

File Name: Supplementary Information

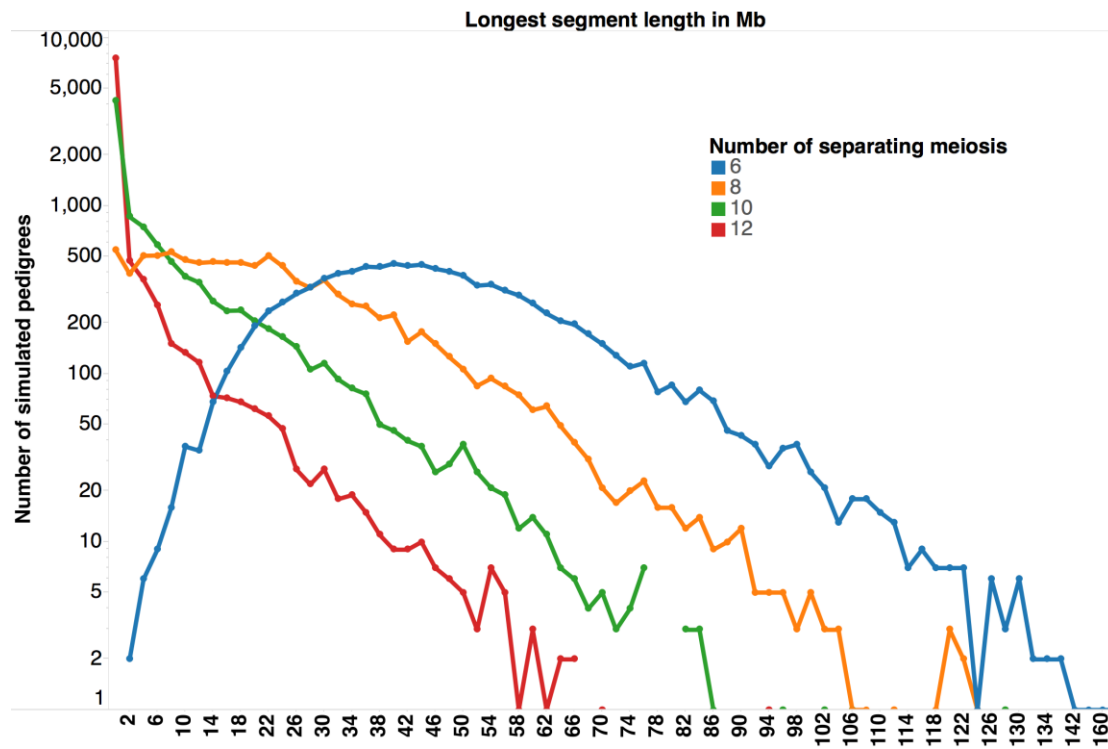
Description: Supplementary Figures, Supplementary Tables and Supplementary Notes.

