

## **TITLE**

A Tissue Engineered Blood Vessel Model of Hutchinson-Gilford Progeria Syndrome Using Human iPSC-derived Smooth Muscle Cells

## **AUTHORS AND AFFILIATIONS**

Leigh Atchison<sup>1</sup>, Haoyue Zhang<sup>2</sup>, Kan Cao<sup>2</sup>, George A. Truskey<sup>1\*</sup>

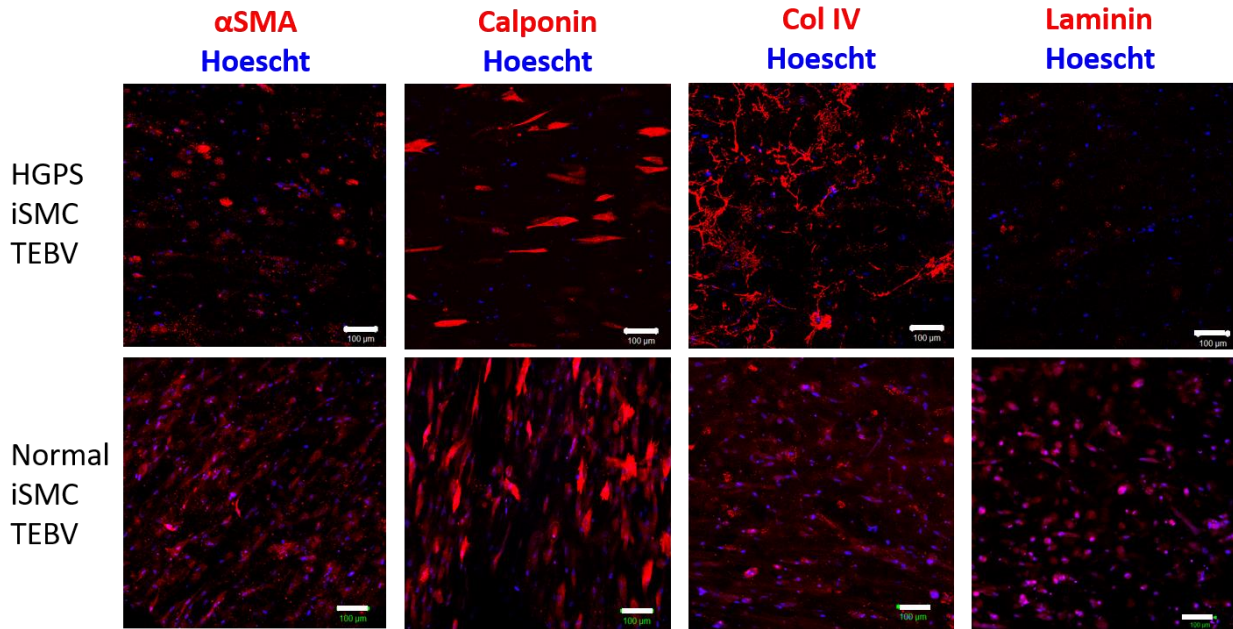
<sup>1</sup>Department of Biomedical Engineering at Duke University, Durham, NC 27708

<sup>2</sup>Department of Cell Biology and Molecular Genetics at University of Maryland, College Park, MD 20742

