

1 **Modified Vaccinia Virus Ankara Preferentially Targets**
2 **Antigen Presenting Cells *In Vitro*, *Ex Vivo* and *In Vivo***

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24 [MOVIE]

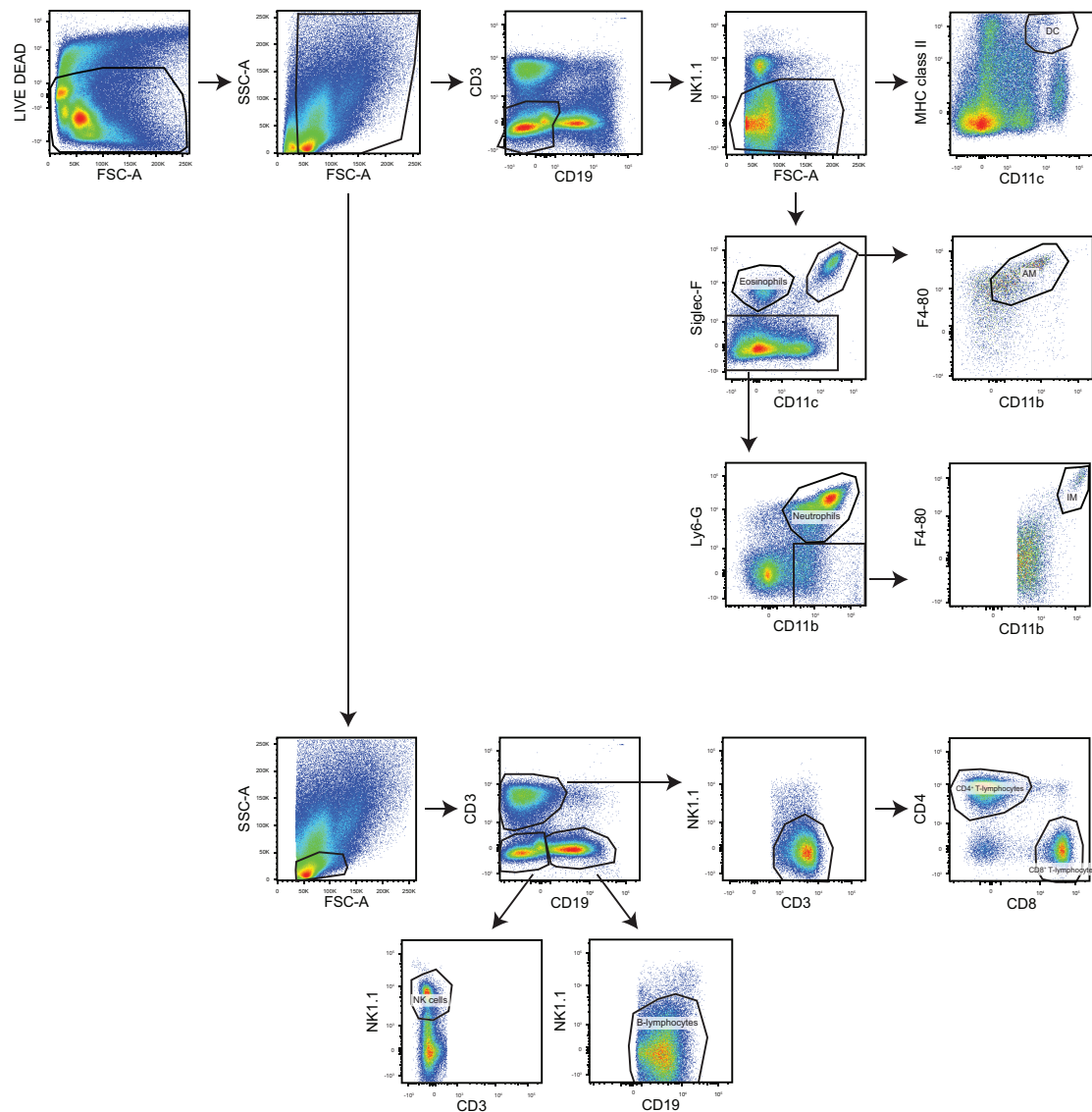
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26 **Supplementary Figure 1. CLSM 3D render of mouse hind leg muscle.** Z-

