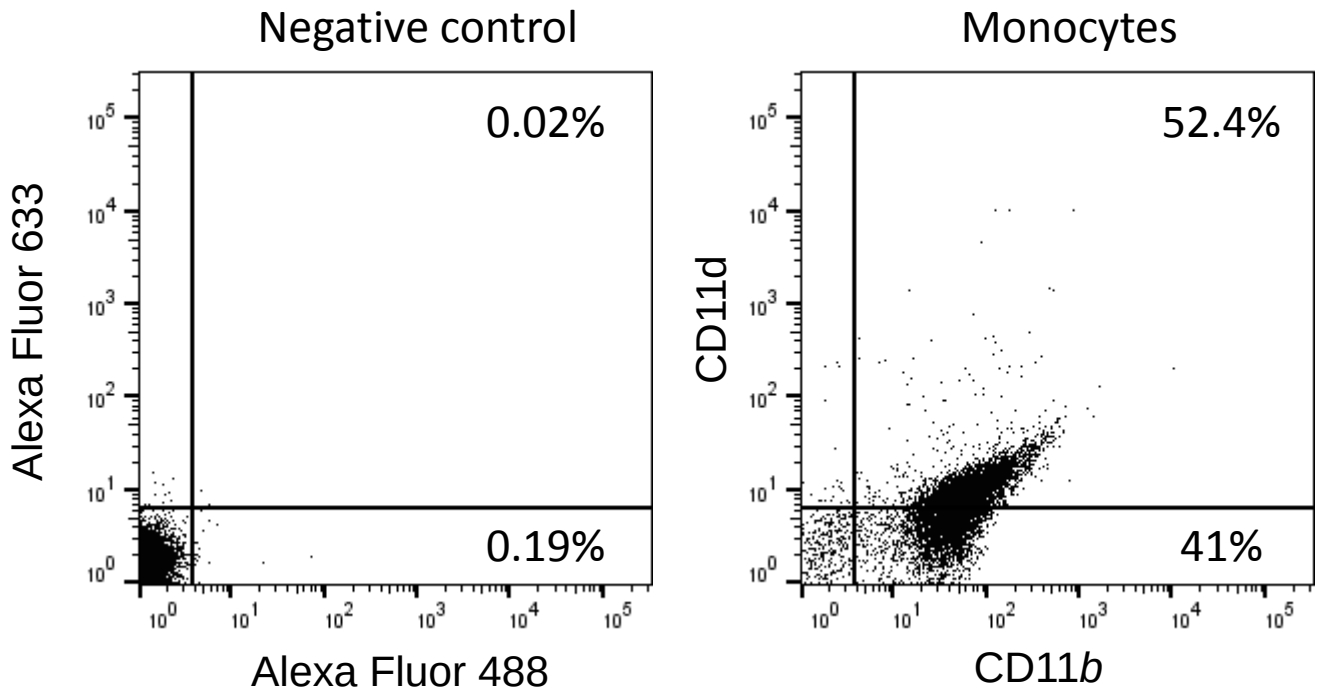
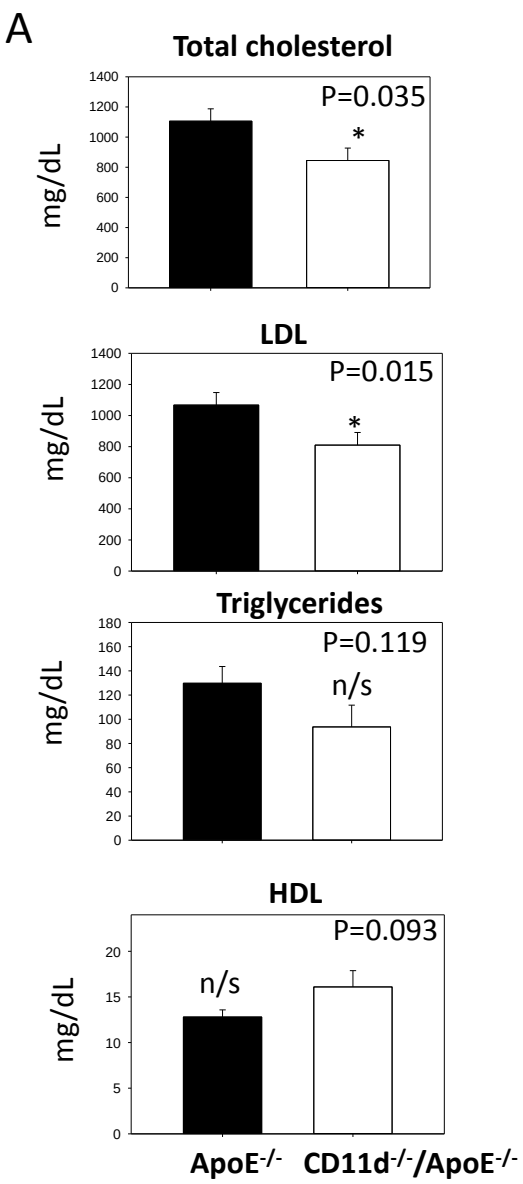
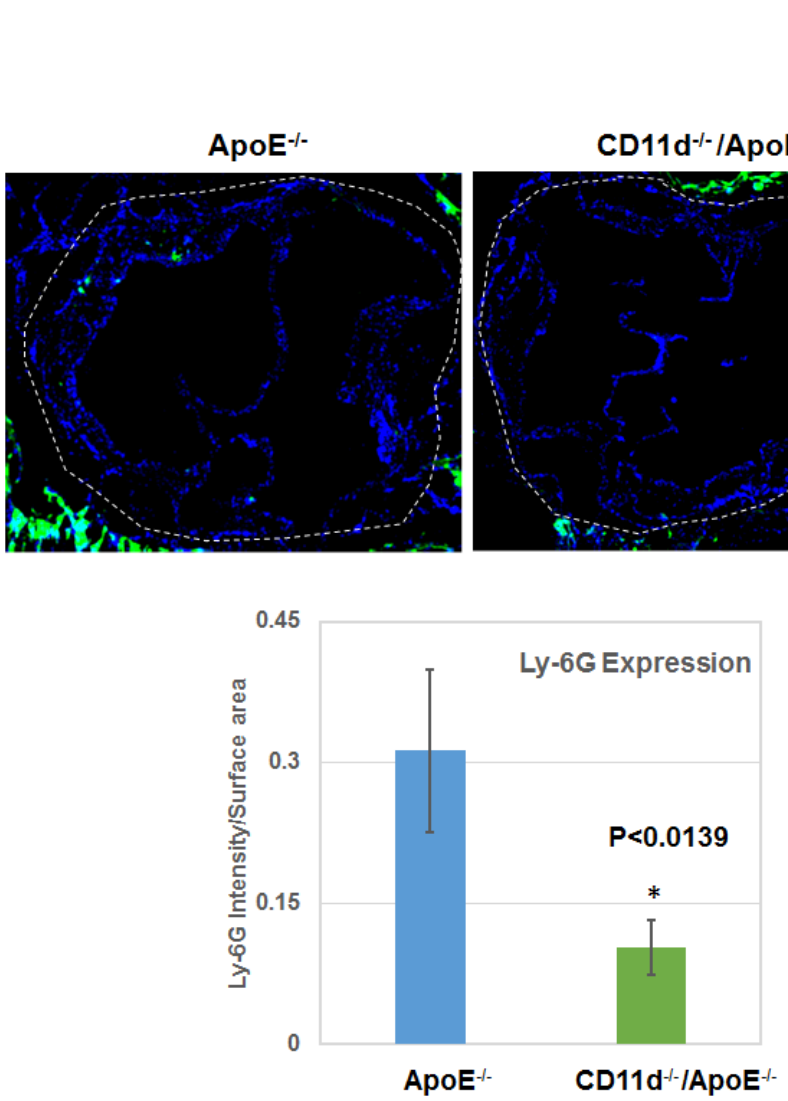
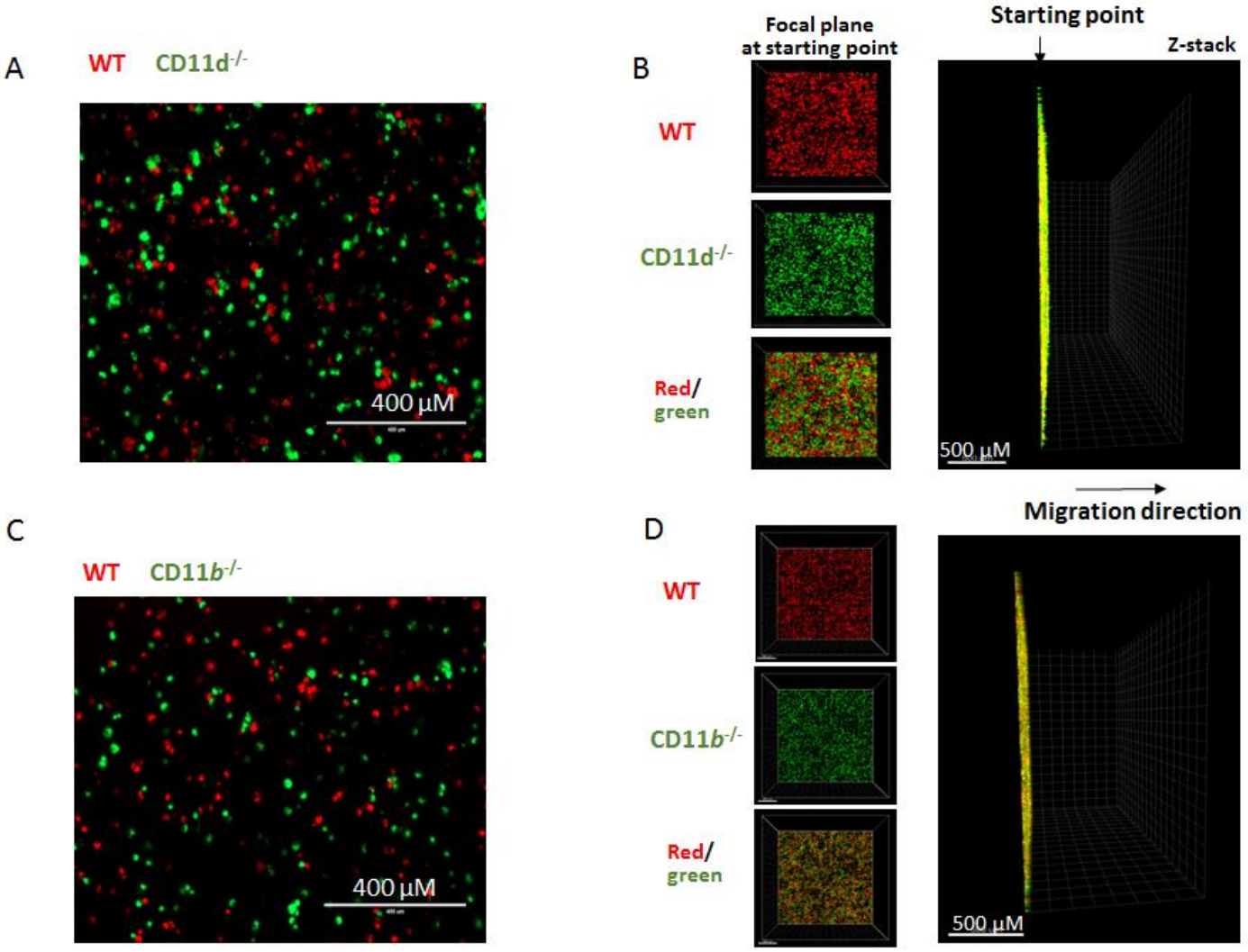


Supplemental figure I. Isolated bone marrow-derived monocytes progenitors were tested by double-staining flow cytometry with anti-CD11*b* (A488) and anti-CD11d (A633) antibodies. Cell purity (CD11*b* positive cells) is 93%. CD11d mean fluorescence is 37.8±3 MFI, which is similar to CD11d density on monocytes in circulation.





Supplemental figure II. A. The levels of total cholesterol, low density lipoprotein, high density lipoprotein and triglycerides in CD11d$^{-/-}$/ApoE$^{-/-}$ and ApoE$^{-/-}$ mice after 16 weeks Western diet were evaluated using an ABBOTT Architect CI-8200 instrument. Statistical analysis was performed using Student's *t*-test. **B.** Neutrophil Ly-6G expression in the aortic sinus section from ApoE$^{-/-}$ and CD11d$^{-/-}$/Apo$^{-/-}$ mice. Representative photographs of a cross section of the aortic sinus stained with Ly-6G (40× magnification) in the mice and the graph represents the quantification of the surface area positive for Ly-6G. The data represent the mean ± SEM Ly-6G positive area in 6 sections of each group.



