

**Mechanisms of Uranium and Chromium Toxicity in *Pseudomonas stutzeri* RCH2 Grown  
Under Anaerobic Nitrate-Reducing Conditions**

Michael P. Thorgersen<sup>a</sup>, W. Andrew Lancaster<sup>a</sup>, Xiaoxuan Ge<sup>a</sup>, Grant M. Zane<sup>b</sup>, Kelly M.  
Wetmore<sup>c</sup>, Brian J. Vaccaro<sup>a</sup>, Farris L. Poole<sup>a</sup>, Adam Younkin<sup>b</sup>, Adam M. Deutschbauer<sup>c</sup>,  
Adam P. Arkin<sup>c</sup>, Judy D. Wall<sup>b</sup> and Michael W. W. Adams<sup>a#</sup>

<sup>a</sup>Department of Biochemistry & Molecular Biology, University of Georgia, Athens, GA, 30602

<sup>b</sup>Department of Biochemistry, University of Missouri, Columbia, Mo 65211

<sup>c</sup>Environmental Genomics and Systems Biology Division, Lawrence Berkeley National  
Laboratory, Berkeley, CA 94720

Running Title: Anaerobic U and Cr Toxicity

**Supplementary Material**

**Tables S1 – S4**

**Figures S1 – S3**

**Table S1. Quality metrics for RB-TNSeq data**

Condition	Median SE	SE Min	SE Max	MAD12	Average Reads/Gene
Control	0.148	0.002	1.7	0.26	1494
120 $\mu$ M K <sub>2</sub> Cr <sub>2</sub> O <sub>7</sub>	0.154	0.003	1.6	0.27	1378
3 mM Uranyl Acetate	0.152	0.002	2.0	0.26	1475

Standard error (SE) was calculated for each gene under all three conditions. The MAD12 is the median absolute difference {Wetmore, 2015}. The number of reads for each gene from all sequencing runs within a condition were summed and the average value is reported (Average Reads/Gene).

**Table S2. Table of primers used for construction of marker-exchange plasmid in this study.**

Primer Name	Primer Sequence*
Psest_2088-upF	<b>CGGCTGCAACTTTGTCATGCTT</b> GCGCTA TAGCTTCGGTGAGGTGC
Psest_2088-upR	<b>TGCCTTCTTGACGAGTTCTTCTGACGTT</b> CTTAATGATTCGCTGCATAGACGC
Psest_2088-dnF	<b>GCGCCCCAGCTGGCAATTCCGGG</b> GCTTCA TTGATACATCCAGAAATTAGGGTGTTCCG
Psest_2088-dnR	<b>CGAGGCATTTCTGTCTGGCTGGGGTAC</b> TCATTTCCAGGCTAGCGTTCTG
SpecRpUC-F	CCAGCCAGGACAGAAATGCCTCG
SpUrT-R	CAAGCATGACAAAGTTGCAGCCG
Kan gene Prom Nterm	CCGGAATTGCCAGCTGGGGCGC
KanR	TCAGAAGAAGTCAAGGCGA
oriT-out	CCTCGTTCAACAGGTCCAGGG
pMO719-XbaI-dn	TGGGTTCGTGCCTTCATCCG
Km_int_Fwd_revcomp	CTCATCCTGTCTCTTGATCAGATCT
Kan_out_C	CTGACCGCTTCTCGTGC

\* Bolded region of primers are overhang used for cloning with plasmid backbone or kanamycin-resistance marker.