27 stack of a hind leg muscle slice from a IM rMVA-GFP injected mouse was

28 obtained by CLSM. A 3D render of the maximum intensity projection was

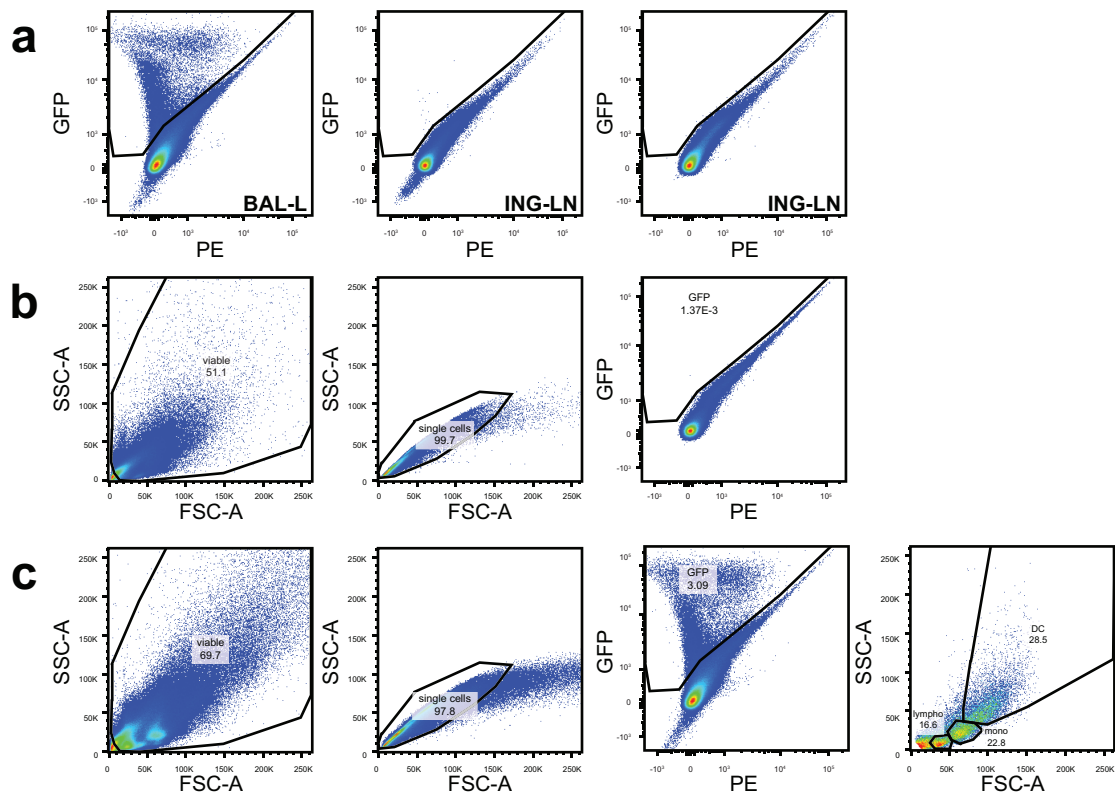
29 generated using the Zen software. GFP = green. Nucleus = red.



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31 **Supplementary Figure 2. Gating strategy to define populations for**
 32 **phenotypic analysis of GFP⁺ cells in the lungs of mice.** Live cells were
 33 gated followed by selection of non-lymphocytes and lymphocytes based on
 34 the forward / sideward scatter. Subsequently, CD3⁻ CD19⁻ NK1.1⁻ cells were
 35 selected in the non-lymphocyte population. MHC class II⁺ CD11c⁺ cells were
 36 defined as DC and Siglec-8⁺ CD11c⁻ were classified as eosinophils. Siglec-8⁺
 37 CD11c⁺ F4-80⁺ CD11b⁺ cells were identified as alveolar macrophages (AM).
 38 Siglec-8-negative cells were further subdivided into a Ly6-G⁺ CD11b⁺
 39 neutrophil population and Ly6-G⁻ CD11b⁺ F4-80⁺ interstitial macrophages (IM).

