1		SUPPLEM	ENTARY INFOR	RMATION	
2					
3					
4	Protein	O-mannosylation	deficiency	increases	LprG-associated
5	lipoarabin	omannan release by I	Mycobacterium	n tuberculosis	and enhances the
6	TLR2-asso	ciated inflammatory r	esponse.		
7					
8	Henar Alonso	o ¹ , Julien Parra ¹ , Wladimir N	Malaga ¹ , Delphine I	Payros ¹ , Chia-Fan	g Liu ¹ , Céline Berrone ¹ ,
9	Camille Robe	rt ¹ , Etienne Meunier ¹ , Odile	e Burlet-Schiltz ¹ , Mi	chel Rivière ^{1*} , Ch	ristophe Guilhot ^{1*}

12	Supple	mentary Figure S1: CID-MS/MS spectra of the selected mannoprotein glycopeptides.		
13	3 Supplementary Figure S2: Construction of <i>M. tuberculosis</i> mutants deficient for the production of			
14	various	s O-mannosylated proteins.		
15	A)	Schematic description of the strategy used to produce and to analyze the various mutants.		
16	B)	PCR analyses of the various <i>M. tuberculosis</i> mutants using selected couples of primers.		
17				
18	Supple	mentary Figure S3: Growth of WT, mutant and complemented strains on 7H11 solid medium		
19	A)	Serial dilutions of adjusted liquid cultures of the various mutants were spotted onto solid 7H11		
20		OADC medium and incubated at 37°C. The results showed that both the Δpmt and the $\Delta lprG$		
21		mutants were impaired in their capacity to form colonies.		
22	B)	Complementation of the $\Delta lprG$ mutant. Several constructs based on an integrative plasmid		
23		harboring either <i>lprG</i> or <i>Rv1410c</i> alone, or the <i>lprG-Rv1410c</i> operon, were transferred in the		
24		$\Delta lprG$ mutant. Complementation was evaluated as the restored capacity to form colonies on		
25		7H11 OADC plate. The results indicated that both <i>lprG</i> and <i>Rv1410c</i> were required for		
26		complementation of the $\Delta l pr G$ mutant.		
27				
28	Supple	mentary Figure S4: Growth of the bacterial population in the lung of infected mice and		
29	propor	tion of each mutant in the inoculum mix.		
30	A)	Number of cfu in the lung of infected mice after 1, 14 and 28 days of infection. Cfu were		
31		evaluated by plating serial dilution of lung homogenates on 7H11 OADC plates. The result for		
32		each mouse is indicated by a circle.		
33	B)	Relative representation of each mutant and WT strain in the inoculum used to infect mice.		

34 Total DNA was extracted from the assembled mix used to prepare the inoculum for infection

- 35 of the mice. The quantity of each mutant was evaluated by qPCR using primers specific for the
 - tags. The amount of each mutant relative to H37Rv is plotted.
 - 37

Supplementary Figure S5: Impact of LprG *O*-mannosylation on the release of the LprG/LAM complex in *M. smegmatis* Δ*pmt* or complemented with *pmt* from *M. tuberculosis*.

Western-blot analyses of LAM associated with LprG purified from the *M. smegmatis* ∆*pmt* mutant and the strain complemented with *pmt* from *M. tuberculosis*. Similar amounts of LprG-His protein purified from bacteria or culture medium were separated by SDS-PAGE. After transfer, the LAM or LprG-His molecules were revealed using anti-LAM or anti-His antibodies. Blots are representative of at least three independent experiments.

45

Supplementary Figure S6: Partial top down ETD fragmentation spectrum of the 22,541 Da molecular mass ion precursor allowing the localization of the unique hexose on the T231.

Peptide sequence of the precursor molecular ion reporting the detected fragment ions. For clarity, the
observed ions are numbered according the sequence of the parent molecular ion that starts from the
K31 of the gene-encoded sequence.

51

52 Supplementary Figure S7: Growth on 7H11 plates of the *M. tuberculosis* recombinant strains 53 producing WT LprG or LprG T231A.