**Table S3. Genes with  $\Delta wCr \geq 1$** 

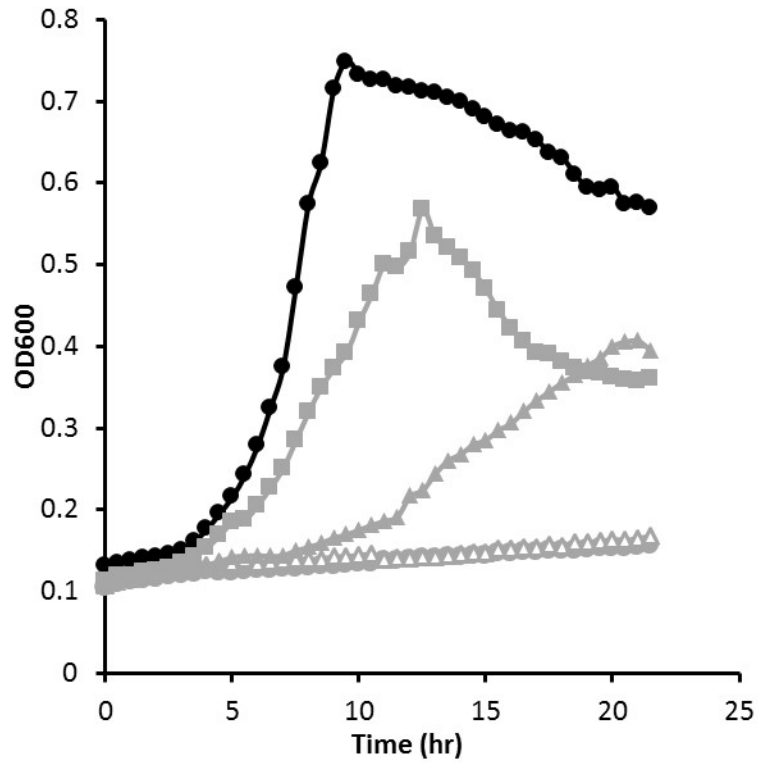
Locus Tag	Gene Function	$w_{ctrl}$	$w_{ctrl}$	$wCr$	$wCr$	$\Delta wCr$
		AVE	STDEV	AVE	STDEV	Delta
<b>DNA Repair</b>						
Psest_3100	RNA polymerase sigma factor RpoE	0.3	0.1	1.3	0.2	1.0
<b>Sulfur Assimilation</b>						
Psest_0186	Protein-disulfide isomerase	-0.5	0.4	0.4	0.3	1.0
<b>Fe Related</b>						
Psest_0954	Fe-S oxidoreductase	-2.2	0.3	0.4	0.1	2.6
Psest_0953	Iron-sulfur cluster-binding protein	-2.2	0.2	0.2	0.3	2.4
Psest_1463	Fe-S center assembly, SufE	-0.6	1.0	0.5	0.6	1.2
<b>Hypothetical</b>						
Psest_3489	Hypothetical protein	-1.3	0.0	1.9	0.2	3.2
Psest_0952	Uncharacterized conserved protein	-2.7	0.5	0.2	0.5	2.8
Psest_4003	TIGR00153 family protein	-1.4	0.5	0.6	0.4	2.1
Psest_2385	Hypothetical protein	-0.6	0.7	0.5	1.2	1.1
<b>Other</b>						
Psest_3490	Signal transduction histidine kinase	-1.2	0.9	1.5	0.6	2.7
Psest_0999	Response regulator	-0.7	0.5	1.0	0.6	1.7
Psest_0951	FAD/FMN-containing dehydrogenases	-0.6	0.2	1.0	0.1	1.6
Psest_2418	Diguanylate cyclase (GGDEF) domain	-0.7	0.4	0.9	0.4	1.6
Psest_3042	Nitrate reductase cytochrome c-type , NapB	-0.6	0.2	0.8	1.3	1.4
Psest_3573	dTDP-4-dehydrorhamnose reductase	0.0	0.5	1.4	0.3	1.4
Psest_1974	Integration host factor, IhfA	-1.0	0.3	0.4	0.0	1.4
Psest_0092	Phosphonate C-P lyase system protein PhnK	-0.9	0.6	0.5	0.3	1.4
Psest_2590	3-isopropylmalate dehydratase, small subunit	-0.9	0.5	0.4	0.2	1.3
Psest_2988	Aspartate kinase, monofunctional class	-4.0	0.7	-2.7	0.3	1.3
Psest_0722	Predicted ATPase	-0.9	0.4	0.3	0.6	1.2
Psest_0466	Lipopolysaccharide kinase (Kdo/WaaP)	-1.2	0.5	0.0	0.2	1.2
Psest_0839	cAMP-binding proteins	-0.1	0.2	1.1	0.2	1.2
Psest_2023	Isocitrate dehydrogenase	-1.3	1.2	-0.1	0.3	1.2
Psest_3323	Ribonuclease, Rne/Rng family	-0.8	0.2	0.4	0.5	1.2
Psest_3377	FKBP-type prolyl cis-trans isomerases	-0.1	0.9	1.0	1.0	1.1
Psest_3226	Predicted membrane protein	-0.6	0.5	0.5	1.1	1.1
Psest_3684	Di- and tricarboxylate transporters	-0.2	0.2	0.8	0.1	1.1
Psest_0277	Non-canonical purine NTP pyrophosphatase	-0.3	0.6	0.7	0.3	1.0
Psest_0575	HrpA-like helicases	-0.8	0.2	0.2	1.1	1.0
Psest_4287	Predicted glutamine amidotransferases	-0.2	0.2	0.8	0.4	1.0
Psest_2123	Carbohydrate kinase	-0.8	0.4	0.2	0.3	1.0
Psest_1994	Amidases related to nicotinamidase	0.1	0.1	1.1	0.2	1.0
Psest_1724	Anti-anti-sigma regulatory factor	-1.5	0.4	-0.5	0.3	1.0
Psest_2949	Aspartyl aminopeptidase	0.0	0.3	1.0	0.2	1.0