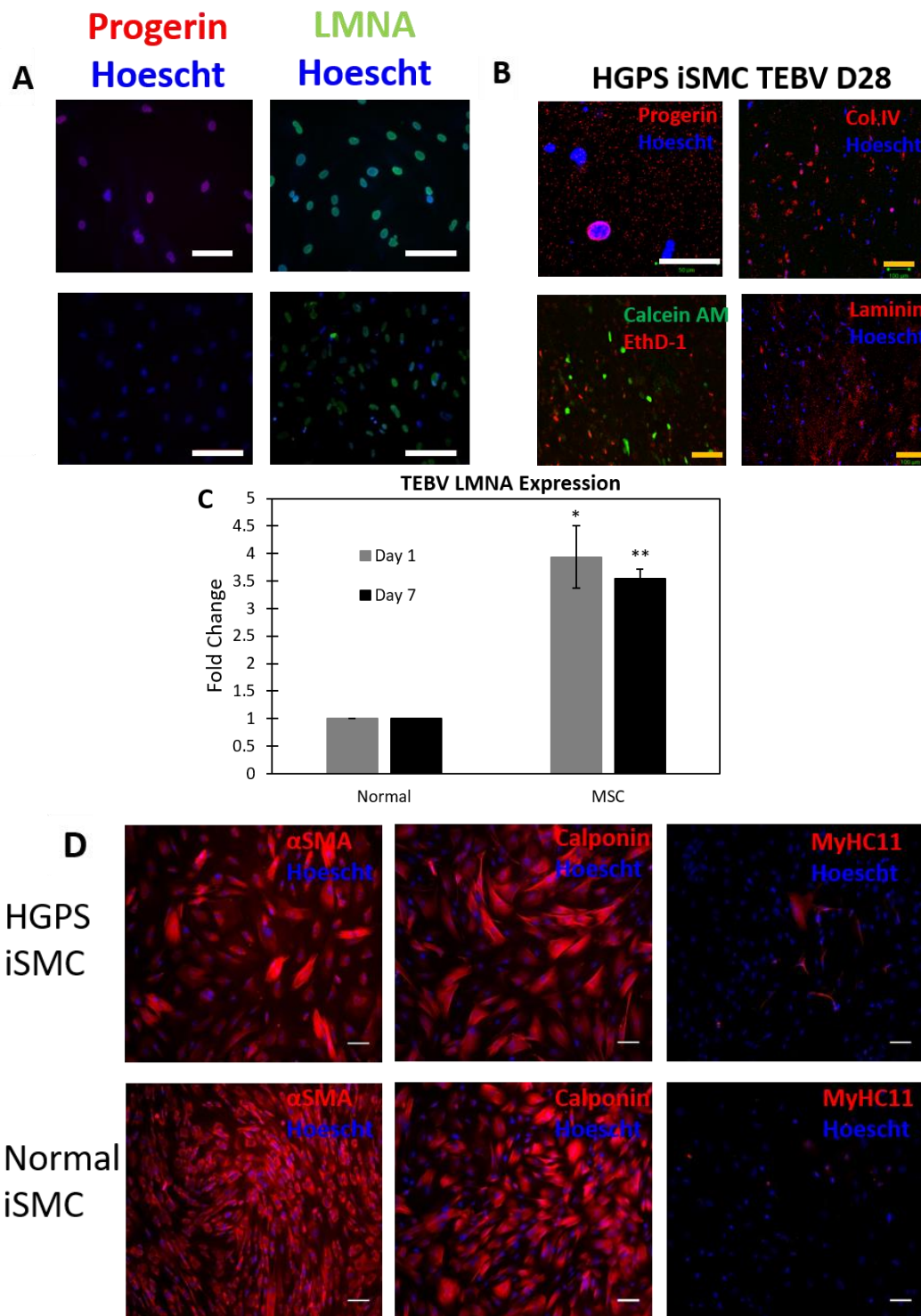
## **CORRESPONDING AUTHOR**

Dr. George A. Truskey (919-660-5147, [george.truskey@duke.edu](mailto:george.truskey@duke.edu)) R. Eugene and Susie E. Goodson  
Professor of Biomedical Engineering Duke University, Durham, NC 27708