Supplemental figure III. Setup for the macrophage 3-D migration in Fibrin gel. M1–activated WT and CD11d$^{-/-}$ macrophages (A,B) or WT and CD11*b*$^{-/-}$ macrophages (C, D) were labeled with PKH26 and PKH67 fluorescent dyes, respectively. Cells were mixed in equal amounts before the experiment. The similar number of cells were verified by cytospin of mixed cells (A,C) and by analysis of macrophage starting points before migration (B, C right panels). The background fluorescence of fibrin gel was verified by scanning samples with confocal microscope before the initiation of migration (B, C, left panels).

Supplemental Table 1.

Cytokine array map.

Position	Targets	Position	Targets	Position	Targets
A-1	POS*	E-1	BLANK***	I-1	Eotaxin-2 (CCL24)
A-2	POS*	E-2	BLANK***	I-2	Eotaxin-2 (CCL24)
A-3	GM-CSF	E-3	IL-2	I-3	IL-9
A-4	GM-CSF	E-4	IL-2	I-4	IL-9
A-5	IL-13	E-5	Leptin	I-5	M-CSF
A-6	IL-13	E-6	Leptin	I-6	M-CSF
A-7	RANTES (CCL5)	E-7	TIMP-1	I-7	TNF RII (TNSFRSF1A)
A-8	RANTES (CCL5)	E-8	TIMP-1	I-8	TNF RII (TNSFRSF1A)
B-1	POS*	F-1	BLC (CXCL13)	J-1	Fas Ligand (TNFSF6)
B-2	POS*	F-2	BLC (CXCL13)	J-2	Fas Ligand (TNFSF6)
B-3	IFN-gamma	F-3	IL-3	J-3	IL-10
B-4	IFN-gamma	F-4	IL-3	J-4	IL-10
B-5	IL-17A	F-5	LIX	J-5	MIG (CXCL9)
B-6	IL-17A	F-6	LIX	J-6	MIG (CXCL9)
B-7	SGF-1 alpha (CXCL12a)	F-7	TIMP-2	J-7	BLANK***
B-8	SGF-1 alpha (CXCL12a)	F-8	TIMP-2	J-8	BLANK***
C-1	NEG**	G-1	CD30 Ligand (TNFSF8)	K-1	Fractalkine (TNFSF8)
C-2	NEG**	G-2	CD30 Ligand (TNFSF8)	K-2	Fractalkine (TNFSF8)
C-3	IL-1 alpha (IL-1F1)	G-3	IL-4	K-3	IL-12 p40/p70
C-4	IL-1 alpha (IL-1F1)	G-4	IL-4	K-4	IL-12 p40/p70
C-5	I-TAC (CXCL1)	G-5	XCL1	K-5	MIP-1 alpha (CCL3)
C-6	I-TAC (CXCL1)	G-6	XCL1	K-6	MIP-1 alpha (CCL3)
C-7	I-309 (TCA-3/CCL1)	G-7	TNF alpha	K-7	BLANK***
C-8	I-309 (TCA-3/CCL1)	G-8	TNF alpha	K-8	BLANK***
D-1	NEG**	H-1	Eotaxin-1 (CCL11)	L-1	GCSF (CCL11)
D-2	NEG**	H-2	Eotaxin-1 (CCL11)	L-2	GCSF (CCL11)
D-3	IL-1 beta (IL-1F2)	H-3	IL-6	L-3	IL-12 p70
D-4	IL-1 beta (IL-1F2)	H-4	IL-6	L-4	IL-12 p70
D-5	KC (CXCL1)	H-5	MCP-1 (CCL2)	L-5	MIP-1 gamma
D-6	KC (CXCL1)	H-6	MCP-1 (CCL2)	L-6	MIP-1 gamma
D-7	TECK (CCL25)	H-7	TNF RI (TNFRSF1A)	L-7	POS*
D-8	TECK (CCL25)	H-8	TNF RI (TNFRSF1A)	L-8	POS*