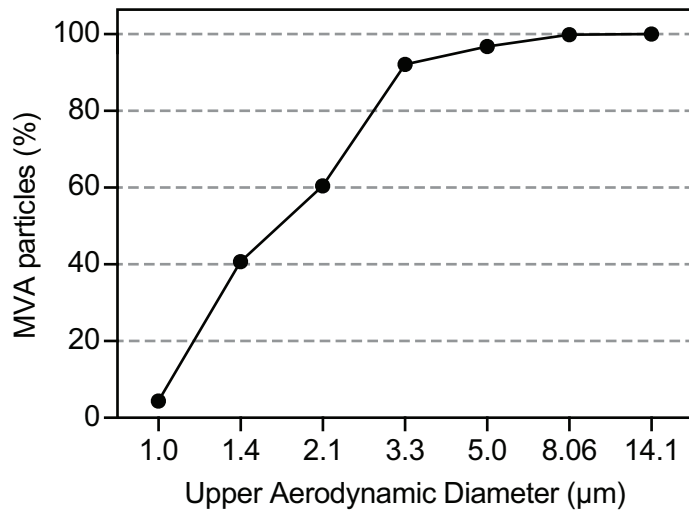
40 Following selection of lymphocytes in the forward / sideward scatterplot, CD3⁺
41 CD19⁻ NK1.1⁻ cells were selected and divided into CD4⁺ and CD8⁺ T-
42 lymphocytes. Furthermore, CD3⁻ CD19⁻ NK1.1⁺ cells were defined as NK cells
43 and CD3⁻ CD19⁺ NK1.1⁻ cells were defined as B-lymphocytes.



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45 **Supplementary Figure 3. Gating strategy to detect GFP⁺ cells in ferret**

46 **tissues. (A)** Example of detection of GFP⁺ cells in unstained single cell
 47 suspensions from different ferret tissues. Detection of GFP⁺ cells in a sample
 48 containing abundant GFP⁺ cells (panel 1, BAL left side after IT inoculation),
 49 background level GFP⁺ cells (panel 2, ING-LN after IM injection) or no GFP⁺
 50 cells (panel 3, ING-LN after IT inoculation) is shown. **(B-C)** Gating strategy to
 51 define GFP⁺ DC-like, monocyte-like and lymphocyte-like cell populations in
 52 ferret tissues. As an example, the gating strategy of BAL is shown after IM
 53 injection **(B, negative control)** or IT inoculation **(C)** with rMVA-GFP. First,
 54 viable cells were gated followed by selection of single cells. Next, all GFP⁺
 55 cells were selected after which lymphocyte-like, monocyte-like or DC-like cell
 56 populations were reversely gated in the scatter plot. Gate name and
 57 percentage of events are indicated in each gate.