54 Serial dilutions of adjusted liquid cultures of various WT, mutant, and complemented strains were 55 spotted on solid 7H11 OADC medium and incubated at 37°C. The results show that the capacity to form 56 colonies on solid medium was restored in the strains producing either the LprG WT or the LprG T231A 57 proteins.

Supplementary Figure S8: Growth of WT or *M. tuberculosis* mutants in human phagocytes and TNF α and IL-10 production by infected cells.

A) Quantification of intracellular bacterial load in hMDM infected with WT, Δpmt , $\Delta lprG$ or complemented strains. For each time point, 50 infected cells were counted and distributed in three groups: cells containing 1-3 bacteria, 4-8 bacteria or more than 8 bacteria. The plotted results are representative of two independent experiments. They indicate that the capacity of both the Δpmt and $\Delta lprG$ to multiply within hMDM was attenuated and that the $\Delta lprG$ phenotype was complemented upon expression of either the *lprG* or the *lprG** genes.

B) Inflammatory response of THP-1 infected with WT, *∆pmt* or complemented strains. Data are
 representative of two independent experiments performed in duplicate.

69

70Supplementary Figure S9: NK-κB activation in HEK-TLR2 cells incubated with LprG/LAM complexes71purified from bacterial cells or culture medium of *M. smegmatis* Δpmt or *M. smegmatis* Δpmt 72complemented with pmt from *M. tuberculosis*

HEK-TLR2 cells were incubated with 50 ng/ml or 10 ng/ml LprG-His and co-purified LAM and SEAP activity was quantified after 24h. PBS 1x or PAM₃CSK₄ (10 ng/ml) were used as negative and positive controls respectively. Data are presented as the mean +/- SD of at least five independent experiments. ** P < 0.01; *P < 0.05.

77

Supplementary Figure S10: Production of TNF- α by bone marrow-derived macrophages from WT, TLR2 -/-, or MyD88-/- mice incubated with LprG/LAM complexes purified from bacterial cells or culture medium of *M. smegmatis* Δpmt or *M. smegmatis* Δpmt complemented with *pmt* from *M. tuberculosis*

82	BMDM were incubated with 100 ng/ml of LprG-His and co-purified LAM and TNF- α was quantified
83	after 24h by ELISA. PBS 1x, LPS (100 ng/ml) or PAM ₃ CSK ₄ (100 ng/ml) were used as negative and positive
84	controls. Data are presented as the mean +/- SD of four independent experiments performed in
85	duplicate. *** P< 0.001; *P < 0.05.
86	
87	

