File name: Supplementary Information Description: Supplementary Figures and Supplementary Tables.

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Supplementary Figure 1. The effect of H1152 on different hESC/iPSC lines.

<u>Related with Figure 1.</u> (a) Intracellular <u>FCM</u> to detect the expression of insulin in DMSO or H1152 treated H1-derived cells.

(b) Quantification of the percentage of c-peptide⁺ cells of DMSO or H1152 treated cells derived from 2 different iPSC lines. *p* values were calculated by unpaired two-tailed Student's t-test. ***p<0.001. <u>N=3-6 independent biological replicates. Error bar is SEM.</u>



Supplementary Figure 2. H1152 promotes the maturation of hESC-derived glucoseresponding cells. <u>Related with Figure 2</u>. Quantification of the intracellular <u>FCM</u> of H1152treated vs DMSO-treated hESC-derived glucose-responding cells in Fig. 2d. *p* values were calculated by unpaired two-tailed Student's t-test. **p*<0.05, ***p*<0.01. <u>N=3-6 independent</u> <u>biological replicates. Error bar is SEM.</u>



Supplementary Figure 3. H1152 promotes the generation and maturation of hESCderived glucose-responding cells in the presence of a distinct protocol. Related with Figure 2. (a) Intracellular <u>FCM</u> quantification for H1152-treated vs DMSO-treated hESCderived glucose-responding cells in Fig. 4c. <u>N=3-6 independent biological replicates</u> (b) NKX6.1 and GCG staining of purified INS-GFP⁺ cells in H1152- and DMSO- treated population. (c) qRT-PCR analysis of H1152-treated or DMSO-treated cells. Scale bar: 100 µm. The data were normalized to primary human islets. N=6-8 independent biological replicates. *p* values were calculated by unpaired two-tailed Student's t-test. **p*<0.05, ***p*<0.01. <u>Error bar is SEM.</u>



Supplementary Figure 4. Immunohistochemistry analysis of grafts of H1152-treated or DMSO-treated cells. Related with Figure 5. Scale bar: 100 μ m. GCG: glucagon; SS: somatostatin; CP:c-peptide.



Supplementary Figure 5. ROCKII, but not ROCK I, inhibition promotes the generation and maturation of glucose-responding cells. Related with Figure 6. (a) Western blotting analysis of lysates from HUES8-derived cells at 48 hours after DMSO or H1152 (10 μ M) treatment. (b) qRT-PCR to validate the knockdown efficiency in hESCs at 5, 10 or 25 μ g/mL dox concentrations. <u>N=3 independent biological replicates</u>. Error bar is <u>SEM</u>. (c) Western blotting analysis of lysates from HUES8-derived cells at 48 hours after 10 μ g/mL doxycycline treatment.



<u>Supplementary Figure 6. Uncropped western blots of Supplementary Figure 5a (a)</u> and Supplementary Figure 5c (b).

Category	Parameter	Description
Assay	Type of assay	Cell-based
	Target	Pancreatic beta cell generation and
		maturation
	Primary measurement	Percentage of insulin+ cells
	Key reagents	Insulin antibody
	Assay protocol	See the section of experimental
		procedures.
	Additional comments	
Library	Library size	More than 4000 compounds
	Library composition	FDA approved drugs, kinase
		inhibitors, signaling pathway
	a.	regulators
	Source	Sigma LOPAC library, Microsource
	A 111/2 1	Spectrum library
9	Additional comments	204 11 1
Screen	Format	384-well plate
	Concentration(s) tested	10 μ M and 1 μ M compounds
	Plate controls	None Maalti daga
	Reagent/ compound	Multidrop
	Detection instrument and	Molecular Device Image Vpress ^{Micro}
	software	Automated High Content Analysis
	software	System
	Assay validation/OC	System Standard deviation of controls and
	Assay variation/QC	background
	Correction factors	Z score=0.59
	Normalization	None
	Additional comments	
Post-HTS	Hit criteria	The compounds that increase the
analysis		percentage of INS ⁺ cells by three folds
5		were picked as primary hits.
	Hit rate	About 0.2% (8/4000)
	Additional assay(s)	Intracelluar FACS to detect the
	• • •	percentage of insulin+ cells and c-
		peptide+ cells
	Confirmation of hit purity and	Compounds were repurchased from
	structure	Tocris
	Additional comments	

Supplementary Table 1. Related with Figure 1. Primary screening data



Supplementary Table 2. Hit compounds from primary screening.