File Name: Supplementary Data 1

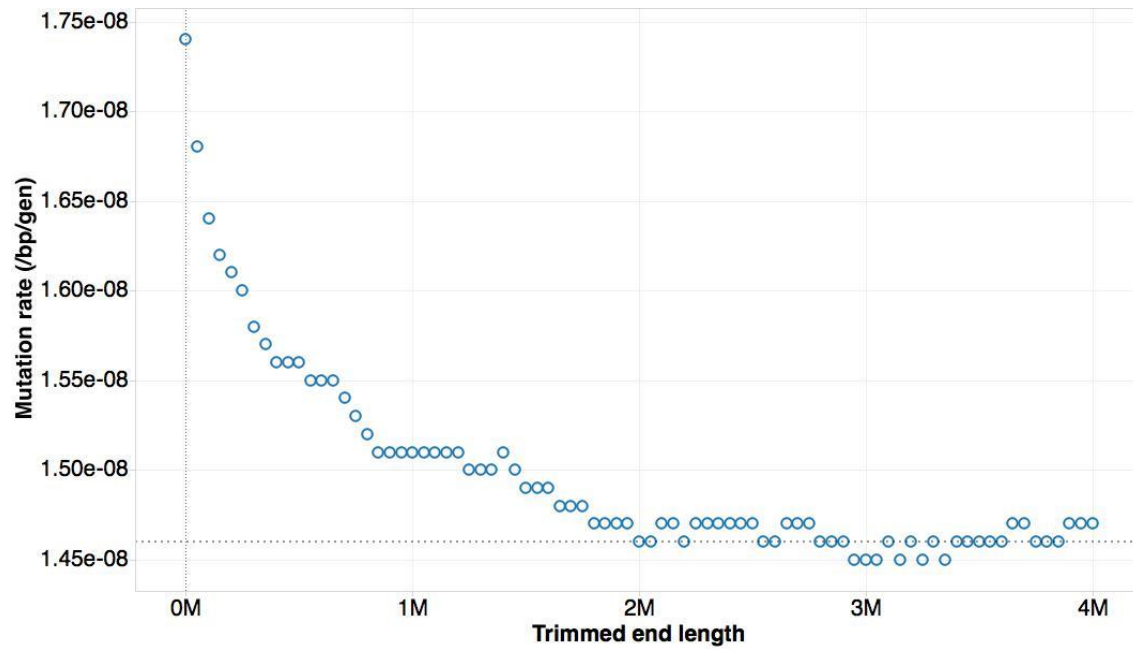
Description: De novo mutations seen in autozygous sequences, PJI Complete Genomics Trios and Scottish Family Health Study, along with their comparisons with the ExAC consortium datasets and partition into mutational spectra.

File Name: Peer Review File

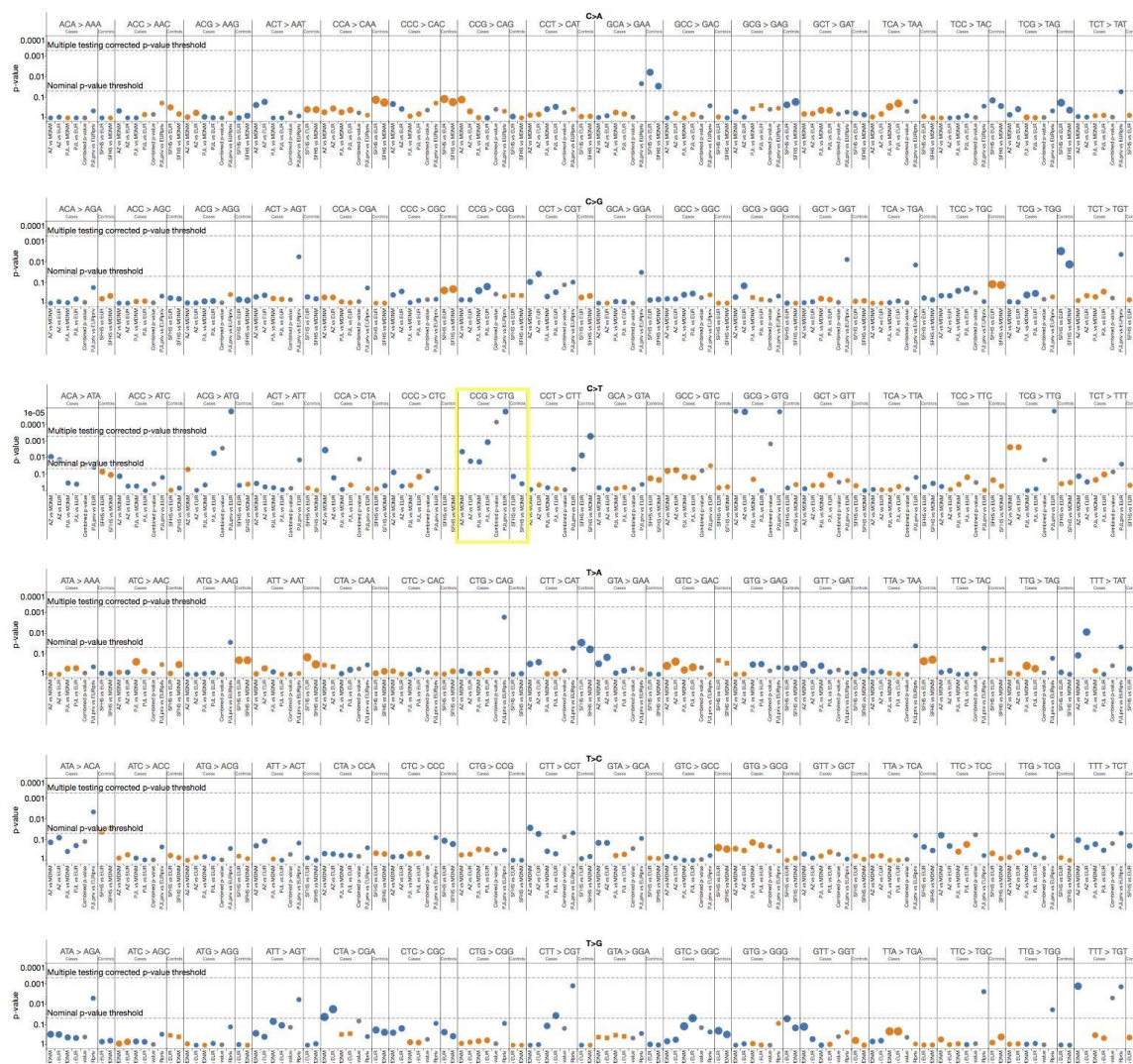
Description:



Supplementary Figure 1. Simulated data of showing histograms of the number of pedigrees for which the longest autozygous segment found is of a certain length. Beyond a separation of 10 meioses to the tMRCA, there are fewer than 8% of pedigrees that have an autozygous segment of at least 10Mb.

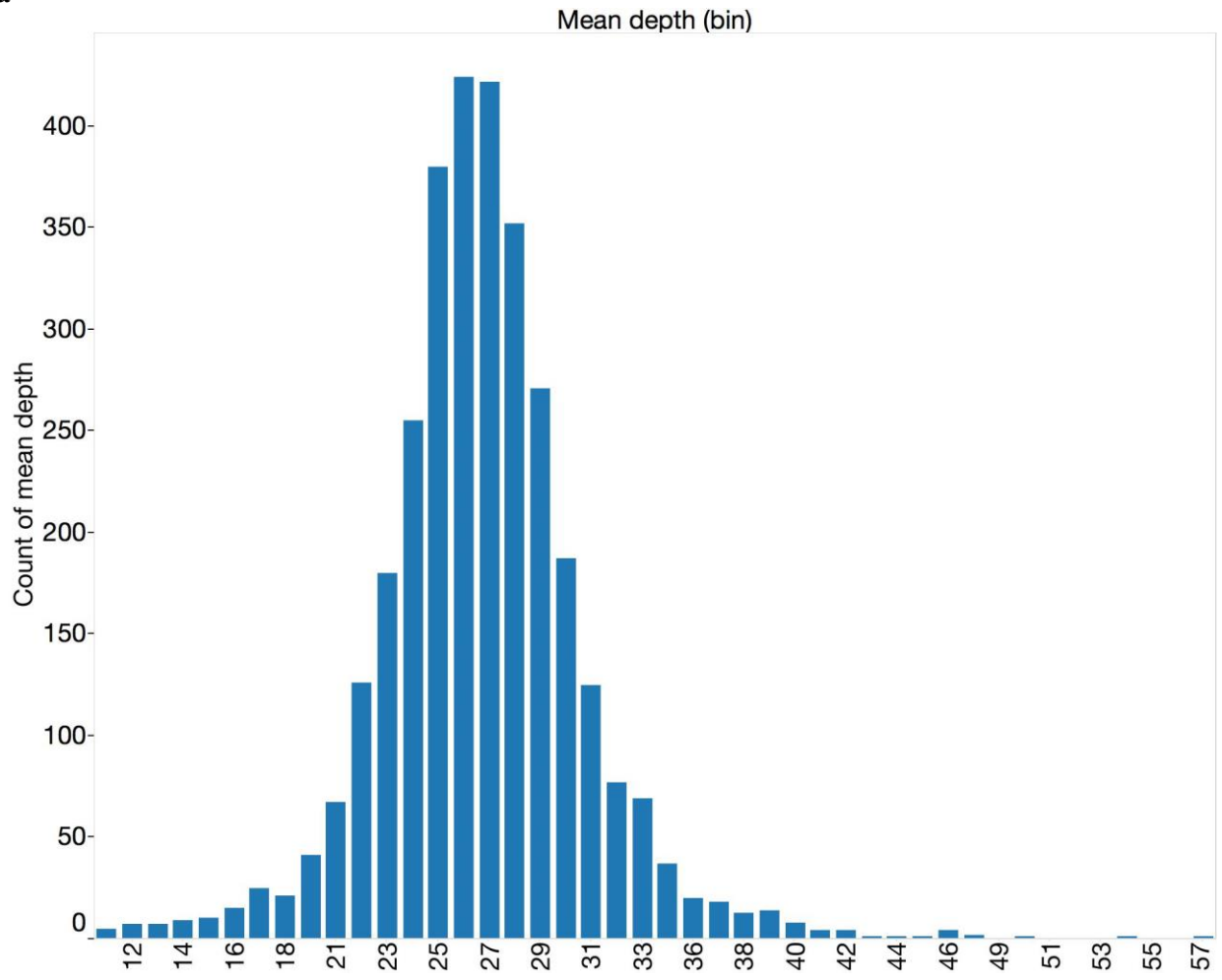