89 Supplementary Table S1: Primers used for the construction analysis of *M. tuberculosis* mutant strains.

Construction of KO mutants					
Reference	Mutant	Primers used for recombineiring system (construction AES)	Primers used for verification	reference	
strain					
		A: AACATTGGCGACGTCAACGAC	Aout: ATTCGGTGTCAGCGGTGG		
	H37Rv	B: <u>ATC</u> TACCAGATGCCAGCTAGCCTTGTTCTGGGCACGCGTCC Bout: AATGCCAACAAGCCAGCC		This work	
	$\Delta Rv0175c$	C : <u>ATC</u> AAGTCCAACACGTCACGCTTCTTCGGGTCCGGATGGC	Cout: AACAACAATGACGGGTCG	THIS WOLK	
		D: GGTTCAAACTTACGACGGGG	Dout: ATAACCACAGCGGATGCGG		
		A: GTTCCTCAGCTCATCCGGC	Aout : TTTGGATTCGATCCCGACG		
	H37Rv	B: <u>ATC</u> TACCAGATGCCAGCTAGCCTAAAGCCCACCGGTTGCC	Bout : TTCTTGATGGGCGTCCG	This work	
	∆Rv0315	C: <u>ATC</u> AACTTCTTCAAGAGCTGCCCGGTGTTGAACCTTGCGG	Cout : AAGACCTCAACGAGCCCATCC	THIS WOLK	
		D: TATGCCTGGCGGATGATGC	Dout : TGCTGATCGGTTCGTTTTCCG		
		A : TATCCCAGGCCTGCAGTGC	Aout : TAAGCGTTCCCAGCTCCG		
	H37Rv Δ Rv0838 osis	B: TTCTACCAGATGCCAGCTAGCCTAGCCCCACCATGAGCAGACG	Bout : TCGGCGACTCGGACCAACGG	This work	
M.		C: <u>TTC</u> ATCAGCAGCAAGAACGCATTGCAACGCAGGGCGTCAG	Cout : TTCGGTCGATGTGACGTTTGC		
tuberculosis		D: AATCCACCAGCTGGTGAGCC	Dout : TATCGCCAGCCAACCGTCCG		
H37Rv		A: CGAATACCTCGACTTCCCCG	Aout : ACCGACAGCGATGCGAGCG		
		B : <u>ATC</u> TACCAGATGCCAGCTAGCCTCTTGAGAGCCTTGGTCTGCG	Bout: GGTTGGTGGTGAGATCGC	This work	
	$(\Delta l prG)$	C : <u>ATC</u> ACTCGAGCAGTGCACGTACTGGCAATTCCGTCCAGATGACC	Cout: CGATCATCAACTGGCACAGG		
		D : AACCACAGCAGCATTCCGGC	Dout: CAATCCGAGGATGTTGTGCC		
		A: TTTCCTGATCCTGTCCCGGC	Aout: TATGCAGATGCCGTGGCG		
	H37Rv	B: <u>TCC</u> TACCAGATGCCAGCTAGCCTCAATTCCCAGTGCGACGG	Bout: AGGAGTAACTGCCATCCG	This work	
	∆Rv3491	C: <u>TCC</u> ACTACGCCATGTCGTCGTCGTCATCGCAGCTCATCGGCCC	Cout:CGTTCAGCGACGAGACGC		
		D: AATTGGGTGGATCTCATGCG	Dout: ATCTATGACCAACTCAGCG		
		TTTTTT <u>TCATGA</u> TAATAATGGTTTCTTAGACG	KmFw: GACCATCAAGCATTTTATCCG		
	km ^R	AAAAAA <u>AAGCTT</u> CTGTGCTGTTGTACATGTCGATACCAGATGCCAG	KmRv: ACCGAGGCAGTTCCATAGG	This work	
		CTAGCATG			
Kanamycin	К1	GACCATCAAGCATTTTATCC			
	К2	ACCGAGGCAGTTCCATAGG		This work	
	КЗ	GCCTAGAGCAAGACGTTTCC			
	K4	AATTTAAAAGGATCTAGGTGAAGATCC			

90 Restriction sites are underlined ; Common region and specific target of each mutant used in qPCR analysis are marked in bold.

91 Supplementary Table S2: Strategy used for the construction of the various complementation plasmids

Complementation strains			
Reference Strain	Complementation	Description	reference
	H37Rv ∆lprG:: Rv1410c	Amplification of the <i>Rv1410c</i> gene with the primers AAG <u>CAGCTG</u> GTCGAGGAGG and TTTT <u>AAGCTT</u> CTTGGTCGGCACCGGC and insertion between the <i>Puv</i> II and <i>Hind</i> III sites of pMV361e Hyg integrative plasmid	This work
	H37Rv ∆lprG:: lprG	Amplification of the <i>lprG</i> gene with the primers TTTTAT <u>CAGCTG</u> GACCTCAAACCAG and TGCAGCAT <u>AAGCTT</u> GCGCC and insertion between the <i>Puv</i> II and <i>Hind</i> III sites of pMV361e Hyg integrative plasmid	This work
M. tuberculosis H37Rv ∆lprG	H37Rv ∆lprG:: lprG-Rv1410c	Amplification of the operon <i>lprG-Rv1410c</i> performed with primers TTTTAT <u>CAGCTG</u> GACCTCAAACCAG and TTTTAAGCTTATCGCCGACGCGATCTTGGTC and insertion between the <i>Puv</i> II and <i>Hind</i> III sites of pMV361e Hyg integrative plasmid	This work
	H37Rv ∆lprG:: lprG-Rv1410c gfp	Amplification of the operon <i>lprG-Rv1410c</i> performed with primers TTTTAT <u>CAGCTG</u> GACCTCAAACCAG and TTTT <u>AAGCTT</u> ATCGCCGACGCGATCTTGGTC and insertion the <i>Puv</i> II and <i>Hind</i> III sites of pMV361e Hyg gfp integrative plasmid	This work
	H37Rv ∆lprG:: lprG*-Rv1410c gfp	The punctual mutation was introduced by amplification of the whole integrative plasmid pMV361 <i>lprG-Rv1410c gfp</i> Hyg with CGAGAAGGTCCAGGTCGCGAAGCCCCCGGTGAGCTGATC GATCAGCTCACCGGGGGGCTTCGCGACCTGGACCTTCTCG	This work

