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Supplemental Information

Adult Mouse Liver Contains Two Distinct Populations of Cholangiocytes

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В С **EpCAM** DAPI 10 MIC1-1C3 PE 1.0 10 10⁰ **ST14** Merge APC Isotype ST14 APC D 60K 0 40K S S 20K 0 40K FSC Н F G Lgr5 mRNA level 100 י 76.0% 62.2% **...** 87.7% fold change 80 CD133 CD133 CD133 60 40 20 N.D. 0 ST^{hi}Org ST^{hi} 2D ST^{lo}Org CD26 ST14 PE CD26 **ST14** ANXA13 SLC34A2 COLLECTRIN **CD24** 13.3% 0.4% 16.5% 5.58% 0.69% 20 15 10

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Figure S1 Related to Figure1. Evaluation of duct-subdividing antibody candidates.

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(A) Representative immunohistochemistry of ST14 (red) and EpCAM (green) colocalization in mouse liver duct cells. DAPI: blue. Scale bar = 50 μm. (B-D) Flow cytometry analyses of M+26-CD45/31/11b-PI- cells stained with (B) secondary APC isotype control.only and (C) the ST14 primary antibody. (D) Size and scatter properties of fully gated ST14IoM+ cells. (E-G) Flow cytometry analyses showing the extent of CD133 and ST14 coexpression in duct cells. (E) 76% of M+CD45/31/11b-PI- cells are CD26 negative. (F) in the M+CD26-CD45/31/11b-PI- cells from (E) 62.2% are CD133 expressing. (G) 87.7% of the M+ST14hi cells are CD133 positive. (H) *Lgr5* mRNA expression was activated by the organoid culture conditions in the ST14hi but not in the ST14lo organoids. (I) Antigen screening for ST14, ANXA13, SLC34A2, COLLECTRIN and CD24 in the CD26-M+ duct cells. Lighter grey delineates the isotype control.

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Figure S2 Related to Figure 2. Clonogenic duct cells survival during ischemia and in the injured liver Flow cytometry analyses for MIC1-1C3+ and ST14. 45% of MIC1-1C3 duct cells were ST14hi after 24hrs of warm ischemia. After warm ischemia organoid formation was strictly limited to only ST14hi ductal cells. Far left panel: unsorted liver NPCs in organoid cell culture. FACS sorted ST14hi (middle panel) and ST14lo (far right panel) cells were embedded in Matrigel droplets at a density of 2,000 per 24 well. Culture date 14. No organoids formed from the ST14lo population. (C-G) ST14 expression in M+ liver duct cells in uninjured controls (D) and CCl4 induced liver injury day 6 (E) and DDC treated liver day 6 (F). CCl4 was diluted in corn oil and administrated via I.P. injection at single dose of 1ml/kg body weight. Sham: corn oil only. Isotype control was used as negative gating. (G) NPCs from livers injured by CCl4 were sorted into 96 well dishes for single cell organoid analysis. There were no significant injury-induced differences in the organoid forming ability of either population. ST14hi cells were superior to ST14lo duct cells under both conditions. Statistical analyses: Mann-Whitney U test for normal vs treated, paired t test for ST14hi vs ST14lo. Independent mice t reated with CCl4 n=3; Independent mice treated with sham n=4. Sorted cells from ST14 large organoid







Figure S3. Related to Figure 3. Characterization of ST14hi derived organoids in vitro.

ST14hi derived organoids at passage 1 were dissociated and resorted using surface maker ST14 as in Figure 2I. (A) Relative *Lgr5* mRNA level from the ST14hi and ST14lo population detected by qPCR. Relative *Lgr5* mRNA expression was normalized to house keeping gene *Gapdh*. Single bands for the *Lgr5* and *Gapdh* PCR products were confirmed by agarose gel. Quantities were calculated with $-\Delta\Delta$ Ct. Independent experiments n=3, upaired t test. *P=0.0286. (B) Immunofluorence staining showed Fah expression in cultured ST14hi and ST14lo derived organoids. Arrow shows FAH+ but EpCAM- cells. Scale bar = 20 µm.

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Figure S4. Related to Figure 5. ST14 protein expression in human liver.

(A) Immunofluorence showed the ST14(red) expression in the EPCAM(green) expressing duct cells in the human liver. Nuclei stained with DAPI as blue. Arrow heads show the ST14 expressing cells. Scale bar = $20 \ \mu m$. (B-E) Primary human liver NPCs were FACS analyzed after staining with EPCAM and ST14. In the EPCAM+ subpopulation, $18.9\pm4.65\%$ cells were ST14hi (Independent patients n=3, unpaired t test, *p=0.001) (E). (F) Sorted cells were put into matrigel organoid culture at a density of 2000 cells/well in the 24 well suspension culture plate. Scale bar = 2 mm.

