrofti and ecology of the vector Culex pipiens fatigans in Pondicherry. *Indian J. Med. Res.* **66**, 200-215 (1977).

32. Chand, D., Singh, M. & WAS, L. Filariasis in Gorakhpur District, Uttar Pradesh. *Indian J. Malariol.* **16**, 269-276 (1962).

33. Jain, D., Ravindranathan, T., Ghosh, T., PILLAI, K. & Rao, C. Prophylactic effect of diethylcarbamazine on W. bancrofti filariasis. *J. Commun. Dis.* **19**, 317-321 (1987).

34. Dondero, T. J.,Jr *et al*. Clinical manifestations of Bancroftian filariasis in a suburb of Calcutta, India. *Am. J. Trop. Med. Hyg.* **25**, 64-73 (1976).