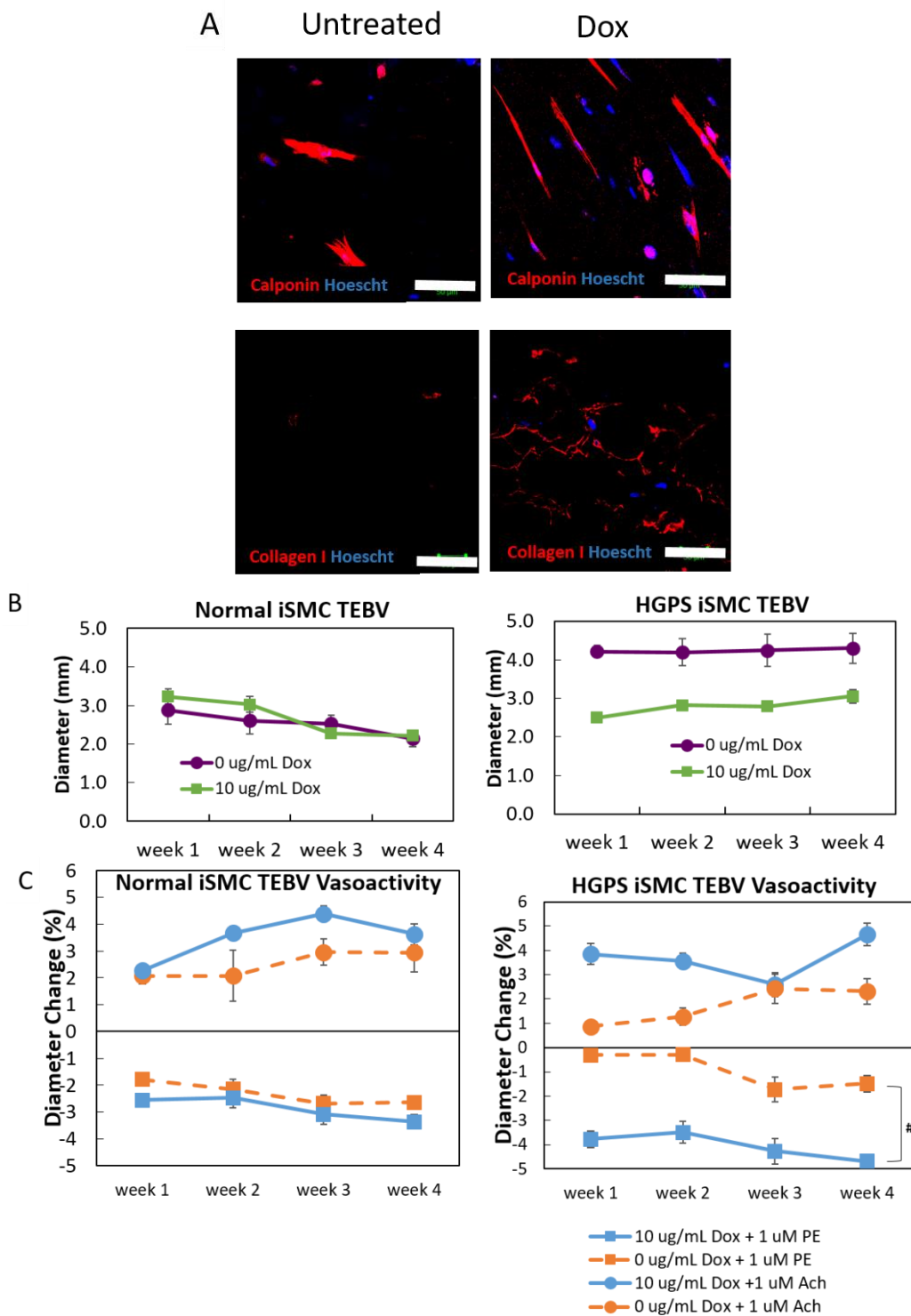
## SUPPLEMENTARY FIGURES



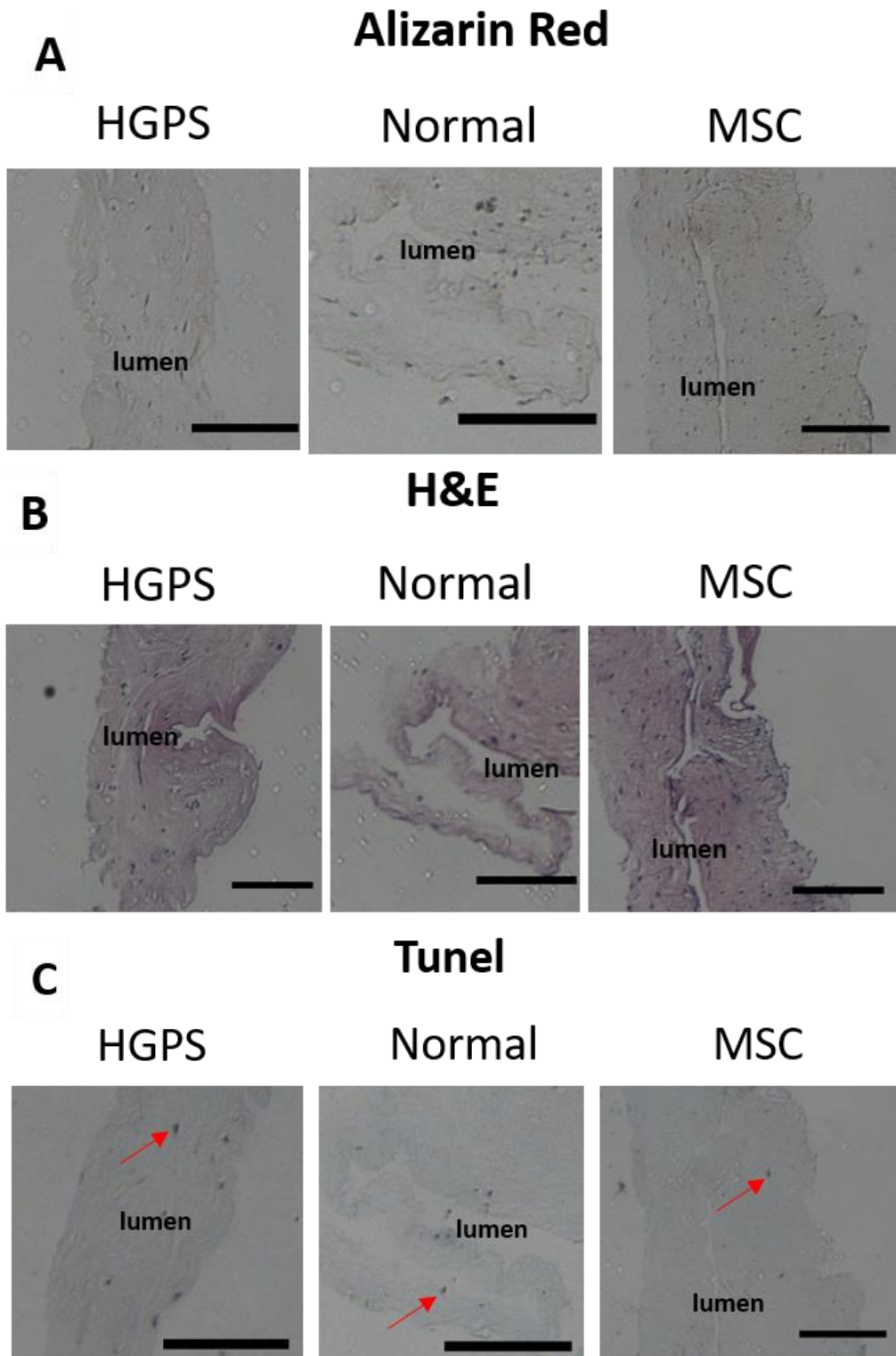
**Figure S1:** Protein characterization of iSMC TEBVs at Day 7. Representative images of immunofluorescent stains with (A) anti-alpha smooth muscle actin, (B) calponin, (C) collagen IV and (D) laminin on both normal and HGPS iSMC TEBVs after 1 week of perfusion culture (Scale bar, 100  $\mu$ m)



