

1 **Supplementary Information for**

2
3 **Cardiac injury of the newborn mammalian heart**
4 **accelerates**
5 **cardiomyocyte terminal differentiation**
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1 **Supplementary Figure Legends**

2 **Supplementary Figure 1: Effect of AR on nuclear localization of pericentrin. (a, b)**

3 Representative images of cryosections of P3 rat heart ventricles. red/cyan: cardiomyocytes
4 (nuclei: Nkx2.5; cytoplasmic membrane: caveolin 3), green: pericentrin (pericentriolar
5 matrix/centrosome), blue: nuclei (DAPI). Dotted circle: vessel. Arrows: examples of
6 cardiomyocytes with pericentrin localized to the nucleus. Scale bars: a: 25 μ m, b: 10 μ m. (a)
7 Note: 1: Non-myocyte with pericentrin localized to its centrosome (asterisk). 2:
8 Cardiomyocyte with pericentrin localized to its centrosome. 3: Cardiomyocyte with
9 pericentrin localized to the nucleus (arrow). (c) Quantitative analysis of cardiomyocytes
10 proximal (within 1 mm) to base or apex/resection with nuclear pericentrin signal from
11 cryosections of P3 or P6 (SHAM and AR) rat hearts. Data are \pm SD. p-values were calculated
12 using two-tailed Student's t-test.

13

14 **Supplementary Figure 2: Gross morphology of AR mouse hearts.** Representative

15 stereomicroscopic images of SHAM- and AR-operated mouse hearts at different postnatal
16 time points. Arrow heads indicate myocardial scar. Scale bars: 1 mm.

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18 **Supplementary Figure 3: Analysis of H3P-positive cardiomyocytes in AR rat hearts.**

19 Representative immunofluorescence images of sham- and AR-operated P6 rat hearts (scale
20 bars: 1 mm). Red: cardiomyocytes (sarcomeric- α -actinin); green: mitotic cells (H3P); blue:
21 nuclei (DAPI).

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23 **Supplementary Figure 4: Characterization of P1 cardiomyocyte cytokinesis *in vivo*.**

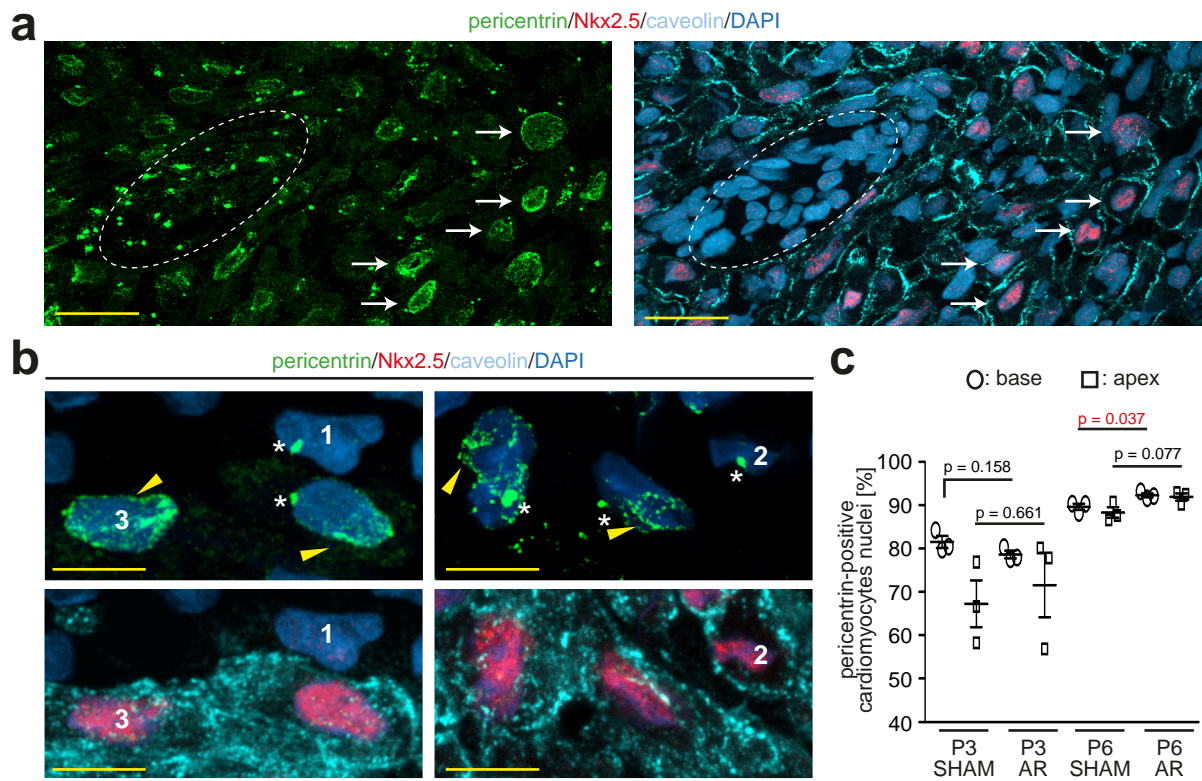
24 Representative images of rat heart sections stained for cytokinesis proteins Aurora B (purple)
25 and Anillin (green) and cardiomyocytes (Troponin I, red). Nuclei were visualized with DAPI
26 (blue). (a) Representative images of P1 SHAM rat cardiomyocytes exhibiting Anillin patterns