Table S1 Selected hepatic biliary and progenitor markers in ST14^{hi} vs ST14^{lo} populations

| Gene | | Transcript | ST14 ^{hi} | ST14 ^{lo} | FC | Pval | FDR |
|-------|---|------------|--------------------|--------------------|-------|------|------|
| St14 | Suppression of tumorigenicity 14 (colon) | NM_011176 | 100 | 100 | 1.1 | 1.09 | 0.35 |
| Krt19 | Keratin 19 | NM_008471 | 617.75 | 428.5 | -1.44 | 0.32 | 0.54 |
| Sox9 | SRY (sex determining region Y)-box 9 | NM_011448 | 114.3 | 119.8 | -1.05 | 0.56 | 0.65 |
| Epcam | Epithelial cell adhesion molecule | NM_008532 | 334.3 | 346.8 | -1.04 | 0.67 | 0.69 |
| Hnf1b | Transcription factor 2, transcription factor 2, | NM_009330 | 195.75 | 207.25 | 1.06 | 0.55 | 0.65 |
| Cftr | Cystic fibrosis transmembrane conductance | NM_021050 | 16.25 | 20.5 | 1.26 | 0.07 | 0.32 |
| Prom1 | Prominin 1 (CD133 antigen) | NM_008935 | 181.75 | 187.75 | 1.03 | 0.83 | 0.73 |
| Lgr5 | Leucine rich repeat containing G protein coupled receptor 5 | NM_010195 | 0.25 | 0 | 2.5 | 0.02 | 0.17 |
| Thy1 | Thymus cell antigen 1, theta | NM_009382 | 3 | 1.25 | 2.4 | 0.11 | 0.37 |
| lcam1 | Intercellular adhesion molecule 1 | NM_010493 | 59.5 | 36.25 | 1.64 | 1 | 1 |
| Bmi1 | Bmi1 polycomb ring finger oncogene | NM_007552 | 32.75 | 34.75 | -1.06 | 0.7 | 0.7 |
| Cd47 | CD47 antigen | NM_010581 | 52.25 | 67.5 | -1.29 | 0.06 | 0.3 |

Tag numbers are given in RPKM for the ST14^{hi} and ST14^{lo} populations; FC = fold change. None of the genes listed were differentially expressed between progenitor and non-progenitor ducts.

Table S2 Selected gene sets significantly enriched in the clonogenic ST14^{hi} population

| NAME | ES | NOM p-val | FDR q-val |
|--|------|-----------|-----------|
| BOQUEST_STEM_CELL_UP | 0.53 | 0.00 | 0.00 |
| ANASTASSIOU_CANCER_MESENCHYMAL_TRANSITION_SIGNATURE | 0.64 | 0.00 | 0.00 |
| LIM_MAMMARY_STEM_CELL_UP | 0.46 | 0.00 | 0.00 |
| LIEN_BREAST_CARCINOMA_METAPLASTIC | 0.67 | 0.00 | 0.00 |
| VECCHI_GASTRIC_CANCER_ADVANCED_VS_EARLY_UP | 0.47 | 0.00 | 0.00 |
| ONDER_CDH1_SIGNALING_VIA_CTNNB1 | 0.53 | 0.00 | 0.00 |
| SERVITJA_ISLET_HNF1A_TARGETS_UP | 0.47 | 0.00 | 0.00 |
| MIKKELSEN_IPS_ICP_WITH_H3K4ME3_AND_H327ME3 | 0.50 | 0.00 | 0.00 |
| STAMBOLSKY_TARGETS_OF_MUTATED_TP53_UP | 0.56 | 0.00 | 0.01 |
| EBAUER_TARGETS_OF_PAX3_FOXO1_FUSION_UP | 0.43 | 0.00 | 0.01 |
| ACEVEDO_LIVER_CANCER_WITH_H3K27ME3_DN | 0.44 | 0.00 | 0.01 |
| KEGG_HEDGEHOG_SIGNALING_PATHWAY | 0.50 | 0.00 | 0.02 |
| PLASARI_TGFB1_TARGETS_10HR_UP | 0.38 | 0.00 | 0.02 |
| BOQUEST_STEM_CELL_CULTURED_VS_FRESH_UP | 0.35 | 0.00 | 0.02 |
| SWEET_KRAS_TARGETS_UP | 0.44 | 0.00 | 0.02 |
| WIEDERSCHAIN_TARGETS_OF_BMI1_AND_PCGF2 | 0.49 | 0.00 | 0.02 |
| DASU_IL6_SIGNALING_SCAR_UP | 0.52 | 0.00 | 0.03 |
| VERRECCHIA_RESPONSE_TO_TGFB1_C2 | 0.54 | 0.00 | 0.03 |
| BOQUEST_STEM_CELL_CULTURED_VS_FRESH_DN | 0.55 | 0.00 | 0.03 |
| TURASHVILI_BREAST_DUCTAL_CARCINOMA_VS_DUCTAL_NORMAL_DN | 0.39 | 0.00 | 0.03 |
| SCHUETZ_BREAST_CANCER_DUCTAL_INVASIVE_UP | 0.35 | 0.00 | 0.03 |
| JECHLINGER_EPITHELIAL_TO_MESENCHYMAL_TRANSITION_UP | 0.44 | 0.00 | 0.03 |
| PID_HEDGEHOG_2PATHWAY | 0.55 | 0.01 | 0.03 |
| GUO_HEX_TARGETS_UP | 0.43 | 0.00 | 0.04 |
| LEE_LIVER_CANCER_HEPATOBLAST | 0.62 | 0.01 | 0.04 |
| IVANOVA_HEMATOPOIESIS_STEM_CELL_LONG_TERM | 0.35 | 0.00 | 0.04 |
| CHIANG_LIVER_CANCER_SUBCLASS_CTNNB1_DN | 0.38 | 0.00 | 0.04 |
| WHITFIELD_CELL_CYCLE_G1_S | 0.37 | 0.00 | 0.06 |
| CONRAD_STEM_CELL | 0.50 | 0.02 | 0.06 |
| SAKAI_CHRONIC_HEPATITIS_VS_LIVER_CANCER_DN | 0.49 | 0.02 | 0.06 |
| PLASARI_TGFB1_TARGETS_10HR_DN | 0.33 | 0.00 | 0.08 |
| ELVIDGE_HIF1A_AND_HIF2A_TARGETS_DN | 0.37 | 0.00 | 0.08 |
| ZHANG_GATA6_TARGETS_DN | 0.39 | 0.01 | 0.09 |
| PID_BETA_CATENIN_DEG_PATHWAY | 0.54 | 0.05 | 0.09 |
| HAN_JNK_SINGALING_DN | 0.45 | 0.02 | 0.09 |
| HOSHIDA_LIVER_CANCER_SURVIVAL_UP | 0.38 | 0.01 | 0.09 |

