1	Supplementary Information
2	for
3	Response of microbial community function to fluctuating
4	geochemical conditions within a legacy radioactive waste trench
5	environment
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20 Results and Discussion

21 Reactive oxygen species detoxification

The rapid trench water redox cycling involving the production of large quantities of 22 Fe(II), along with elevated concentrations of organic compounds, creates conditions 23 conducive to reactive oxygen species production (Page et al., 2013; Minella et al., 2015; 24 Klüpfel et al., 2014; Page et al., 2012; Tong et al., 2016). Superoxide dismutase (SOD, 25 SUPEROX-DISMUT-RXN, EC:1.15.1.1) was found to be the predominant RXN related to 26 reactive oxygen species (ROS) detoxification at all time points, with a maximum at day 47 27 (Figure 5F). Catalase peaked at day 4 and superoxide reductases (SOR, 1.15.1.2-RXN) at day 28 47, both exhibiting similar relative abundances at the lowest values (days 0 and 4). 29

The general classic concept of strict anaerobes being unable to cope with O_2 and reactive oxygen species has been long obsolete (Imlay, 2002). While SODs are well distributed amongst aerobic and anaerobic organisms, catalases are more commonly found in aerobes, and SORs in anaerobes (Sheng *et al.*, 2014). This is consistent with the results presented above.

Levels of SOD have been correlated with the aerotolerance of anaerobes (Hassan, 1989; Tally *et al.*, 1977). Gene copy numbers have been correlated with expression levels for numerous proteins. The high relative abundance of SOD during the anaerobic phase could relate to the physiological needs of the anaerobes and aerotolerant microbes thriving in the trenches to deal with transient high oxygen concentrations (Brioukhanov *et al.*, 2002; Sheng *et al.*, 2014).

41 **References**

Brioukhanov AL, Thauer RK, Netrusov AI. (2002). Catalase and superoxide dismutase in the
 cells of strictly anaerobic microorganisms. *Microbiology* 71: 281–285.

- Hassan HM. (1989). Microbial superoxide dismutases. In: Scandalios JG (ed) Vol. 26. *Advances in Genetics*. Academic Press: New York, NY, pp 65–97.
- Imlay JA. (2002). How oxygen damages microbes: Oxygen tolerance and obligate
 anaerobiosis. In: Physiology B-A in M (ed) Advances in Microbial Phisiology Vol. 46.
 Academic Press, pp 111–153.
- Klüpfel L, Piepenbrock A, Kappler A, Sander M. (2014). Humic substances as fully
 regenerable electron acceptors in recurrently anoxic environments. *Nat Geosci* 7: 195–200.
- 51 Minella M, De Laurentiis E, Maurino V, Minero C, Vione D. (2015). Dark production of 52 hydroxyl radicals by aeration of anoxic lake water. *Sci Total Environ* **527–528**: 322–327.
- Page SE, Kling GW, Sander M, Harrold KH, Logan JR, McNeill K, *et al.* (2013). Dark
 formation of hydroxyl radical in arctic soil and surface waters. *Environ Sci Technol* 47:
 12860–12867.
- Page SE, Sander M, Arnold WA, McNeill K. (2012). Hydroxyl radical formation upon oxidation of reduced humic acids by oxygen in the dark. *Environ Sci Technol* 46: 1590–1597.
- Sheng Y, Abreu IA, Cabelli DE, Maroney MJ, Miller A-F, Teixeira M, *et al.* (2014).
 Superoxide Dismutases and Superoxide Reductases. *Chem Rev* 114: 3854–3918.
- Tally FP, Goldin BR, Jacobus NV, Gorbach SL. (1977). Superoxide dismutase in anaerobic
 bacteria of clinical significance. *Infect Immun* 16: 20–25.
- Tong M, Yuan S, Ma S, Jin M, Liu D, Cheng D, *et al.* (2016). Production of abundant
 hydroxyl radicals from oxygenation of subsurface sediments. *Environ Sci Technol* 50: 214–
 221.

66 Supplementary Figures





Figure S1. Daily rainfall at Lucas Heights (ANSTO) Meteorological Station from April to July 2015 (bottom). Corresponding trench water levels across the sampling period, along with the ground surface elevation shown by the dashed line (top). Circles depict dates of chemical and microbial sampling. Rainfall data is courtesy of the Australian Government, Bureau of Meteorology.

RXN-12625







ISOCIT-CLEAV-RXN

Figure S3. ISOCIT-CLEAV-RXN. Marker for the glyoxylate pathway.

COENZYME-F420-HYDROGENASE-RXN



Figure S4. COENZYME-F420-HYDROGENASE-RXN. Indicator of methanogenesis from H₂ and CO₂.



Figure S5. SULFITE-DEHYDROGENASE-RXN (EC:1.8.2.1). Assimilatory sulfate reduction.





86 Figure S6. Relevant RXNs related to the nitrogen cycle.



RXN0-310



Figure S7. PHOSPHOKETOLASE-RXN and RXN0-310. Markers for heterolactic and propionate
 fermentation respectively.



92 Figure S8. Bray-Curtis similarity tree of the functional profiles for the individual sampling 93 replicates.

94 Supplementary Files

- 95 Supplementary file 1. Spreadsheet with YSI data.
- 96 Supplementary file 2. Krona HTML file with the taxonomy.
- 97 Supplementary file 3. Spreadsheet with the relative abundances of all the RXNs.
- 98 Supplementary file 4. Table with the raw HUMAnN2 output.
- All supplementary files can be viewed at: https://dx.doi.org/10.6084/m9.figshare.3817356