1 associated with “normal” cytokinesis. **(b)** Representative images of P1 SHAM rat
2 cardiomyocytes exhibiting Anillin patterns associated with “abnormal” cytokinesis. Scale
3 bars: 10 μm .

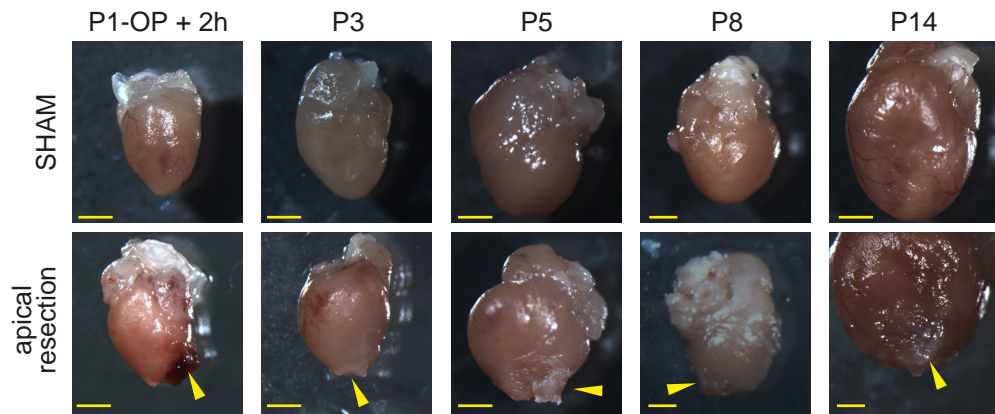
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5 **Supplementary Figure 5: Hypothetical model for the amount of H3P events to support**
6 **proliferation along-side of binucleation after AR.** **(a)** SHAM, binucleation with no
7 proliferation. Experimental data: binucleation at P5 = 20%. The number of H3P events at P3
8 = 1-fold. **(b)** AR, binucleation with no proliferation. Experimental data: binucleation at P5 =
9 30%. The number of H3P events at P3 = 2-fold. **(c, d)** Assumption: all **(c)** or the additional **(d)**
10 H3P events after AR indicate proliferation. This would result in a decrease in the binucleation
11 rate which has not been observed. **(e)** Any significant increase in proliferation induced by AR
12 requires a significant increase in binucleation and thus a higher fold-increase in H3P-positive
13 cardiomyocytes than observed. P: postnatal day. Dashed arrows: no cell cycle activity from
14 P3 to P5. Green arrows: mitosis (i.e. H3P event) from P3 to P5. Note, it is known that H3P
15 events post-P3 are strictly followed by binucleation during normal (i.e. SHAM) neonatal heart
16 development.

Zebrowski DC *et al.*, Figure S1



Zebrowski DC *et al.*, Figure S2

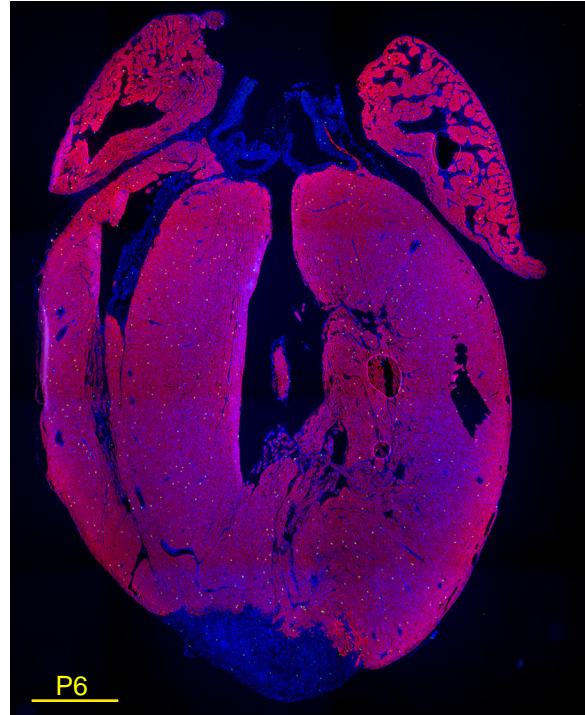
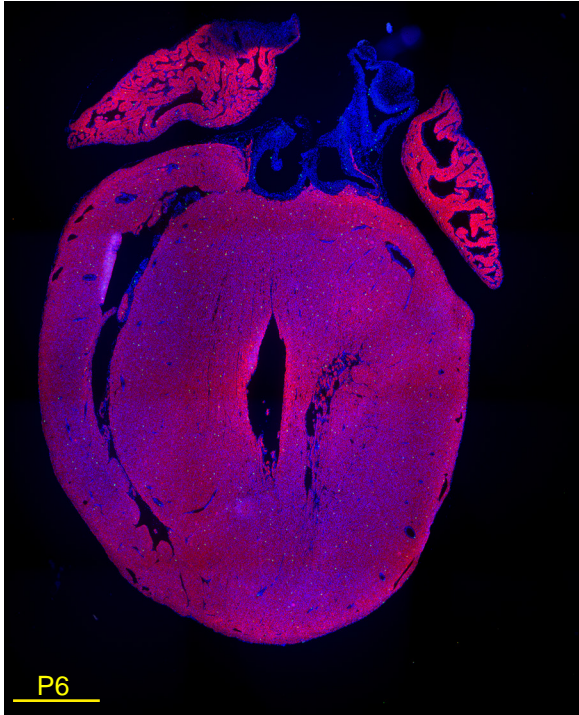