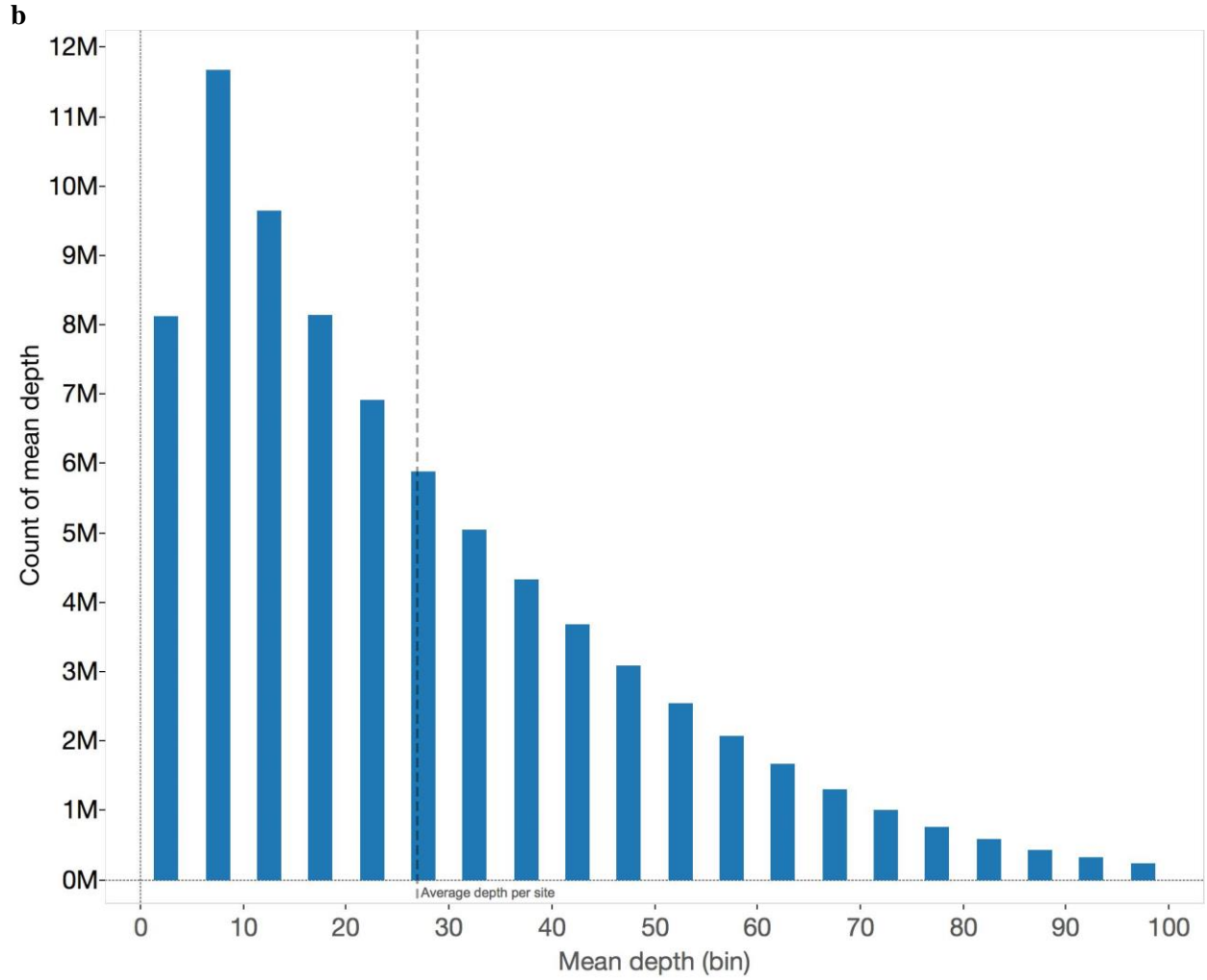
Supplementary Figure 2. The mutation rate estimated from autozygous segments at least 10Mb long that have been further trimmed from each end at a distance given on the x-axis. We see that there is minimal change to the mutation rate estimate beyond 2Mbs of trimming.