Psest_0800	Dehydrogenases	-0.1	0.1	0.9	0.0	1.0
Psest_3047	Methionyl-tRNA synthetase	-0.2	0.6	0.8	1.0	1.0
Psest_0393	Methylase of chemotaxis methyl-accepting	-2.2	0.4	-1.2	0.2	1.0
Psest_0325	Putative threonine efflux protein	0.2	0.1	1.2	0.1	1.0

**Table S4. Genes with  $\Delta wU \geq 1$**

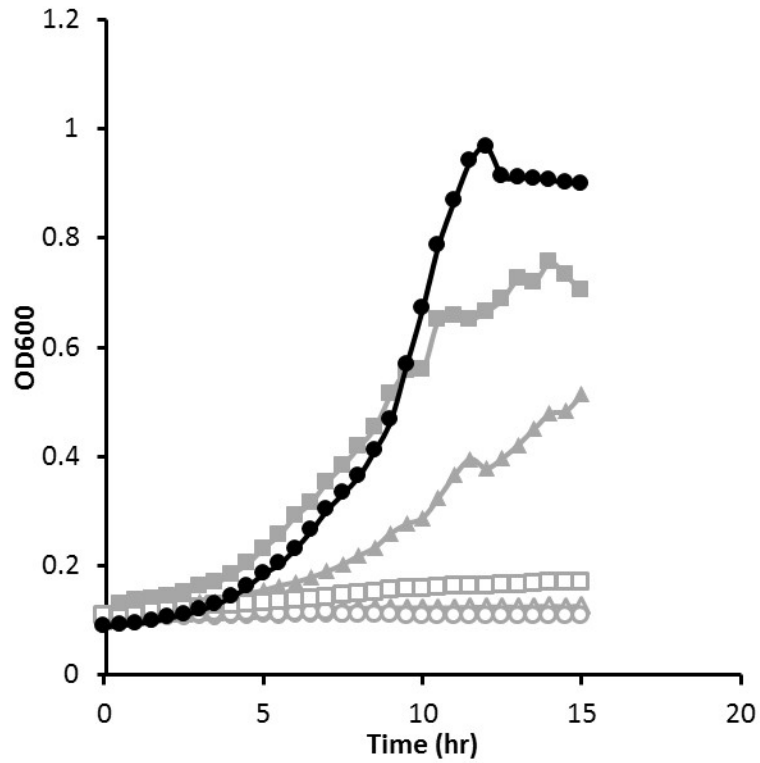
Locus Tag	Gene Function	$w_{ctrl}$	$w_{ctrl}$	$wU$	$wU$	$\Delta wU$
		AVE	STDEV	AVE	STDEV	Delta
<b>Fe Related</b>						
Psest_0954	Fe-S oxidoreductase	-2.2	0.3	-0.7	0.2	1.5
Psest_1463	Fe-S center assembly, SufE	-0.6	1.0	0.8	1.0	1.4
Psest_0953	Iron-sulfur cluster-binding protein	-2.2	0.2	-0.8	0.3	1.4
<b>Hypothetical</b>						
Psest_0952	Uncharacterized conserved protein	-2.7	0.5	-0.8	0.5	1.9
Psest_3146	Hypothetical protein	-0.9	0.4	1.0	0.5	1.8
Psest_1920	Uncharacterized protein conserved in bacteria	-0.9	0.5	0.6	1.1	1.5
<b>Other</b>						
Psest_2243	Glyceraldehyde-3-phosphate dehydrogenase	-0.4	0.8	0.7	2.0	1.2
Psest_1082	Enoyl-CoA hydratase/carnithine racemase	-1.5	0.3	-0.4	0.3	1.1