Table S3 List of antibodies used

| 1 st Antibodies | | | | | |
|----------------------------|------------|------------|----------|----------------|------------|
| Name | IgG type | Use | Dilution | Company | Lot |
| ALB | Goat pAB | IF | 100 | Bethyl | A90-134A-6 |
| ANXA13 | Rabbit pAB | IF | 100 | Sigma-Aldrich | HPA019650 |
| CD11b | Rat mAB | FACS | 100 | BD biosciences | 552850 |
| CD133 | Biotin | FACS | 100 | eBioscience | 13-1331-82 |
| CD24 | Rat mAB | FACS | 100 | BD biosciences | 561079 |
| CD31 | Rat mAB | FACS | 100 | BD biosciences | 561410 |
| CD45 | Rat mAB | FACS | 100 | BD biosciences | 552848 |
| CK19 | Rabbit pAB | IF | 500 | Cell Lab Tech | A03189 |
| COLLECTRIN | Sheep pAB | FACS | 100 | R&D systems | AF4965 |
| EpCAM | Rat mAB | IF | 100 | BD biosciences | G8.8 |
| EpCAM | Mouse mAB | huIF, FACS | 100 | DAKO | Ber-EP4 |
| FAH | Rabbit pAB | IF | 50 | Grompe lab | |
| HNF4A | Rabbit pAB | IF | 100 | Santa Cruz Bio | sc-8987 |
| MIC1-1C3 | Rat mAB | FACS | 20 | Grompe lab | |
| SLC34A2 | Rabbit pAB | FACS | 100 | Abbiotech | 251343 |
| ST14 | Rabbit pAB | FACS | 100 | Abcam | ab28266 |
| ST14 | Rabbit pAB | huIF, FACS | 100 | Invitrogen | PA5-29764 |

2nd Antibodies

| Name | lgG type | Use | Dilution | Company | Lot |
|------------------|--------------|------|----------|---------------|-------------|
| Anti rat DL649 | Mouse | FACS | 200 | JacksonImmuno | 212-496-168 |
| Anti rabbit APC | Donkey | FACS | 200 | JacksonImmuno | 711-056-152 |
| Anti rabbit PE | Donkey | FACS | 200 | JacksonImmuno | 711-116-152 |
| Anti Rat PE | Goat | FACS | 200 | JacksonImmuno | 112-116-143 |
| Anti rabbit Cy3 | Donkey | IF | 250 | JacksonImmuno | 711-166-152 |
| Anti sheep AF647 | Donkey | FACS | 250 | Jacksonimmuno | 713-606-147 |
| APC/cy7 | Streptavidin | FACS | 250 | Biolegend | 405208 |
| | | | | | |

Table S4 Human samples information

| Use | Gender | Age | Source |
|------------------|---------|---------|---|
| IHC | Unknown | Unknown | Gift from Strom lab at University of Pittsburg, U.S.A |
| FACS | Female | 17 | Lonza, LLC. U.S.A |
| FACS | Male | 29 | Lonza, LLC. U.S.A |
| FACS, culture | Female | 48 | Lonza, LLC. U.S.A |