S1 (3	1 Day	MCDB131	100 ng/Ml activin A
days)	-	1.5g /L Sodium bicarbonate	3 μM Chir99021
-		10 mM glucose	
		0.5% Fatty acid free BSA	
	1 Day	MCDB131	100 ng/mL activin A
		1.5 g/L Sodium bicarbonate	0.3 μM Chir99021
		10 mM glucose	
		0.5% Fatty acid free BSA	
	1 Day	MCDB131	100ng/mL activin A
		1.5g/L Sodium bicarbonate	
		10mM glucose	
		0.5% Fatty acid free BSA	
S2 (2	2 Days	MCDB131	0.25 mM ascorbic acid
days)		1.5 g/L Sodium bicarbonate	50 ng/mL FGF7
		10 mM glucose	
		0.5% Fatty acid free BSA	
S3 (2	2 Days	MCDB131	0.25 mM ascorbic acid
days)		2.5 g/L Sodium bicarbonate	50 ng/mL FGF7
		10 mM glucose	0.25 μM SANT-1
		2% Fatty acid free BSA	1 μM retinoic acid
			100 nM LDN193189
			1:200 ITS-X
			200 nM TPB
S4 (3	3 Days	MCDB131	0.25 mM ascorbic acid
days)		2.5 g/L Sodium bicarbonate	2 ng/mL FGF7
		10 mM glucose	0.25 μM SANT-1
		2% Fatty acid free BSA	0.1 μM retinoic acid
			100 nM LDN193189
			1:200 ITS-X
			200 nM TPB
S5 (3	3 Days	MCDB131	0.25 μM SANT-1
days)		1.5 g/L Sodium bicarbonate	0.05 µM retinoic acid
		20 mM glucose	100 nM LDN193189
		2% Fatty acid free BSA	1:200 ITS-X
			1 μM T3
			10 µM ALK5i
			10 µM Zinc Sulfate
			10 µg/mL heparin
S6-1		MCDB131	100 nM LDN193189
(7		1.5g/L Sodium bicarbonate	1:200 ITS-X
days)		20mM glucose	1 uM T3
5~7		2% Fatty acid free BSA	10 µM ALK5i
			10 µM Zinc Sulfate
	1		pure mille Noricette

Supplementary Table 3. Related with Figure 4. Differentiation Protocol.

		10 μg/mL heparin 100 nM GS inh XX
S6-2	MCDB131	100 nM LDN193189
(7	1.5 g/L Sodium bicarbonate	1:200 ITS-X
days)	20 mM glucose	1 μM T3
	2% Fatty acid free BSA	10 μM ALK5i
		10 μM Zinc Sulfate
		10 μg/mL heparin
		10 μM H1152

Gene	Forward	Reverse
NKX6.1	CCGAGTCCTGCTTCTTCTTG	ATTCGTTGGGGGATGACAGA G
MafA	GAGTTGGCACTTCTCGCTCT	TTCAGCAAGGAGGAGGTCA T
ABCC8	GAGAAGTCGGCCTCTTTGAA	GGGCCTTTGCCATCTATACC
UCN3	GATGGGCTTGGCTTTGTAGA	GGAGGGAAGTCCACTCTCG
G6PC2	ACACTCCAAAGAAATGACCAG G	CGCATCCTGTGTCTGGTATG
INS	GCAGCCTTTGTGAACCAACAC	CCCCGCACACTAGGTAGAG A
KCNK1	GGCCTTACCTCCATCTGACA	TGGAACTGGGACTTCACCTC
KCNK3	GAACATGCAGAACACCTTGC	GGCTCCTTCTACTTCGCCAT
PAX4	GAGGGTCTGGTTTTCCAACA	TGTGCAGAGATGATTCCTGG
SOX9	GTACCCGCACTTGCACAAC	GTGGTCCTTCTTGTGCTGC
NKX2.2	TCTACGACAGCAGCGACAAC	GGAGCTTGAGTCCTGAGGG
NEUROD 1	ATGACCAAATCGTACAGCGAG	GTTCATGGCTTCGAGGTCGT

Supplementary Table 4. Related with Figure 2,4. Primer for qRT-PCR

Supplementary Table 5. Related with Figure 6. shRNA target sequences.

ROCKI	TCCATTCCATGGGTTTTAT
ROCKII	ATCAGACAGCATCCTTTCT
Scramble	GGACTACTCTAGACGTATA