Supplementary Figure 3. Comparisons of the proportion of each of the 96 tri-nucleotide signatures across datasets. Differences in context-specific mutation rate. y-axis: significance of the difference in proportion of DNMs for each signature between 1152 mutations from the autozygosity dataset (AZ) and 850 DNMs from the Complete Genomics trio dataset (PJL) in comparison with 6948 mutations from the meta-analysis dataset (MDNM) and mutations private to Europeans in the 1000 Genomes Project (EURpriv). Additional comparisons for mutations private to the PJL population from the 1000 Genomes Project (PJLpriv) and private to Europeans (EURpriv) shown in rightmost panel. As controls significance of the difference in 747 DNMs from the Scottish Family Health Study (SFHS); Colors (Orange, first population has a lower proportion, Blue, otherwise) and size reflect the sign and fold difference of the test. Comparisons for which de novo mutations have 0 counts shown in squares. The only tri-nucleotide context, 5' CCG → CTG 3' that shows experiment wide significance, and consistent direction of effect shown in yellow box.

a





Supplementary Figure 4. Mean depth across sites and individuals on the evaluated portion of the genome

a Histogram of the distribution of mean depth per individual across all sites. **b** Histogram of the mean depth per site across all individuals.

		Self stated parental relatedness					
		First cousin	First cousin once removed	Second cousin	Other blood	Other marriage	Do not know
Inferred Meiosis	6 (First cousin)	835	7	33	29	2	528
	8 (Second cousin)	423	1	47	63	15	621
	10 (Third cousin)	78	1	13	17	11	356
	>10 (Not considered)	19	0	6	14	0	103

Supplementary Table 1. Most probable number of separating meioses giving rise to autozygous segment lengths as compared with those from self-stated parental relatedness.