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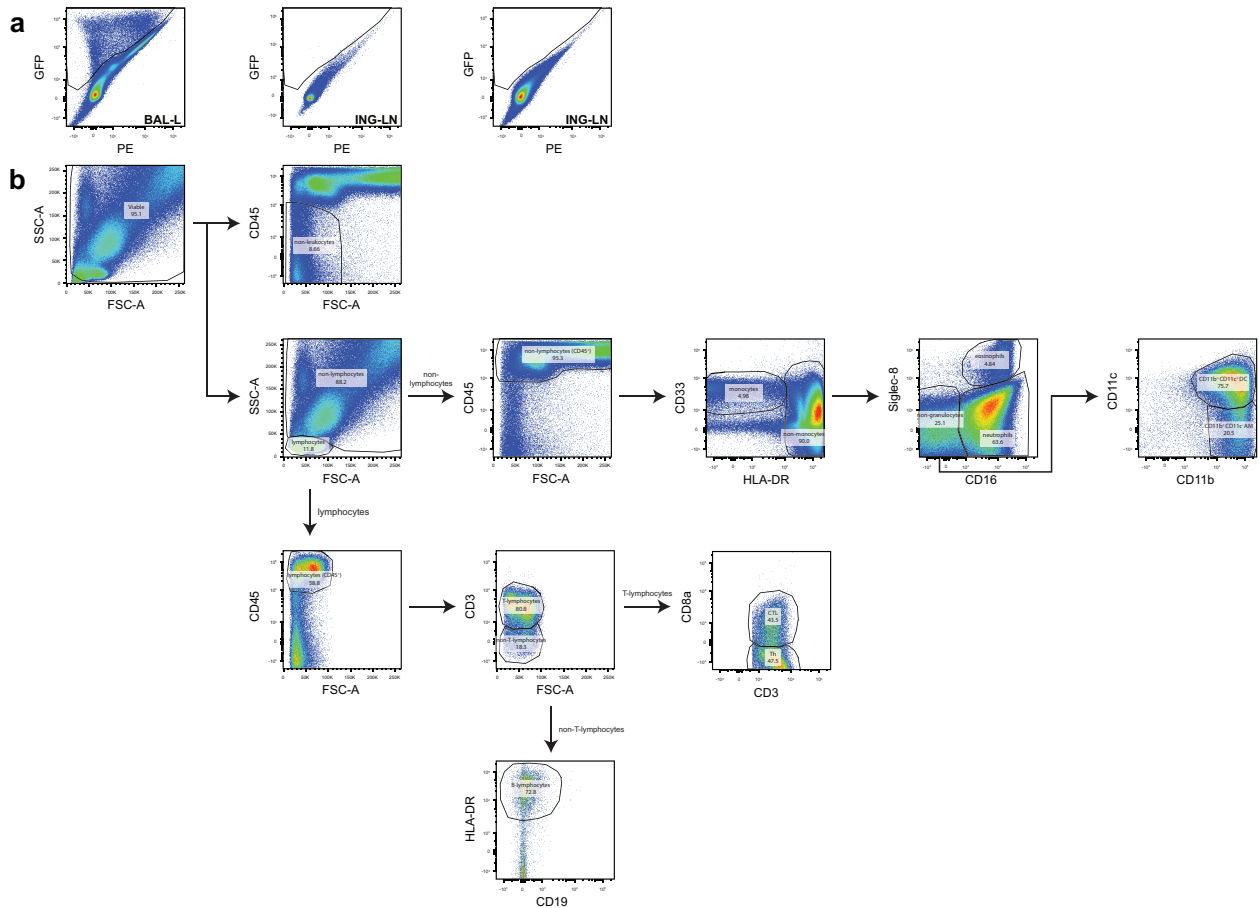
59 **Supplementary Figure 4. Droplet size characterization of rMVA-GFP**

60 **aerosol.** rMVA-GFP was nebulized and several droplet fractions of increasing
61 size between 0.98 and 14.1µm in diameter were collected using a cascade
62 impactor. Cumulative distribution of rMVA-GFP particles across the range of
63 droplet diameters is shown.

64 [MOVIE]

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66 **Supplementary Figure 5. CLSM 3D render of macaque lung.** Z-stack was
67 obtained of a lung slice from a macaque that received rMVA-GFP via AER
68 inhalation. A 3D render of the maximum intensity projection was generated
69 using Zen software. GFP = green. Nucleus = red.



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71 **Supplementary Figure 6. Gating strategy to detect GFP⁺ cells in macaque tissues.**

72 (A) Example of detection of GFP⁺ cells in unstained single cell suspensions from
 73 different non-human primate tissues. Detection of GFP⁺ cells in a sample containing
 74 abundant GFP⁺ cells (panel 1, BAL left side after AER inhalation), background level
 75 GFP⁺ cells (panel 2, ING-LN after IM injection) or no GFP⁺ cells (panel 3, ING-LN after
 76 IT inoculation) is shown. (B) Gating strategy to define populations for phenotypic
 77 analysis of GFP⁺ cells in BAL of macaques. Viable cells were selected after which non-
 78 leukocyte (e.g. CD45⁻ epithelial cells) were gated. Viable cells were further discriminated
 79 into lymphocytes and non-lymphocytes based on FSC / SSC plot. CD45⁺ cells outside
 80 the lymphogate were selected in which the CD33⁺ monocytes were gated. The HLA-

81 DR⁺ population was divided into Siglec-8⁺ CD16⁺ eosinophils, Siglec-8⁻ CD16⁺
82 neutrophils and Siglec-8⁻ CD16⁻ non-granulocytes, which were further phenotyped into
83 CD11b⁺ CD11c⁺ DC or CD11b⁺ CD11c⁻ AM. The CD45⁺ lymphocytes were divided into
84 CD3⁺ T-lymphocytes, further discriminated into CD8⁺ cytotoxic T-lymphocytes (CTL)
85 and CD8⁻ T helper (Th)-lymphocytes, and CD3⁻ HLA-DR⁺ B-lymphocytes. In this
86 example CD19 staining did not work, thus was not included in the analysis. Gate name
87 and percentage of events is indicated in each gate.