Supplementary material for Takahashi *et al*. (August 7, 2001) *Proc. Natl. Acad. Sci. USA*, 10.1073/pnas.171209998

Microarray Experiments. Microarray production was performed as described, with slight modification. Briefly, 21,632 cDNA clones were PCR amplified directly from bacterial stocks purchased from Research Genetics (Huntsville, AL). After ethanol precipitation and transfer to 384-well plates, clones were printed onto poly-L-lysine-coated glass slides by using a home-built robotic microarrayer (http://cmgm.stanford.edu/pbrown/). Slides were blocked by using a BSA solution (1% BSA/5× SSC/0.1% SDS), as described. Two micrograms of poly(A)$^+$ RNA from each kidney tumor and 2 μg of poly(A)$^+$ RNA from each patient-matched normal kidney tissue were reverse transcribed with oligo(dT) primer and Superscript II (Life Technologies, Rockville, MD) in the presence of Cy5-dCTP and Cy3-dCTP (Amersham Pharmacia Biotech), respectively. The Cy3- and Cy5-labeled cDNA probes were mixed with a 2× hybridization solution (50% formamide/10× SSC/0.2% SDS) and hybridized to prewarmed (42ºC) slides for 20 hours at 42ºC. After hybridization, slides were washed in 1× SSC/0.1% SDS at 42ºC for 5 min, followed by 0.2× SSC/0.1% SDS at room temperature (RT) for 5 min, 0.2× SSC at RT for 5 min twice, and 0.1× SSC at RT for 5 min. Slides were dried by snap centrifugation and scanned immediately by using a commercially available confocal fluorescent scanner equipped with lasers operating at 532- and 635-nm wavelengths (Scan Array Lite, GSI Lumonics, Billerica, CA).