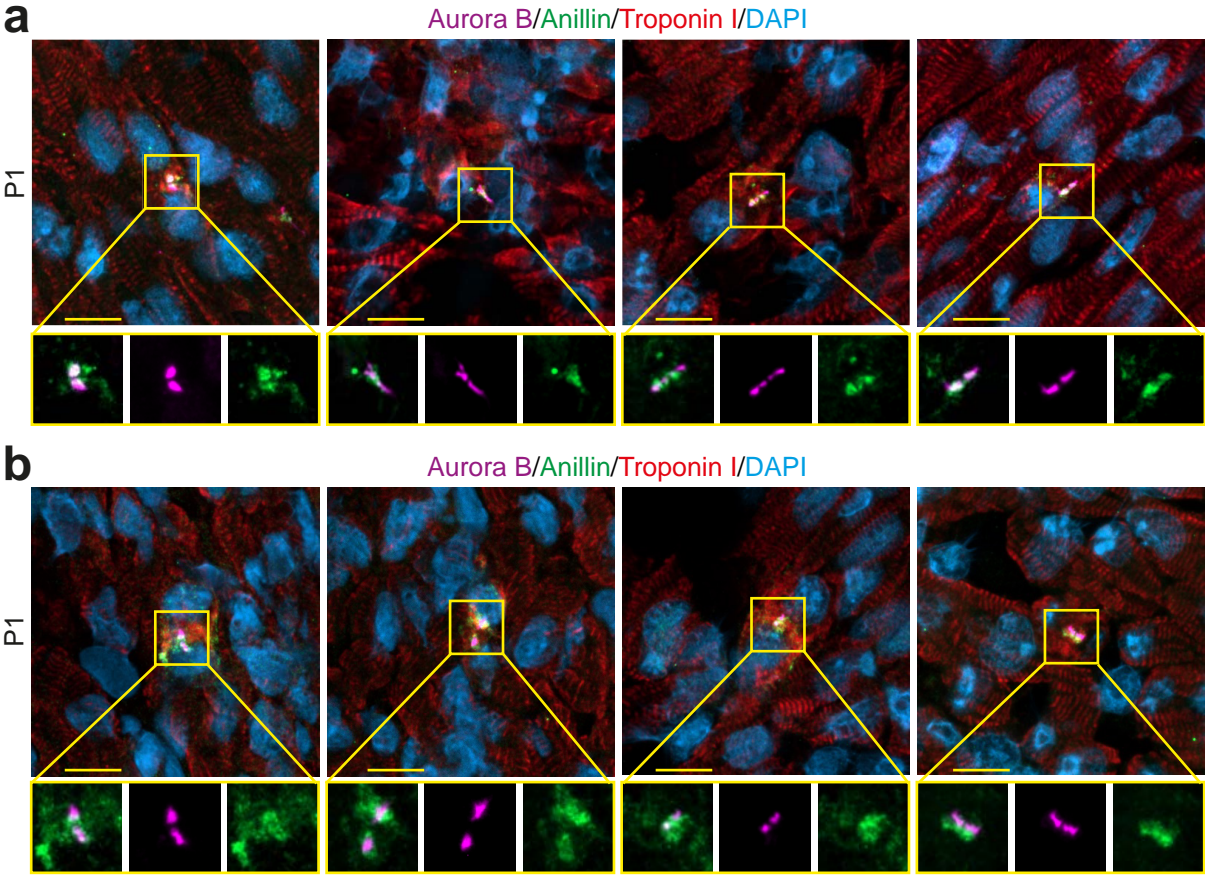
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H3P/Actinin/DAPI

SHAM

apical resection





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