		Pedigree ascertained relatedness				
		Double First cousin	First cousin	First cousin once removed	Second cousin	Third cousin
Inferred Meiosis	6 (First cousin)	2	46	2	0	0
	8 (Second cousin)	0	2	0	5	0
	10 (Third cousin)	0	0	0	0	1

Supplementary Table 2. Most probable number of separating meioses giving rise to autozygous segment lengths as compared with those from well studied pedigrees.

Allele Frequency	Percentage of sites identified correctly	Percentage of genotypes identified correctly	False negative rate	False positive rate
Singleton	83.77	NA	16.23	3.60
10%	94.55	97.22	8.23	1.05
20%	94.27	98.10	7.63	0.86
30%	93.91	98.17	7.92	0.94

Supplementary Table 3. Estimates of the false negative rates on the allele frequency based on our approach of altering reads to contain a new mutation then remapping them and recalling. Two components to the false negative rate are measured: first the percentage of introduced sites that failed to be called, and second the fraction of introduced heterozygous genotypes that failed to be called at a site that was already known to be polymorphic based on other individuals. The total false negative rate is reflected by aggregating both of these types of error.

Class of mutation	PJL	MDNM
5' CCG 3' → 5' CTG 3'	54	310
not 5' CCG 3' → 5' CTG 3'	795	6592

Supplementary Table 4. 2x2 table showing the number of mutations of the particular class, 5' CCG 3' → 5' CTG 3' in the PJL complete genomics trios and those from a set of meta de novo mutations ascertained in Europeans.

Dataset	Total Number of DNMs	Type of sequencing	Ancestry
Autozygosity, this dataset (AZ)	1152	28x WES illumina HiSeq 2500	Pakistani
Scottish Family Health Study (SFHS) ⁶	747	30x WGS illumina HiSeq 2500	European
Meta de novo mutations ⁶	6902	Variable coverage WGS	European
PJL Complete Genomics Trios ³¹	850	80x Complete genomics	Pakistani
Mutations private to Europeans in the 1000 Genomes Project excluding singletons ¹²	2452300	7.4x WGS illumina HiSeq 2500	European
Mutations private to PJL in the 1000 Genomes Project excluding singletons ¹²	162855	7.4x WGS illumina HiSeq 2500	Pakistani

Supplementary Table 5. Table showing a listing of various datasets their acronyms, the total number of DNMs seen and the sequencing technology used along with their ancestry.

Supplementary Note 1

Equation 1:

$$P(\text{het in dup 2} \mid \text{het in dup 1}, \alpha, \beta, f) = \frac{P(\text{het in dup 2}, \text{het in dup 1} \mid \alpha, \beta, f)}{P(\text{het in dup 1} \mid \alpha, \beta, f)}$$

Equation 2:

$$\begin{aligned} &= \frac{P(\text{het in dup 1}, \text{het in dup 2} \mid \text{reality is hom alt}, \alpha, \beta, f)P(\text{reality is hom alt}) + P(\text{het in dup 1}, \text{het in dup 2} \mid \text{reality is het}, \alpha, \beta, f)P(\text{reality is het}) + P(\text{het in dup 1}, \text{het in dup 2} \mid \text{reality is ref}, \alpha, \beta, f)P(\text{reality is hom ref})}{P(\text{het in dup 1} \mid \text{reality is hom alt}, \alpha, \beta, f)P(\text{reality is hom alt}) + P(\text{het in dup 1} \mid \text{reality is het}, \alpha, \beta, f)P(\text{reality is het}) + P(\text{het in dup 1} \mid \text{reality is ref}, \alpha, \beta, f)P(\text{reality is hom ref})} \\ &= \frac{2f^2(1-\beta)^2 + 2f(1-f)(1-\alpha)^2(1-\beta)^2 + 2f(1-f)\alpha^2\beta^2 + 4f(1-f)(1-\alpha)(1-\beta)(\alpha\beta)}{2f^2(1-\beta) + 2f(1-f)(1-\alpha)(1-\beta) + 2f(1-f)(\alpha\beta) + 2(1-f)^2\alpha} \end{aligned}$$