94 Supplementary Table S3: Primers used for the qPCR analysis

Name	Sequence
Rv0175c variable	GTCTTCTCGACCACCACAGAT
Rv0315 variable	GGGCAGCTCTTGAAGAAGTT
Rv0838 variable	ATGCGTTCTTGCTGCTGAT
Rv3491 variable	GACGACGACATGGCGTAGT
Rv1411c variable	AGTACGTGCACTGCTCGAGT
qPCR km Fw	CGATGAGTTTTTCTAACTGTCAGACC
qKmFint	TGATGCGCTGGCAGTGTT
qKmRint (commonR)	CGCGATCGCTGTTAAAAGGA

MS/MS Fragmentation of DCVAATQAPDAGAMSASMQK

Found in RVBD_0175T.1 in MTBtuberculist_R27, RVBD_0175T.1|RVBD_0175|Mycobacterium tuberculosis H37Rv (Broad V3) MCE-associated membrane protein (214 aa)

Match to Query 77813: 2170.921448 from(1086.468000,2+) index(78042)

Variable modifications:

C2 : Carbamidomethyl (C) S17 : HexINL (ST), with neutral losses 162.052823(shown in table), 0.000000 Ions Score: 121 Expect: 6.9e-012



MS/MS Fragmentation of PAGPTPAPAAPAAATGGLLFHDEFDGPAGSVPDPSK Found in RVBD_0315T.1 in MTBtuberculist_R27, RVBD_0315T.1|RVBD_0315|Mycobacterium tuberculosis H37Rv (Broad V3) hypothetical protein (295 aa)

Match to Query 15435: 3544.704495 from(1182.575441,3+) index(2655) Monoisotopic mass of neutral peptide Mr(calc): 3544.699829 Variable modifications:

S35 : Hex(1)NLI(ST) (ST), with neutral loss 162.052823



MS/MS Fragmentation of VQVTKPPVS Found in RVBD_1411cT.1 in MTBtuberculist_R27, RVBD_1411cT.1|RVBD_1411c|Mycobacterium tuberculosis H37Rv (Broad V3) lipoprotein lprG (237 aa)

Match to Query 16700: 1115.607248 from(558.810900,2+) index(80564)

Monoisotopic mass of neutral peptide Mr(calc): 1115.607422

Variable modifications:

T4 : HexINL (ST), with neutral losses 0.000000(shown in table), 162.052823 Ions Score: 25 Expect: 0.039 (help)



MS/MS Fragmentation of TTTPSGPVPPVSEAAR Found in RVBD_0838T.1 in MTBtuberculist_R27, RVBD_0838T.1|RVBD_0838|Mycobacterium tuberculosis H37Rv (Broad V3) D-alanyl-Dalanine dipeptidase (257 aa)

Match to Query 70844: 1889.908848 from(945.961700,2+) index(55551)

Monoisotopic mass of neutral peptide Mr(calc): 1889.910599 Variable modifications:

S5 : HexINL (ST), with neutral losses 162.052823(shown in table)
S12 : HexINL (ST), with neutral losses 162.052823(shown in table)
Ions Score: 55 Expect: 1.4e-005



MS/MS Fragmentation of QPFSLQLIGPPPSPVQR Found in Rv3491_Rv3491 in MTBtuberculist_R27, Rv3491_Rv3491

Match to Query 75159: 2022.089448 from(1012.052000,2+) index(76792)

Monoisotopic mass of neutral peptide Mr(calc): 2022.078598 Variable modifications:

S13 : HexINL (ST), with neutral losses 162.052823(shown in table)
Ions Score: 49 Expect: 3.4e-005



Figure S1



A)

B)





dil.

B)



Figure S3







Figure S4



Figure S5



Figure S6



Figure S7



B)





Figure S9