**Figure S2:** (A) Representative immunofluorescent images with anti-lamin A/C and progerin antibodies of normal and HGPS iSMC after 14 days in 2D culture (Scale bar, 100  $\mu$ m). (B) Representative immunofluorescent images of HGPS iSMC TEBVs after 28 days of perfusion with anti-progerin, anti-collagen IV, and anti-laminin antibodies and Calcein AM and EthD-1 (white scale bar = 50  $\mu$ m, yellow scale bar = 100  $\mu$ m). (C) qRT-PCR of Lamin A/C gene expression on HGPS and normal iSMC TEBVs at D1 and D7 of perfusion culture normalized to HGPS iSMC TEBVs; n=3 TEBVs, \*P<.05, \*\*P<.01. (D) Representative immunofluorescent images in 2D culture, prior to formation of TEBVs, of HGPS and normal iSMCs with anti-alpha smooth muscle actin, anti-calponin and anti-smooth muscle myosin heavy chain 11 (scale bar= 100  $\mu$ m).



**Figure S3:** (A) Representative immunofluorescent images of normal iSMC TEBVs after four weeks of perfusion with 10  $\mu\text{g}/\text{mL}$  doxycycline or untreated and stained with anti-calponin and anti-collagen I (scale bar=100  $\mu\text{m}$ ). (B) Weekly outer diameter measurements of normal iSMC TEBVs seeded with hCB-EPCs or HGPS iSMC TEBVs seeded with hCB-EPCs for four weeks. (C) Weekly vasoactive response to 1  $\mu\text{M}$  phenylephrine or 1  $\mu\text{M}$  acetylcholine of normal iSMC TEBVs seeded with hCB-EPCs or HGPS iSMC TEBVs seeded with hCB-EPCs for four weeks.  $n=3$  TEBVs,  $\#P<.0001$  at each week.



**Figure S4:** Histochemical analysis of iSMC or MSC TEBVs at Day 7. (A) Alizarin Red stain (B) H&E stain and (C) Tunel stain of HGPS iSMC, normal iSMC or MSC TEBVs after 1 week of perfusion culture (Scale bar, 200  $\mu$ m).

<b>Cell Type</b>	<b>Source</b>	<b>Cell Line</b>	<b>Donor Age</b>	<b>Passage Range</b>
<b>MSC</b>	<b>Texas A&amp;M Institute for Regenerative Medicine</b>	<b>N/A</b>	<b>unknown</b>	<b>p5-p8</b>
<b>Normal iSMC</b>	<b>Progeria Research Foundation</b>	<b>HGADFN167</b>	<b>8 yrs 5 months</b>	<b>p1-p3</b>
<b>HGPS iSMC</b>	<b>Progeria Research Foundation</b>	<b>HGFDFN168</b>	<b>40 yrs 5 months</b>	<b>p1-p5</b>
<b>hCB-EPCs</b>	<b>Carolina Cord Blood Bank</b>	<b>N/A</b>	<b>newborn</b>	<b>p5-p8</b>

**Table S1:** TEBV cell sources