Figure S1



Anaerobic growth of *Pseudomonas stutzeri* RCH2 without (black) and with (gray) the addition of 80  $\mu\text{M}$   $\text{K}_2\text{Cr}_2\text{O}_7$  to the growth medium (not containing yeast extract). Various exogenously added sulfur sources include: none (closed circles), 0.1 mM sulfate (open triangles), 1 mM sulfate (closed triangles), and 0.1 mM sulfide (closed squares).

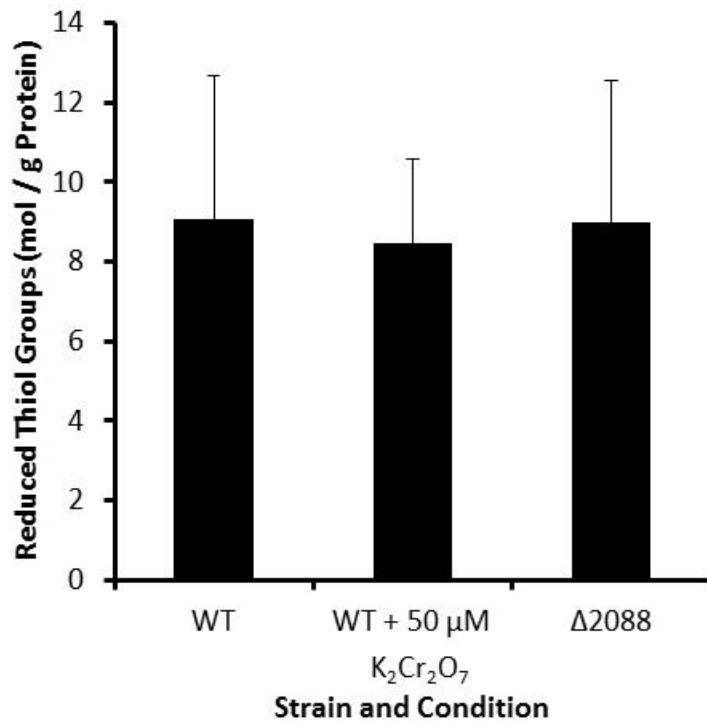
Figure S2



**Aerobic growth of *Pseudomonas stutzeri* RCH2 WT (black) and  $\Delta$ 2088 (gray) without yeast extract (closed circles) and with various exogenously added sulfur sources: 1 mM sulfate (open triangles), 1 mM sulfite (open circles), 0.3 mM sulfide (closed squares), 0.3 mM cysteine (open squares), and 0.1 mM thiosulfate (closed triangles).**



Figure S3



Reduced intracellular thiol concentrations of cell free extracts prepared anaerobically from anaerobically grown cultures of *Pseudomonas stutzeri* RCH2.